

Figures

Fig. 1. Effects of PSK- α on non-embryogenic cell proliferation in the presence of different concentrations of 2,4-D. Non-embryogenic cells were incubated in liquid medium containing PSK- α at the indicated concentrations with various concentrations of 2,4-D (A, 4.5×10^{-6} M; B, 4.5×10^{-7} M). The cell density was adjusted to 0.2 ml PCV/l and cells were counted on the 14th day of culture. Data are the means of three replicates \pm SD (the bar is not shown where it was too small to be displayed).

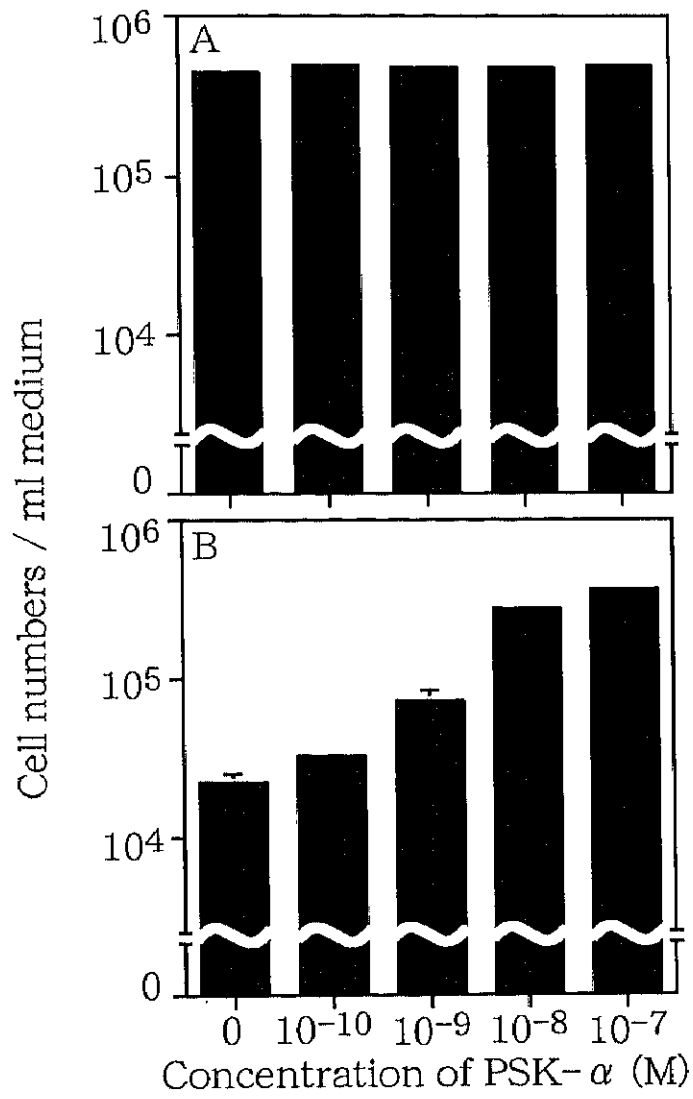


Fig. 2. Effects of PSK- α on non-embryogenic cell proliferation in the presence of different concentrations of IAA (A) and NAA (B). Non-embryogenic cells were incubated in liquid medium containing PSK- α at the indicated concentrations with various concentrations of IAA (a, 4.5×10^{-6} M; b, 4.5×10^{-7} M; c, 4.5×10^{-8} M) and NAA (a, 4.5×10^{-6} M; b, 4.5×10^{-7} M; c, 4.5×10^{-8} M). The cell density was adjusted to 0.2 ml PCV/l, and cells were counted on the 14th day of culture. Data are the means of three replicates \pm SD (the bar is not shown where it was too small to be displayed).

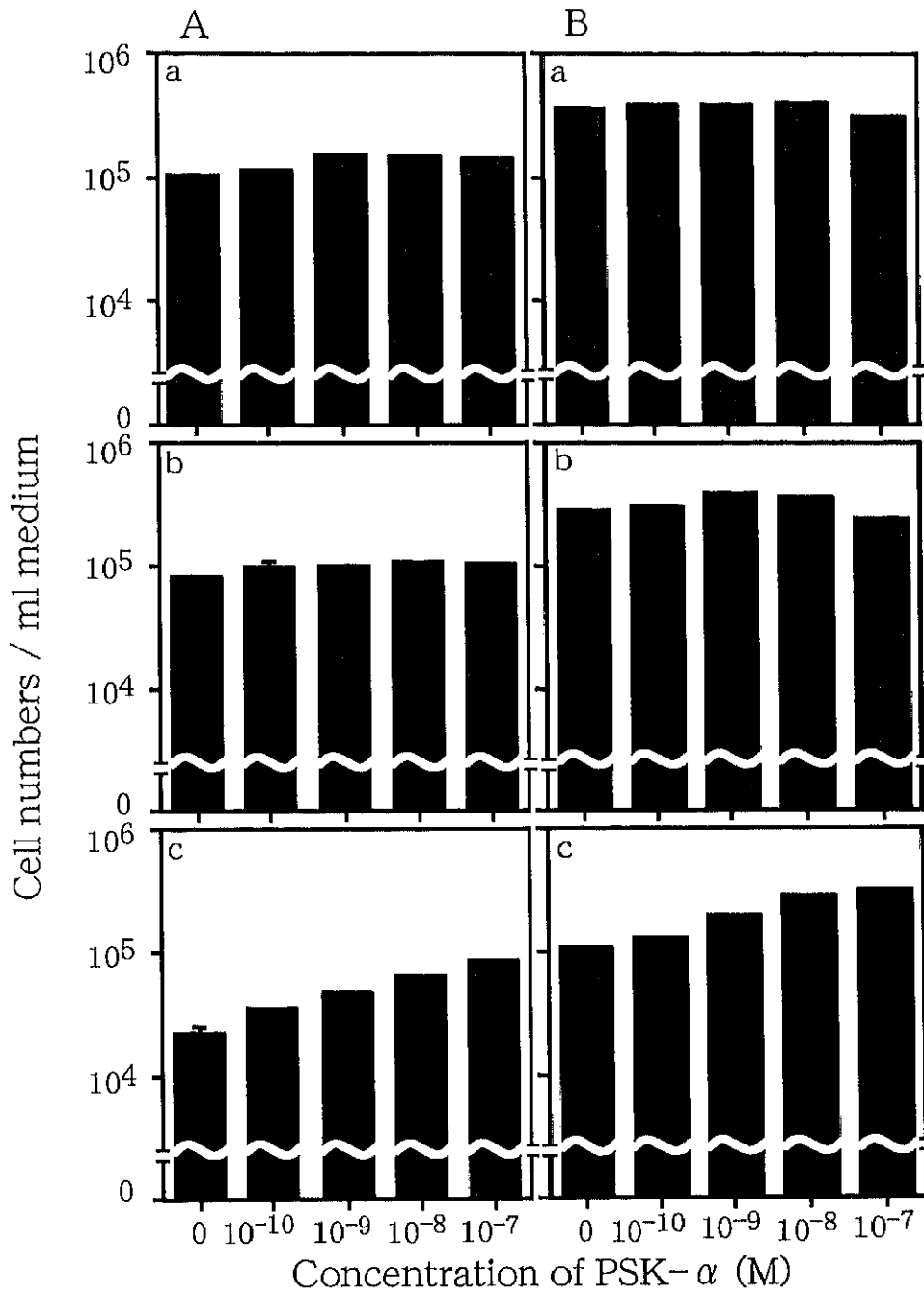


Fig. 3. PSK- α accumulation in CM derived from non-embryogenic cell culture in the presence of high (H) or low (L) concentrations of auxin (2,4-D, IAA, and NAA). Non-embryogenic cells were suspended at 0.2 ml PCV/l and cultured in liquid medium that contained 4.5×10^{-6} M (H) or 4.5×10^{-7} M (L) of auxin. The PSK- α concentration in the medium was determined by competitive ELISA. Data are the means of duplicates \pm SD (the bar is not shown where it was too small to be displayed).

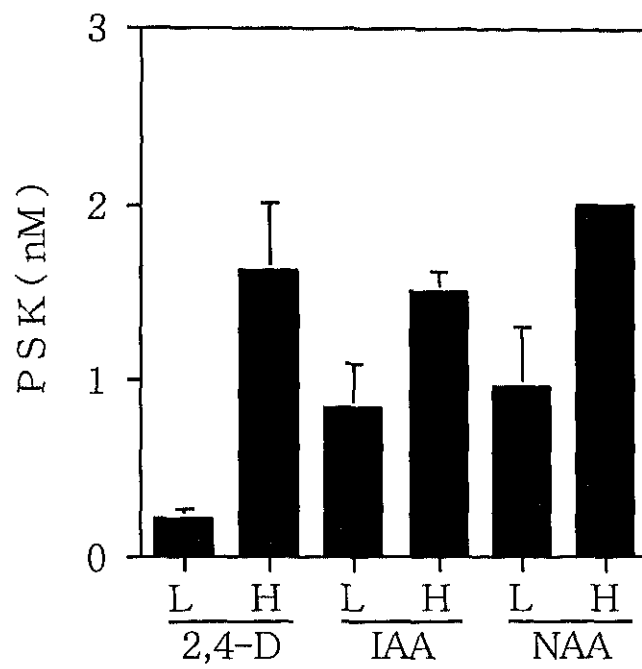


Fig. 4. Effects of PSK- α on non-embryogenic cell proliferation in the absence of 2,4-D. Non-embryogenic cells were precultured in medium without 2,4-D for 14 days and then incubated in liquid medium containing PSK- α at the indicated concentrations without 2,4-D. The cell density was adjusted to 0.2 ml PCV/l, and cells were counted on the 14th day of culture. Data are the means of three replicates \pm SD.

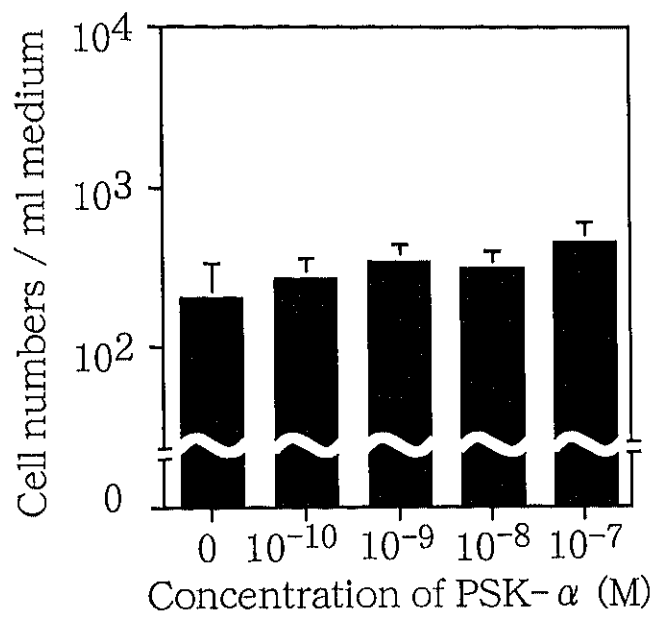


Fig. 5. Flow cytometry analysis of non-embryogenic cells after release from propyzamide blocking in M phase. A time course of the accumulation of cells in G1- or G2/M- phases of the cell cycle was examined in cell suspensions cultured in the presence of a low concentration (4.5×10^{-7} M) of 2,4-D without (A and D) or with (B and E) PSK- α (1×10^{-7} M), and in the presence of a high concentration (4.5×10^{-6} M) of 2,4-D without PSK- α (C and F). Closed and open symbols indicate the percentage of the cells accumulated in the G1 phase or G2/M phases, respectively. Each experiment was performed in duplicate, and the results of each experiment are shown separately (1st and 2nd experiment).

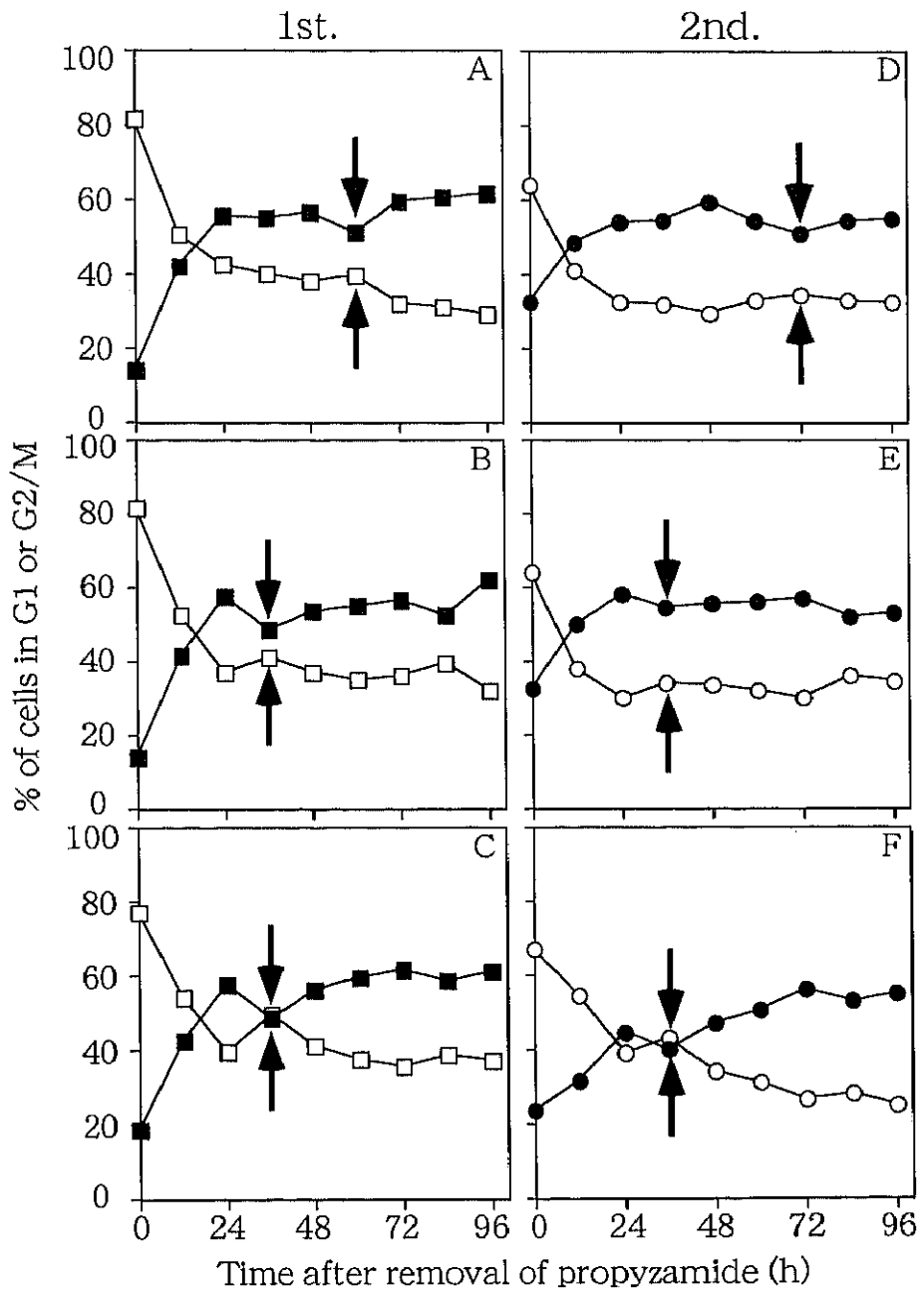


Fig. 6. Effects of PSK- α on cell division in non-embryogenic cells after release of propyzamide blocking. Cell numbers were counted each day for four days. The initial cell density was adjusted to 0.2 ml PCV/l. Each experiment includes three replicates, and all experiments were performed in duplicate. Data are the means with standard deviation (where the bar is not shown, it was too small to be displayed). ■, low-concentration (4.5×10^{-7} M) of 2,4-D without PSK- α ; ●, low concentration (4.5×10^{-7} M) of 2,4-D with PSK- α (1×10^{-7} M); ▲, high concentration (4.5×10^{-6} M) of 2,4-D without PSK- α .

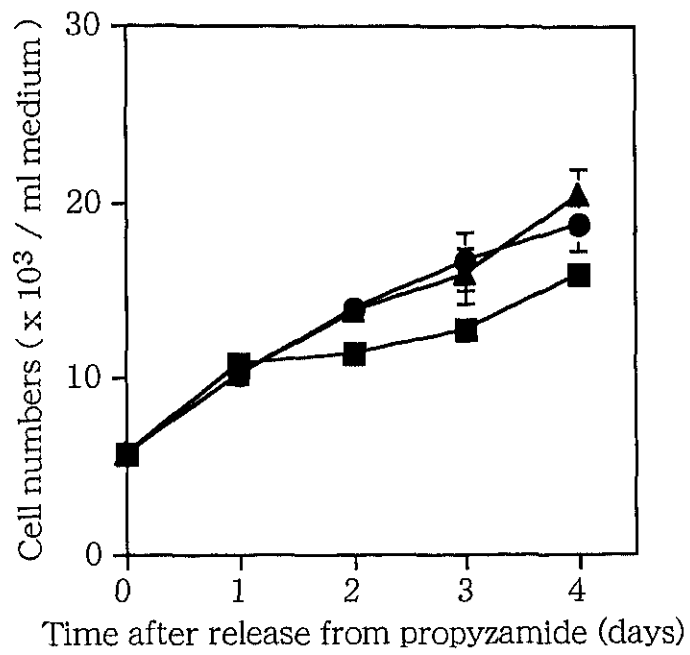
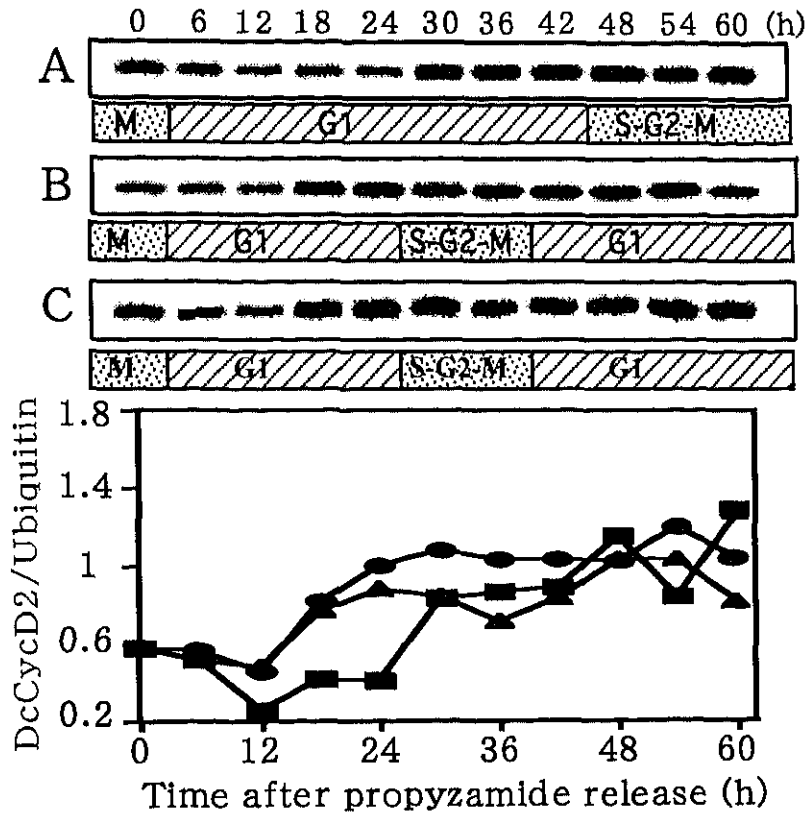


Fig. 7. Nucleotide sequence of the carrot *CycD2* cDNA and deduced amino acid sequence. The initiation codon of the open reading frame is indicated by a double underline, and the LxCxE motif is shown in boldface letters. The cyclin box is shown in black, and a putative poly A signal sequence is underlined.

1 ACTCACTATAGGGCTCGAGCGGCCGCCCGGGCAGGTGGAGACACATATAAAATTACATAT 60
61 GCTCACACAACACAGCAACACACATTTATAAATGTATATATAAAGCTCACTTCTTTGCTA 120
121 CAGTCTATCTATCCCAAACCTTCATTTCACTACTTTACAGCCATGCTTCTTGATTTCTTTA 180
181 TATCAAGTTCTCTAATTTCTTATTTCATTTTAGATGAGATTCACTAGGGTTTACACAGAGG 240
M R F T R V Y T E V
241 TTTTCAATATGACAGACCATAGCCTCCTCTGCACAGAAACCAATAAATTGTGTTTTGATG 300
F N M T D H S L L C T E T N N L C F D D
301 ATCTTGAGGCTAGAGATGATCAGGACCCGAGAATAGATTGTGAAAATGTGGTGGGTAATG 360
L E A R D D Q D P R I D C E N V V G N E
361 AATCAGAAGCTTTGATTTGTGCCGTCCCATTACAGAGAGATGAAGATTTTGTGTTTGTGT 420
S E A L I C A V P L Q R D E D F V F V F
421 TTGAAGCAAGGTGAATTTTTGCCAGAGGTGATTTTCTCCACAGATAGAAGTGGTGAGC 480
E S K V N F C P E V I F S T D R S G E L
481 TTGATTTGTGTGTCAGAAAAGAGGCCCTTGATTGGATTTATAAGGCTCATGCTCATTACA 540
D L C V R K E A L D W I Y K A H A H Y N
541 ATTTTGGCAGCATTGACTGTTTGGCTTAGCAGTGAATTATTTGGATCGCTTTCTTTCTGTT 600
F G A L S V C L A V N Y L D R F L S L Y
601 ATCAATTCGCCACTGCAAAAAGACTGCACTGTCATTTGTTAGCTGTTGGCTGTTTATCAC 660
E L P S G K K W T V Q L L A V A C L S L
661 TGGCAGCAAAAATGGAGCAGCTTAATGTCGCACTAAGTGTGATTTACAGGTGGCTGATC 720
A A K M E E V N V P L T V D L Q V A D P
721 CTAAGTTGCTGTTGGAGCGCAAAAAGCATTAAAAGAATGGAGCTTTTGGTGTGAGCACCT 780
K F V F E A K T I K R M E L L V L S T L
781 TGAAATGGAGATGCAAGCCTGCAAGCCTTGTTCATTCATAGATTACTTCCTTCGAAAAA 840
K W R M Q A C T P C S F I D Y F L R K I
841 TCAACAATGCTGATGCGCTTCCATCGGGGTCTCTGATCGATAGGTGCGATTCAAGTTCATTT 900
N N A D A L P S G S L I D R S I Q F I L
901 TGAAAACGATGAAAGGTATTGATTTTCTGGAATTCAGGCCCTCAGAAAATTCAGCAGCTG 960
K T M K G I D F L E F R P S E I S A A V
961 TGGCAATTTGTGTAACAAGAGAAGCACAAACACTAGACATTAATAAGGCAATGTCTAATA 1020
A I C V T R E A Q T L D I N K A M S N I
1021 TCATACCTGTTGAAAAGGATAGAGTATTCAAGTGTATTGAAATGATTCAAGATCTGACAT 1080
I P V E K D R V F K C I E M I Q D L T L
1081 TGGTTACTGAGACTAGTAATGTAGCTAGTGGTAGAACCAAGAGCACAAGTGCCACAAAGTC 1140
V T E T S N V A S G R T R A Q V P Q S P
1141 CTGTTGGGGTGTGATGCTGCATGCTTGAGCTATAAGAGTGATGAGAGAACAGTTGGGT 1200
V G V L D A A C L S Y K S D E R T V G S
1201 CATGTCCTAATTCTTCTTTACATACTGAGACTAGTCCACACACTaAAAGGAGGAAGCTGA 1260
C P N S S L H T E T S P H T K R R K L I
1261 TTGAGATCATGAAATGTGGATTTTACTCTATCACTCAGTTTCAGATTTTTAGTGAATGGG 1320
E I M K C G F Y S I T Q F Q I F S E W E
1321 AGTTTTGGTGTGGCTGCCTTTGCAAGCGACCaCCTTTATGATAGAGGAAAAATAATATAT 1380
F W C G C L C K R P P L * *
1381 AAATATAAAAAATATGTAGAGAGAgAGAgAGGGGAgAgAGAATcTTGTGCCTTGAGCATGA 1440
1441 ATATAGTTGTGACCAGCTGTTAATAAAAATGTCATTTTAAACAGTAAAACCTTAGTGTGGGGG 1500
1501 AAGTGGCCGGCCAGGAGTAaAgAGGTTTTTGCTATTTCAAGTTGTTTTATTAATTATTATT 1560
1561 ACTACATACAGAATATATTTCTCAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA 1620
1621 CAAAAAAAAAAAAAAAAA 1636

Fig. 8. RT-PCR for the *DcCycD2* and ubiquitin mRNA in non-embryogenic cells after release from propyzamide blocking. Culture conditions after propyzamide release were as follows: (A, ■), low concentration (4.5×10^{-7} M) of 2,4-D without PSK- α ; (B, ●), low concentration (4.5×10^{-7} M) of 2,4-D with PSK- α (1×10^{-7} M); (C, ▲), high concentration (4.5×10^{-6} M) of 2,4-D without PSK- α . Hybridization signals were quantified using a Fuji BAS5000 Imaging Analyzer. Quantified data were equalized by normalizing to the amount of ubiquitin signal. Each experiment was performed in duplicate and the figure shows representative experiments. Rectangular bars below each expression data show the cell cycle expected from FCM analysis of Fig. 5.

DcCycD2



Ubiquitin

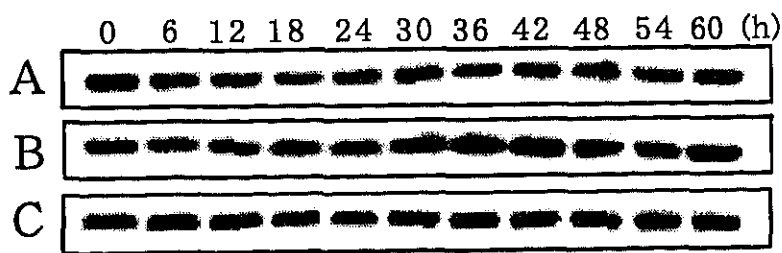


Fig. 9. RT-PCR for the *DcH4* mRNA in non-embryogenic cells after release from propyzamide blocking. Culture conditions after propyzamide release were as follows: (A, ■), low concentration (4.5×10^{-7} M) of 2,4-D without PSK- α ; (B, ●), low concentration (4.5×10^{-7} M) of 2,4-D with PSK- α (1×10^{-7} M); (C, ▲), high concentration (4.5×10^{-6} M) of 2,4-D without PSK- α . Hybridization signals were quantified using a Fuji BAS5000 Imaging Analyzer. Quantified data were equalized by normalizing to the amount of ubiquitin signal. Each experiment was performed in duplicate and the figure shows representative experiments. Rectangular bars below each expression data show the cell cycle expected from FCM analysis of Fig. 5.

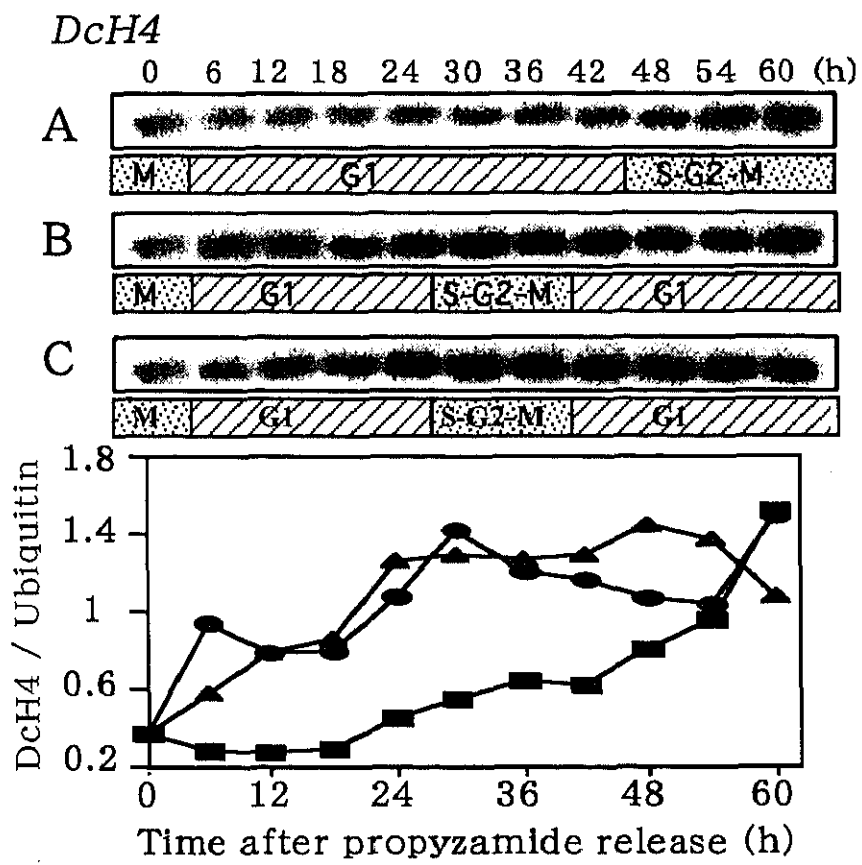


Fig. 10. RT-PCR for the *DcCycB1;1* mRNA in non-embryogenic cells after release from propyzamide blocking. Culture conditions after propyzamide release were as follows: (A, ■), low concentration (4.5×10^{-7} M) of 2,4-D without PSK- α ; (B, ●), low concentration (4.5×10^{-7} M) of 2,4-D with PSK- α (1×10^{-7} M); (C, ▲), high concentration (4.5×10^{-6} M) of 2,4-D without PSK- α . Hybridization signals were quantified using a Fuji BAS5000 Imaging Analyzer. Quantified data were equalized by normalizing to the amount of ubiquitin signal. Each experiment was performed in duplicate and the figure shows representative experiments. Rectangular bars below each expression data show the cell cycle expected from FCM analysis of Fig. 5.

DcCycB1;1

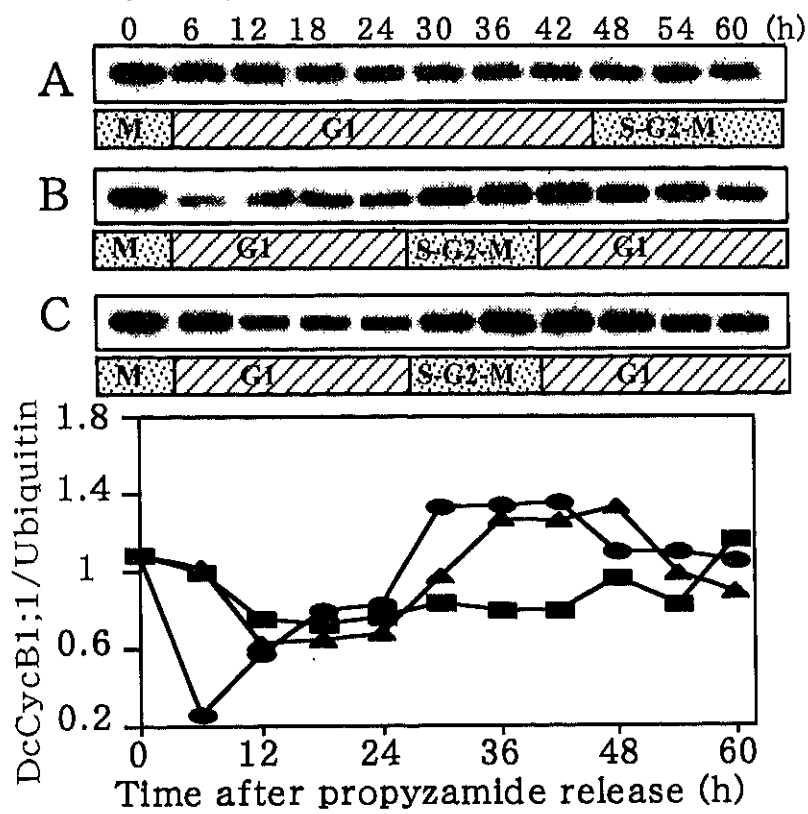


Fig. 11. Flow cytometry analysis of non-embryogenic cells after addition of propyzamide with aphidicolin blocking release. A time course of the accumulation of cells in the G2/M phase of the cell cycle was examined in a cell suspension cultured in the presence of a low concentration (4.5×10^{-7} M) of 2,4-D without (■ and □) or with (● and ○) PSK- α (1×10^{-7} M), and in the presence of a high concentration (4.5×10^{-6} M) of 2,4-D without PSK- α (▲ and △). Each experiment was performed in duplicate, and the results of each experiment are shown separately (1st and 2nd experiment).

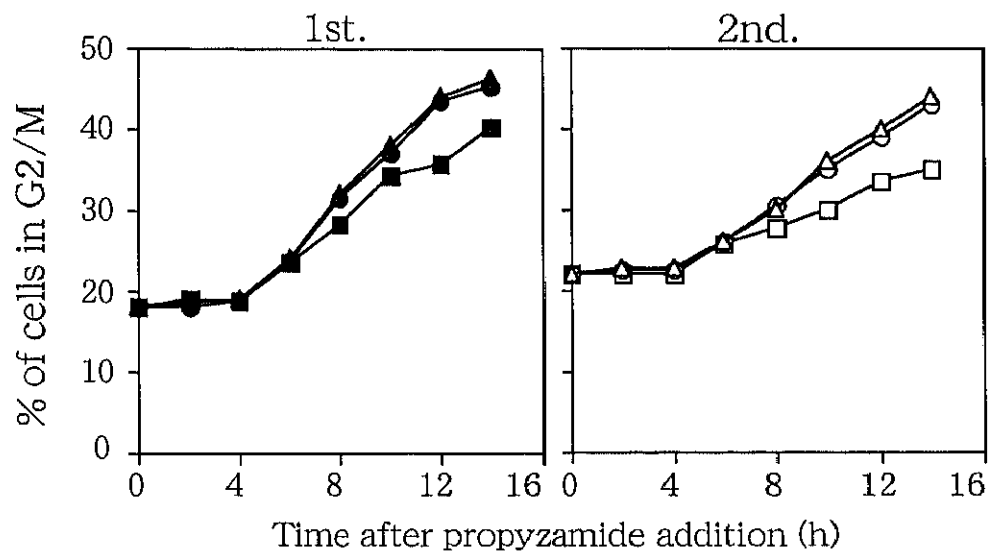


Fig. 12. The nuclear DNA C-value of quiescent non-embryogenic cells. Non-embryogenic cells cultured in phytohormone-free MS medium for seven days were used in FCM analysis.

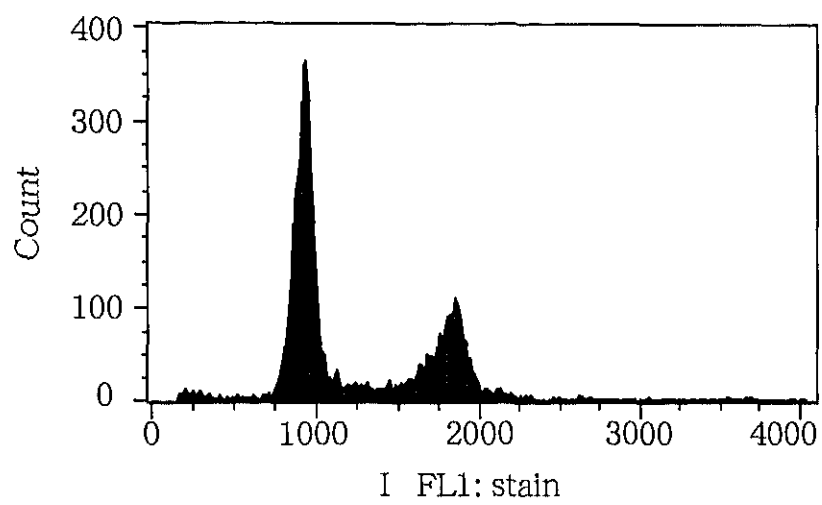
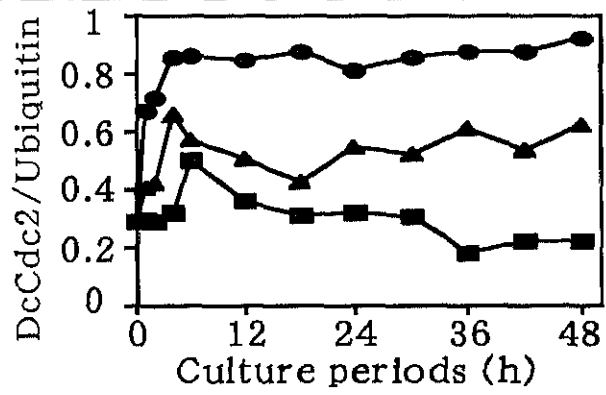
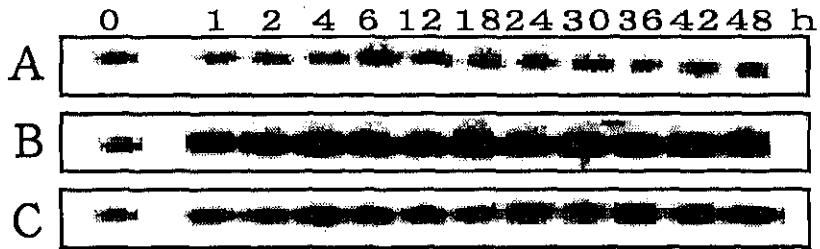


Fig. 13. RT-PCR for *DcCdc2* and ubiquitin mRNA in non-embryogenic cells after re-entry into the cell cycle from the quiescent state. Quiescent non-embryogenic cells were cultured under the following conditions: (A, ■), low concentration (4.5×10^{-7} M) of 2,4-D without PSK- α ; (B, ●), low concentration (4.5×10^{-7} M) of 2,4-D with PSK- α (1×10^{-7} M); (C, ▲), high concentration (4.5×10^{-6} M) of 2,4-D without PSK- α . Hybridization signals were quantified using a Fuji BAS5000 Imaging Analyzer. The quantified data were equalized by normalizing to the amount of ubiquitin signal. Each experiment was performed in duplicate and the figure shows representative experiments.

DcCdc2



Ubiquitin

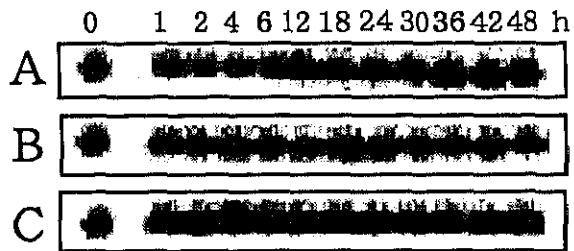


Fig. 14. RT-PCR for *DcCycD2* mRNA in non-embryogenic cells after re-entry into the cell cycle from the quiescent state. Quiescent non-embryogenic cells were cultured under the following conditions: (A, ■), low concentration (4.5×10^{-7} M) of 2,4-D without PSK- α ; (B, ●), low concentration (4.5×10^{-7} M) of 2,4-D with PSK- α (1×10^{-7} M); (C, ▲), high concentration (4.5×10^{-6} M) of 2,4-D without PSK- α . Hybridization signals were quantified using a Fuji BAS5000 Imaging Analyzer. The quantified data were equalized by normalizing to the amount of ubiquitin signal. Each experiment was performed in duplicate and the figure shows representative experiments.

DcCycD2

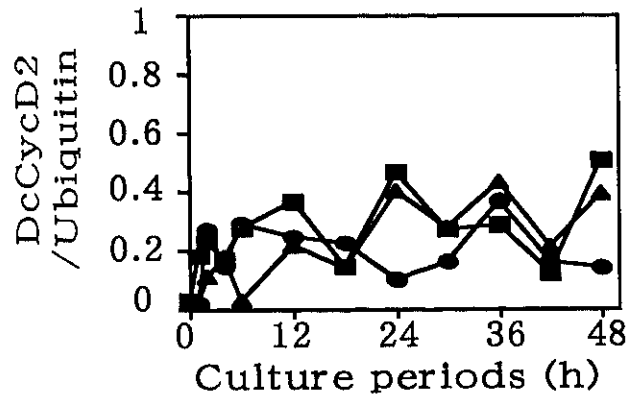
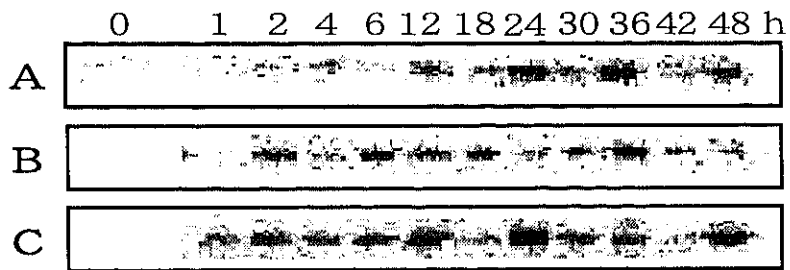


Fig. 15. RT-PCR for *DcH4* mRNA in non-embryogenic cells after re-entry into the cell cycle from the quiescent state. Quiescent non-embryogenic cells were cultured under the following conditions: (A, ■), low concentration (4.5×10^{-7} M) of 2,4-D without PSK- α ; (B, ●), low concentration (4.5×10^{-7} M) of 2,4-D with PSK- α (1×10^{-7} M); (C, ▲), high concentration (4.5×10^{-6} M) of 2,4-D without PSK- α . Hybridization signals were quantified using a Fuji BAS5000 Imaging Analyzer. The quantified data were equalized by normalizing to the amount of ubiquitin signal. Each experiment was performed in duplicate and the figure shows representative experiments.

DcH4

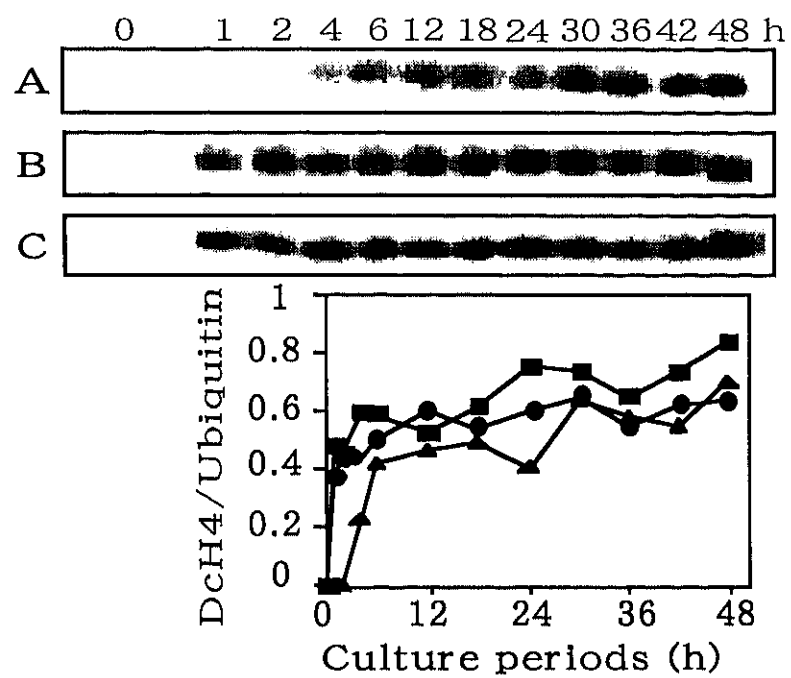


Fig. 16. RT-PCR for *DcCycB1;1* in non-embryogenic cells after re-entry into the cell cycle from the quiescent state. Quiescent non-embryogenic cells were cultured under the following conditions: (A, ■), low concentration (4.5×10^{-7} M) of 2,4-D without PSK- α ; (B, ●), low concentration (4.5×10^{-7} M) of 2,4-D with PSK- α (1×10^{-7} M); (C, ▲), high concentration (4.5×10^{-6} M) of 2,4-D without PSK- α . Hybridization signals were quantified using a Fuji BAS5000 Imaging Analyzer. The quantified data were equalized by normalizing to the amount of ubiquitin signal. Each experiment was performed in duplicate and the figure shows representative experiments.

DcCycB1;1

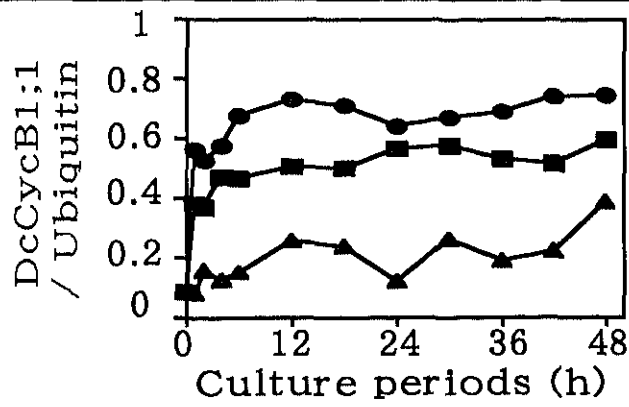
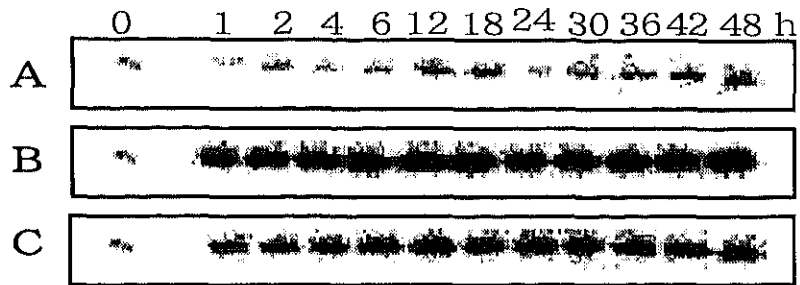


Fig. 17. A schematic model for the promotion of cell proliferation by PSK- α . Auxin involves in the production of PSK- α and the PSK- α stimulates the cell proliferation by activating the cell cycle induced by auxin.

