

1 **Stability and performance of algal-bacterial granular sludge in shaking photo-**
2 **sequencing batch reactors with special focus on phosphorus accumulation**

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14 **Abstract**

15 The granular stability, nutrients removal and phosphorus (P) accumulation of algal-bacterial
16 aerobic granular sludge(AGS) was examined by using shaking photoreactors (at a fixed light/dark
17 cycle of 12h/12h). During the 25 days' operation, algal-bacterial AGS possessed good granular
18 integrity ($8.4\pm 0.6\%$), and excellent removals of dissolved organic carbon ($94.8\pm 1.6\%$) and total
19 nitrogen ($71.1\pm 3.3\%$). More extracellular proteins (153.7 ± 2.3 mg/g) were excreted from the
20 granules with a high proteins/polysaccharides ratio(~ 7.4) on day 25, especially the tightly bound
21 proteins mainly responsible for granular stability. Decrease in P content, especially non-apatite
22 inorganic P relating to Fe-PO₄ precipitates, was detected in the granules to some extent, although
23 $54.8\pm 17.1\%$ of total P removal was achieved during the light-on cycles. Still, high P bioavailability
24 (92.0%) was kept in the algal-bacterial AGS throughout the test period. Further optimization of
25 light-on/light-off cycle and hydraulic/sludge retention time is demanding for better and stable P
26 accumulation in the algal-bacterial granules with high bioavailability.

27 **Keywords:** Algal-bacterial aerobic granular sludge; Shaking photoreactor; Granular stability;
28 Nutrients removal; Phosphorus accumulation

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30 **1. Introduction**

31 Algal-bacterial consortia is regarded as a competitive alternative to conventional activated
32 sludge (AS) process due to its unique features such as less energy demand for degradation of
33 organic matter, enhanced nutrients removal, and higher potential for resources recovery (Quijano et
34 al., 2017; Sforza et al., 2018). In a conventional wastewater treatment plant (WWTP) or AS
35 process-based WWTP, up to 60% of the energy consumption is contributed by aeration units which
36 provide oxygen (O₂) necessary for the growth of aerobic bacteria; while in the algal-bacterial
37 consortia, algae can produce sufficient O₂ that is required by aerobic bacteria to efficiently remove

38 nutrients from wastewater (Boelee et al., 2014). Thus, wastewater treatment systems using algal-
39 bacterial consortia attract much attention.

40 On the other hand, algae harvesting and separation from the treated water is challenging the
41 promising algal-bacterial consortia systems due to the small size of microalgae cells (3-30 μm),
42 poor settleability and low density (Zhou et al., 2017; Hu et al., 2017). Recently, there's a growing
43 interest in the use of innovative solutions that combine microalgae technology with aerobic granular
44 sludge (AGS) process, aiming to solve this problem. In comparison to the AS process-based
45 wastewater treatment, algal-bacterial granular consortia with a compact granular structure and good
46 biomass settleability reflects a prospective option for more nutrients uptake or removal from
47 wastewater (Ahmad et al., 2017; Liu et al., 2018; Zhang et al., 2018). In addition, the application of
48 algal-bacterial AGS for potentially less energy consumption during wastewater treatment, especially
49 in aeration units, has become a research hotspot. Tiron et al. (2015) claimed that the activated algae
50 granular system could efficiently remove organic matter and nutrients from low-strength wastewater
51 with O_2 being provided only by algae through photosynthesis. Most recently, Abouhend et al.
52 (2018) attempted to treat wastewater with oxygenic photogranules in stirred-tank reactors under no
53 aeration condition, achieving efficient removals of chemical oxygen demand (COD) and nitrogen
54 (N). Although much efforts have been devoted to the algal-bacterial granular system, especially on
55 energy saving via less aeration or no aeration, previous works mainly focused on organics or
56 nutrients removal efficiencies under the test conditions.

57 As it is known, phosphorus (P) recovery from P-rich resources is very important due to the
58 limited global reserves of high-quality phosphate rock, especially in Japan. In recent years, many
59 researches concentrated on P recovery from sewage sludge produced from WWTPs due to its high P
60 content (Ye et al., 2016). In addition, algal biomass produced from wastewater treatment can serve
61 as P fertilizer in agriculture (Mulbry et al., 2005). A recent research (Zhao et al., 2018) reported that

62 high P content (2.8-3.0%) with extremely high P bioavailability (up to 98%) could be achieved in
63 the algal-bacterial AGS treating low influent carbon wastewater, which is easily used for P resource
64 recovery. Besides, loss of granular stability during the long-term operation is a major concern for
65 practical application of this promising biotechnology. However, very little information is available
66 regarding P content and its bioavailability as well as stability of algal-bacterial granules in no air
67 bubbling systems which are usually applied for cultivation of bacterial AGS or algal-bacterial AGS.

68 This preliminary trail aimed to investigate the characteristics and performance of the mature
69 algal-bacterial AGS in shaking photoreactors instead of air bubbling systems usually used for
70 bacterial AGS or algal-bacterial AGS process. Besides, P accumulation and its bioavailability in the
71 algal-bacterial granules were detected. Results from this work are expected to provide basic
72 information for the design of this cost-effective system to treat wastewater and the subsequent
73 resources recovery from the algal-bacterial AGS biomass.

74 **2. Materials and methods**

75 *2.1. Synthetic wastewater and seed algal-bacterial granules*

76 Synthetic wastewater used in this work was the same as the influent to R₁ in a previous study
77 (Zhao et al., 2018), which was mainly characterized as 400 mg COD/L, 50 mg NH₄⁺-N/L, and 10
78 mg PO₄³⁻-P/L to mimic the domestic wastewater in China.

79 Prior to the experiments, mature algal-bacterial AGS was cultivated in the laboratory-scale
80 sequencing batch reactor (SBR) which has been operated stably for more than 3 months. The
81 dominant algae species, *Leptolyngbya*, was the same as that in Zhao et al. (2018). The initial
82 concentration of mixed liquor volatile suspended solids (MLVSS) was around 3.87±0.03 g/L in
83 each photoreactor. The mean diameter of the granules was 2.32±0.52 mm with a sludge volume
84 index at 5 minutes (min) (SVI₅) about 38.1±0.4 mL/g.

85 *2.2. Photoreactors' operation and analysis*

86 Six 250 mL glass flasks (FB-501-250, Fisher Scientific, USA) with a working volume of 200
87 mL each were shaken constantly in a multi shaker (MMS-310, EYELA, Japan) at 150 rpm and
88 room temperature (25 ± 2 °C), resulting in suspended granular biomass during shaking. These
89 reactors were operated in parallel under the same conditions. Two artificial solar lights
90 (NLSS15CBM-AC, Nikki Trading Corp., Japan) were applied to the reactors with the light intensity
91 on the surface of the outer wall being 88-122 $\mu\text{mol}/(\text{m}^2\cdot\text{s})$ when the artificial solar light was on
92 (light-on/light-off=12 h/12 h). All the experiments were carried out without air bubbling (used for
93 bacterial AGS or algal-bacterial AGS cultivation and treatment processes), during which oxygen
94 could be provided by the algae through photosynthesis when the light was on. Each cycle (12 hours)
95 consisted of 1 min filling, 715 min shaking, 2 min settling, and 2 min effluent discharge. The
96 volumetric exchange ratio was kept at 50%, resulting in a hydraulic retention time (HRT) of about
97 24 h. Sludge retention time (SRT) was controlled at approximately 30 days through timely and
98 quantitative sludge discharge (once every two days). The whole test lasted for 25 days till the stable
99 system performance being achieved.

100 All the analytical methods were described elsewhere (Zhao et al., 2018), including the
101 quantification and characterization of granular biomass (ML(V)SS, SVI, chlorophyll *a*, granular
102 size and distribution, granular strength, P fractionation in biomass), water quality (total nitrogen
103 (TN), ammonia nitrogen ($\text{NH}_4^+\text{-N}$), nitrite nitrogen ($\text{NO}_2^-\text{-N}$), nitrate nitrogen ($\text{NO}_3^-\text{-N}$), phosphorus
104 ($\text{PO}_4^{3-}\text{-P}$), dissolved organic carbon (DOC), dissolved inorganic carbon (DIC), dissolved total
105 carbon (DTC)), and extracellular polymeric substances (EPS) extraction and determination.

106 2.3. Statistical analysis

107 All analyses were carried out in triplicate with the results being expressed as the mean \pm
108 standard deviation. Pearson's correlation analysis was performed by using Origin 2017 to clarify the

109 correlation between P fractions and metal ions in the granules. The significant difference was
110 assumed at $p < 0.05$.

111 3. Results and discussion

112 3.1. Properties of algal-bacterial AGS in shaking photoreactors

113 3.1.1. Changes in morphology and biomass retention

114 At the beginning, the mature algal-bacterial AGS was green, reflecting an irregular, compact
115 and dense structure. After day 20, filamentous bacteria were observed on the outer surface of the
116 granules. Gradually, the number of large-sized granules with superficial filamentous bacteria
117 increased in all the reactors. Moreover, most parts of the granules turned to dark green with a larger
118 particle diameter (3.77 ± 1.09 mm) compared to the seed granules (2.32 ± 0.52 mm). Restated,
119 although filamentous bacteria co-existed in the reactors till the end of experiment, no obvious
120 disintegration was observed in the algal-bacterial granules which maintained their compact and
121 integrated structure throughout the whole shaking test.

122 The reactors had an initial MLSS of 4.99 ± 0.06 g/L (MLVSS/MLSS= $77.6 \pm 1.1\%$), and an
123 obvious decrease in MLSS was detected after 25 days of operation (3.77 ± 0.12 g/L,
124 MLVSS/MLSS= $77.6 \pm 2.0\%$), most probably attributable to a short SRT (about 30 days) applied in
125 this study. As it is known, chlorophyll *a* concentration can be used to represent the growth of algal
126 biomass (Lee et al., 2015), which was detected to increase from the initial 7.99 ± 0.13 to 13.40 ± 0.01
127 mg/g-MLVSS in the granules from the reactors on day 25. This observation signals the fast growth
128 of algae in the algal-bacterial granules under the shaking condition.

129 3.1.2. Granular settleability and stability

130 Effective biomass separation through the formation of fast settling biomass is one of the major
131 advantages of AGS technology. Under the designed shaking condition, an obvious increase in SVI₅
132 (from 38.1 ± 0.4 to 77.5 ± 1.8 mL/g) was noticed, suggesting somewhat worsen settleability of algal-

133 bacterial granules. This observation was partially in agreement with the finding by Abouhend et al.
134 (2018) who attributed the increase in SVI_{30} of oxygenic photogranules to the overgrowth of
135 filamentous green algae in the stirred-tank reactors without aeration. The presence of filamentous
136 bacteria and the low hydrodynamic shear force provided by the shaker might be the key factors for
137 the resultant slightly lower granular settleability than the seed algal-bacteria granules. Thus some
138 decrease in granular settling velocity (from 29.1 ± 3.9 to 21.4 ± 6.2 m/h) was observed in this study.
139 Still, the SVI_5 of algal-bacterial AGS on day 25 was better than or comparable to the conventional
140 AGS (55.3-82.8 mL/g) or algae granules (~125 mL/g) in the air bubbling systems (Cai et al., 2019;
141 Wang et al., 2019).

142 Besides, the stability of granular sludge is a crucial factor to ensure the effluent quality from the
143 algal-bacterial granule system. In this study, the granular stability was expressed in terms of
144 integrity coefficient (%), and a lower value denotes a better stability (Muda et al., 2010). It was
145 found that the integrity coefficient of the granules only slightly increased from the initial 7.4 ± 0.3 to
146 $8.4 \pm 0.6\%$ on day 25, implying that the algal-bacterial AGS possesses a strong enough structure to
147 maintain its granular stability even under the constantly shaking condition.

148 *3.2. Performance on carbon and nutrients removal*

149 *3.2.1. DOC removal*

150 The changes in effluent carbon concentration from the reactors were recorded during the 25
151 days' operation. The effluent DOC concentration from the reactors was always < 14 mg/L from day
152 2 onwards, achieving an average DOC removal efficiency of $94.8 \pm 1.6\%$ and $94.4 \pm 1.4\%$ during the
153 light-on and light-off cycles of the whole test period. This observation indicates that the algal-
154 bacterial AGS system could also have good performance on organics removal under shaking
155 condition, and the light illumination exhibited no obvious influence on organics removal by the
156 algal-bacterial AGS. A similar organic removal efficiency was also achieved from the AGS systems

157 with bacterial AGS (averagely 96.0%) or algal-bacterial AGS (averagely 96.2%) under air bubbling
158 (Cai et al., 2018; Zhao et al., 2018). It is worth noting that an enhanced DIC reduction was detected
159 during the light cycle from day 11 onwards, which might be attributable to the photosynthesis of
160 algae. That is, light illumination can influence DIC removal process to some extent.

161 3.2.2. *Nutrients removal*

162 In this study, the algal-bacterial AGS exhibited excellent performance on $\text{NH}_4^+\text{-N}$ removal (>
163 99%) during the light-on and light-off cycles from the very beginning of this test, indicating that
164 stable nitrification could be quickly established in this shaking system. TN removal efficiencies in
165 the reactors were almost at the same levels, $71.1\pm 3.3\%$ and $69.2\pm 4.1\%$ during the light-on and light-
166 off cycles, respectively from day 3 to the end of experiment. Compared to a previous report by
167 using the same algal-bacterial AGS in the air bubbling system (Zhao et al. 2018), a higher TN
168 removal efficiency was detected in this study. This suggests that algal-bacterial AGS possesses
169 great potential for N removal even under shaking condition, which could be further optimized with
170 the relationship between DO change in bulk liquor and light illumination duration being considered.
171 Interestingly, the TN removal efficiency in the photoreactors demonstrated an evidently increasing
172 tendency along with the operation. This observation might be attributable to the bigger granule size
173 with the operation, creating a more beneficial environment for denitrification (Li et al., 2017).

174 The effluent P concentration from the reactors during the light-on and light-off cycles, however,
175 showed an obviously different trend during the operation. When fed with 10 mg/L of P, the reactors
176 produced the effluents with P concentrations about 4.5 ± 1.7 mg/L (varied from 2.8 to 8.5 mg/L) and
177 13.5 ± 4.1 mg/L (ranged between 9.1-20.9 mg/L), respectively during the light-on and light-off
178 cycles from day 3 to the end of experiments. Biotic assimilation and abiotic precipitation are
179 reported as the main P removal means from wastewater (de Godos et al., 2009). Algae can absorb
180 and accumulate large quantities of P through luxury uptake (Solovchenko et al., 2016). In the case

181 of algal-bacterial AGS system, co-precipitation of phosphate with other co-existing metals and
182 algae assimilation are the major contributors to P removal (Zhao et al., 2018). As seen, almost no P
183 removal was detected during the light-off cycles in the shaking photoreactors, suggesting algae
184 growth and assimilation might be responsible for the enhanced P removal with light illumination in
185 this study. Further research is necessary for the optimization of light cycle for the photoreactors to
186 improve P uptake and effluent quality.

187 3.2.3. Cycle tests on day 20

188 In order to have a better understanding of the algal-bacterial AGS system under shaking
189 condition, typical cycle tests (including DOC, TP, DO and N profiles) were also conducted and
190 compared during the light-on and light-off cycles on day 20.

191 DO concentration decreased from the initial 2.7 and 2.6 mg/L to 1.37 and 0.83 mg/L at the 60th
192 min during the light-on and light-off cycles, respectively. The observation indicates that the activity
193 of aerobic bacteria (nitrifiers and heterotrophs) could be maintained in the shaking photoreactors.
194 From the 60th min to the 720th min, the DO values continuously increased, and reached around 8.2
195 and 3.5 mg/L at the end of light-on and light-off cycles. As for the DOC concentration, a similar
196 decreasing trend was observed during the light-on and light-off cycles, which sharply declined to
197 37.5 and 32.1 mg/L respectively at the 60th min, and then gradually decreased to 5.8 and 7.7 mg/L
198 at the end of each cycle. On the other hand, the decrease of $\text{NH}_4^+\text{-N}$ concentration in the reactors
199 was slightly faster during the light-on cycle than that during the light-off cycle, indicating that
200 besides algae assimilation, nitrification with simultaneous denitrification processes may occur under
201 shaking and illumination conditions.

202 As for P, TP concentration was firstly observed to release and then uptake during the subsequent
203 period, suggesting that P was not only taken up by algae but also removed by phosphorus
204 accumulating organisms (PAOs) in this study. Interestingly, a higher P release (22.6 mg/L at the

205 120th min) during the light-off cycle was detected, in comparison to 16.7 mg/L of P release during
206 the light-on cycle (at the 60th min). In addition, the effluent P concentrations were averagely about
207 2.5 and 10.0 mg/L at the end of light-on and light-off cycles, respectively. Almost little contribution
208 of algal-bacterial AGS to P removal was observed during the light-off period, most probably
209 attributable to the designed light-on/light-off cycle (12h/12h) which might not be suitable for the
210 metabolism of PAOs. More in-depth research is still ongoing with respect to the optimization of
211 light-on/light-off cycle and hydraulic/sludge retention time.

212 3.3. P fractionation in algal-bacterial granules

213 The P fractions in granules are presented in Fig. 1, including total phosphorus (TP), organic
214 phosphorus (OP), inorganic phosphorus (IP), non-apatite inorganic phosphorus (NAIP) and apatite
215 phosphorus (AP). **The quantitative relationship among these P fractions can be expressed as**
216 **follows: $TP = OP + IP$, and $IP = NAIP + AP$.** According to Huang et al. (2015), NAIP is mainly
217 composed of the fraction associated with Al, Fe and Mn, which together with OP is regarded as
218 bioavailable P that can be potentially released and utilized by microorganisms and plants. AP is the
219 Ca-bound P that is hard to be utilized by microorganisms and plants. It is clearly seen that the
220 content of each P fraction in the seed algal-bacterial AGS was much higher than those in the
221 granules on day 15 and day 25 (Fig. 1a). The TP content in the granules decreased obviously, from
222 the initial 30.4 ± 0.4 to 23.8 ± 0.3 on day 15 and to 21.5 ± 0.4 mg/g-MLSS on day 25, respectively.
223 Correspondingly, the OP and NAIP contents decreased from the initial 5.8 ± 0.3 (occupying 19% of
224 TP) and 22.5 ± 0.3 mg/g-MLSS (74% of TP) to 3.1 ± 0.2 and 16.7 ± 0.1 mg/g-MLSS on day 25,
225 accounting for 14% and 78% of TP in the granules, respectively. However, the AP content
226 maintained at around 1.6 ± 0.1 mg/g-MLSS in the granules during the whole process, signaling its
227 relative stability in quantity under the test condition. This observation also indicates that the
228 decrease of TP content in the granules is mainly contributed by OP and NAIP under the shaking

229 condition. Specifically, Fe content in the granules reflected a similar decrease trend as NAIP, and a
230 strongly positive correlation ($R = 0.999$) was found between the amount of Fe accumulated in the
231 granules and its NAIP content. This result suggests that it might be difficult to maintain the stable
232 Fe-PO₄ precipitates in the algal-bacterial granules under the designed operation condition, which
233 needs further investigation. However, the proportion of potentially bioavailable P ((OP+NAIP)/TP)
234 was relatively stable, which varied from the initial 93.2% to 92.0% in the granules on day 25. As a
235 result, the test condition has limited effect on the bioavailable P in the algal-bacterial granules,
236 which can be potentially recovered.

237 *3.4. EPS secretion from algal-bacterial granules*

238 EPS play an essential role in the formation and stability of granule (Guo et al., 2016). As shown
239 in Fig. 1b. The total EPS content increased significantly from the initial 100.0 ± 1.5 mg/g-MLVSS to
240 174.5 ± 4.8 mg/g-MLVSS on day 25, in which proteins (PN) was much more abundant than
241 polysaccharides (PS). During the operation, PN increased from the initial 86.2 ± 1.9 to 153.7 ± 2.3
242 mg/g-MLVSS on day 25, and the PN/PS ratio rose from 6.3 to 7.4, correspondingly. The increment
243 in PN excretion in this study indicates that PN might function as protective agents for bacteria and
244 maintain the stability of algal-bacterial granules under the shaking condition (Zhang et al., 2018).

245 Previous works indicate that EPS may have a different content distribution in the layered
246 structure of AGS (Adav et al., 2008), which can be mainly divided into two different fractions, i.e.
247 loosely bound EPS (LB-EPS) and tightly bound EPS (TB-EPS) (Li and Yang, 2007). Fig. 1b also
248 shows the changes of LB-EPS and TB-EPS contents with their major components (PN and PS)
249 during the 25 days' operation. In general, the PN and PS contents in the LB-EPS and TB-EPS
250 increased with a similar trend as those in the total EPS in the granules. The PN/PS ratio slightly
251 decreased from the initial 5.6 to 5.5 in LB-EPS, whereas it increased from 6.6 to 9.0 in TB-EPS on
252 day 25. This observation suggests that PN in the TB-EPS is more sensitive than those in the LB-

253 EPS of algal-bacterial granules under the test condition. The above results also imply that PN play a
254 crucial role in the maintenance of the stable inner structure of algal-bacterial AGS.

255 **4. Conclusions**

256 **The stability and performance of mature algal-bacterial AGS was examined by using shaking**
257 **photoreactors instead of air bubbling systems.** The algal-bacterial AGS possessed good granular
258 integrity ($8.4\pm 0.6\%$), excellent DOC removal ($94.8\pm 1.6\%$) and high TN removal ($71.1\pm 3.3\%$)
259 during 25 days' operation. The stability of Fe-PO₄ precipitates in the granules was affected to some
260 extent **under the test condition**, during which a high P bioavailability (92.0%) was still maintained.
261 Further research is directed to the optimization of light-on/light-off cycle and hydraulic/sludge
262 retention time in order to realize the minimum energy consumption for wastewater treatment by
263 using this novel algal-bacterial AGS system.

264 **Acknowledgments**

265 This work was supported by JSPS KAKENHI Grant Numbers JP18H03403 and JP16H06382.

266 **Appendix. A. Supplementary data**

267 E-supplementary data associated with this article can be found in the online version.

268 **References**

- 269 1. Abouhend, A. S., McNair, A., Kuo-Dahab, W. C., Watt, C., Butler, C. S., Milferstedt, K.,
270 Hamelin, J., Seo, J., Gikonyo, G. J., El-Moselhy, K. M., Park, C., 2018. The oxygenic
271 photogranule process for aeration-free wastewater treatment. *Environ. Sci. Technol.* 52, 3503-
272 3511.
- 273 2. Adav, S. S., Lee, D. J., Tay, J. H., 2008. Extracellular polymeric substances and structural
274 stability of aerobic granule. *Water Res.* 42, 1644-1650.
- 275 3. Ahmad, J. S. M., Cai, W., Zhao, Z., Zhang, Z., Shimizu, K., Lei, Z., Lee, D.-J., 2017. Stability
276 of algal-bacterial granules in continuous-flow reactors to treat varying strength domestic
277 wastewater. *Bioresour. Technol.* 244, 225-233.
- 278 4. Boelee, N. C., Temmink, H., Janssen, M., Buisman, C. J. N., Wijffels, R. H., 2014. Balancing
279 the organic load and light supply in symbiotic microalgal-bacterial biofilm reactors treating

- 280 synthetic municipal wastewater. *Ecol. Eng.* 64, 213-221.
- 281 5. Cai, W., Jin, M., Zhao, Z., Lei, Z., Zhang, Z., Adachi, Y., Lee, D. -J., 2018. Influence of ferrous
282 iron dosing strategy on aerobic granulation of activated sludge and bioavailability of
283 phosphorus accumulated in granules. *Bioresour. Technol. Rep.* 2, 7-14.
- 284 6. Cai, W., Zhao, Z., Li, D., Lei, Z., Zhang, Z., Lee, D. -J., 2019. Algae granulation for nutrients
285 uptake and algae harvesting during wastewater treatment. *Chemosphere*, 214, 55-59.
- 286 7. de Godos, I., Blanco, S., García-Encina, P.A., Becares, E., Muñoz, R., 2009. Long-term
287 operation of high rate algal ponds for the bioremediation of piggery wastewaters at high loading
288 rates. *Bioresour. Technol.* 100, 4332-4339.
- 289 8. Guo, J., Wang, S., Lian, J., Ngo, H. H., Guo, W., Liu, Y., Song, Y., 2016. Rapid start-up of the
290 anammox process: Effects of five different sludge extracellular polymeric substances on the
291 activity of anammox bacteria. *Bioresour. Technol.* 220, 641-646.
- 292 9. Hu, Y., Hao, X., van Loosdrecht, M., Chen, H., 2017. Enrichment of highly settleable
293 microalgal consortia in mixed cultures for effluent polishing and low-cost biomass production.
294 *Water Res.* 125, 11-22.
- 295 10. Huang, W., Cai, W., Huang, H., Lei, Z., Zhang, Z., Tay, J. H., Lee, D.-J., 2015. Identification of
296 inorganic and organic species of phosphorus and its bio-availability in nitrifying aerobic
297 granular sludge. *Water Res.* 68, 423-431.
- 298 11. Lee, C. S., Lee, S. A., Ko, S. R., Oh, H. M., Ahn, C. Y., 2015. Effects of photoperiod on nutrient
299 removal, biomass production, and algal-bacterial population dynamics in lab-scale
300 photobioreactors treating municipal wastewater. *Water Res.* 68, 680-691.
- 301 12. Li, X.Y., Yang, S.F., 2007. Influence of loosely bound extracellular polymeric substances
302 (EPS) on the flocculation, sedimentation and dewaterability of activated sludge. *Water Res.* 41,
303 1022-1030.
- 304 13. Li, X., Luo, J., Guo, G., Mackey, H. R., Hao, T., Chen, G., 2017. Seawater-based wastewater
305 accelerates development of aerobic granular sludge: a laboratory proof-of-concept. *Water Res.*
306 115, 210-219.
- 307 14. Liu, L., Zeng, Z., Bee, M., Gibson, V., Wei, L., Huang, X., Liu, C., 2018. Characteristics and
308 performance of aerobic algae-bacteria granular consortia in a photo-sequencing batch reactor. *J.*
309 *Hazard. Mater.* 349, 135-142.
- 310 15. Muda, K., Aris, A., Salim, M. R., Ibrahim, Z., Yahya, A., van Loosdrecht, M. C., Ahmad, A.,
311 Nawahwi, M. Z., 2010. Development of granular sludge for textile wastewater treatment. *Water*

- 312 Res. 44, 4341-4350.
- 313 16. Mulbry, W., Westhead, E. K., Pizarro, C., Sikora, L., 2005. Recycling of manure nutrients: use
314 of algal biomass from dairy manure treatment as a slow release fertilizer. *Bioresour. Technol.*
315 96, 451-458.
- 316 17. Quijano, G., Arcila, J. S., Buitrón, G., 2017. Microalgal-bacterial aggregates: applications and
317 perspectives for wastewater treatment. *Biotechnol. Adv.* 35, 772-781.
- 318 18. Sforza, E., Pastore, M., Sanchez, S.S., Bertucco, A., 2018. Bioaugmentation as a strategy to
319 enhance nutrient removal: Symbiosis between *Chlorella protothecoides* and *Brevundimonas*
320 *diminuta*. *Bioresour. Technol. Rep.* 4, 153-158.
- 321 19. Solovchenko, A., Verschoor, A. M., Jablonowski, N. D., Nedbal, L., 2016. Phosphorus from
322 wastewater to crops: an alternative path involving microalgae. *Biotechnol. Adv.* 34, 550-564.
- 323 20. Tiron, O., Bumbac, C., Patroescu, I. V., Badescu, V. R., Postolache, C., 2015. Granular activated
324 algae for wastewater treatment. *Water Sci. Technol.* 71, 832-839.
- 325 21. Wang, X., Chen, Z., Shen, J., Zhao, X., Kang, J., 2019. Impact of carbon to nitrogen ratio on the
326 performance of aerobic granular reactor and microbial population dynamics during aerobic
327 sludge granulation. *Bioresour. Technol.* 271, 258-265.
- 328 22. Ye, Y., Ngo, H. H., Guo, W., Liu, Y., Zhang, X., Guo, J., Ni, B., Chang, S. W., Nguyen, D. D.,
329 2016. Insight into biological phosphate recovery from sewage. *Bioresour. Technol.* 218, 874-
330 881.
- 331 23. Zhang, B., Lens, P.N., Shi, W., Zhang, R., Zhang, Z., Guo, Y., Cui, F., 2018. Enhancement
332 of aerobic granulation and nutrient removal by an algal-bacterial consortium in a lab-scale
333 photobioreactor. *Chem. Eng. J.* 334, 2373-2382.
- 334 24. Zhao, Z., Yang, X., Cai, W., Lei, Z., Shimizu, K., Zhang, Z., Utsumi, M., Lee, D. J., 2018.
335 Response of algal-bacterial granular system to low carbon wastewater: Focus on granular
336 stability, nutrients removal and accumulation. *Bioresour. Technol.* 268, 221-229.
- 337 25. Zhou, D., Zhang, C., Fu, L., Xu, L., Cui, X., Li, Q., Crittenden, J. C., 2017. Responses of the
338 microalga *Chlorophyta* sp. to bacterial quorum sensing molecules (N-acylhomoserine lactones):
339 Aromatic protein-induced self-aggregation. *Environ. Sci. Technol.* 51, 3490-3498.

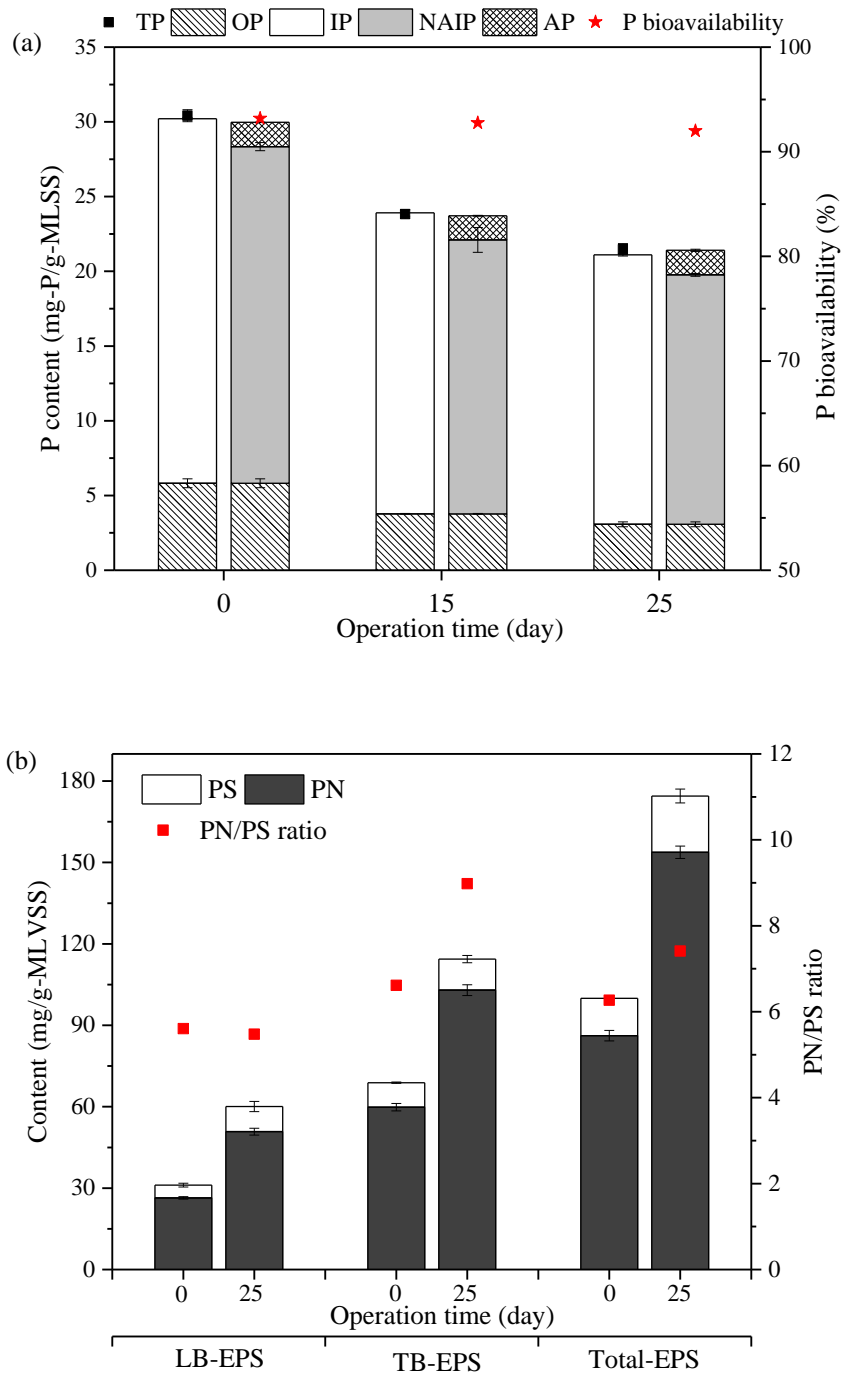


Fig. 1. Changes in P content and its fractions (a), and loosely bound EPS (LB-EPS), tightly bound EPS (TB-EPS), and total EPS contents and their major components (proteins (PN) and polysaccharides (PS)) in the algal-bacterial granules.

Highlights

- Highly stable algal-bacterial granules were maintained after shaking 25 days
- Algal-bacterial granules removed about 95% dissolved organic C and 71% total N
- 55% of total P removal was achieved during light-on cycles in shaking photoreactors
- 92% of total P was still bioavailable in the algal-bacterial AGS after the test
- Tightly bound extracellular proteins played a crucial role in this study

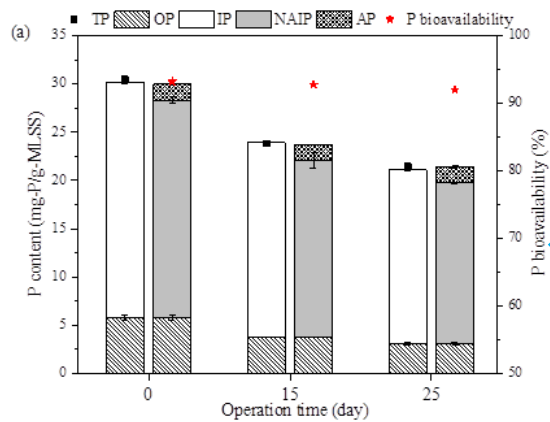
Graphical abstract

Good granular integrity (~8.4%)

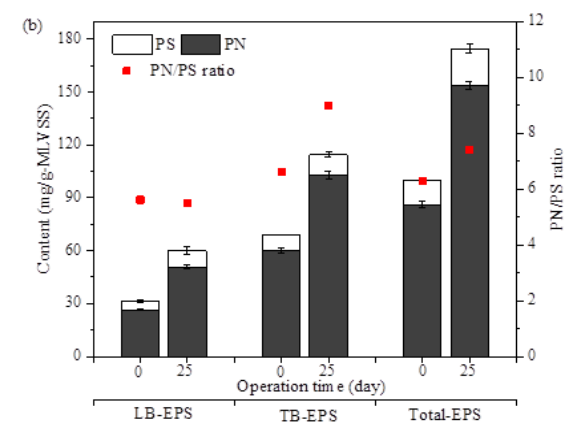
Excellent DOC removal ($94.8 \pm 1.6\%$)

High TN removal ($71.1 \pm 3.3\%$)

High TP ($54.8 \pm 17.1\%$) removal (light-on)



High P bioavailability (92%)



More extracellular proteins (153.7±2.3mg/g)

Algal-bacterial AGS after 25 days' shaking

(instead of air bubbling used for bacterial AGS or algal-bacterial AGS cultivation)

Supplementary Materials

Stability and performance of algal-bacterial granular sludge in shaking photo-sequencing batch reactors with special focus on phosphorus accumulation

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Materials and Methods

Determination of granular strength

Integrity coefficient (%) was used to estimate the strength of algal-bacterial granules by a shock method as described by Ghangrekar et al. (2005), which was defined as the ratio of solids in the supernatant after being shaken on a platform shaker at 200 rpm for 5 min to the total weight of the granular sludge used for this test.

Reference

1. Ghangrekar, M. M., Asolekar, S. R., Joshi, S. G., 2005. Characteristics of sludge developed under different loading conditions during UASB reactor start-up and granulation. *Water Res.* 39, 1123-1133.

Results and discussion

Table S1

Comparison in characteristics of algal-bacterial AGS in the shaking photoreactors between day 0 and day 25.

Parameter	Day 0	Day 25
MLSS (g/L)	4.99±0.06	3.77±0.12
MLVSS/MLSS (%)	77.6±1.1	77.6±2.0
SVI ₅ (mL/g)	38.1±0.4	77.5±1.8
Mean diameter (mm)	2.32±0.52	3.77±1.09
Integrity coefficient (%)	7.4±0.3	8.4±0.6
Settling velocity (m/h)	29.1±3.9	21.4±6.2
Chlorophyll <i>a</i> (mg/g-MLVSS)	7.99±0.13	13.40±0.01

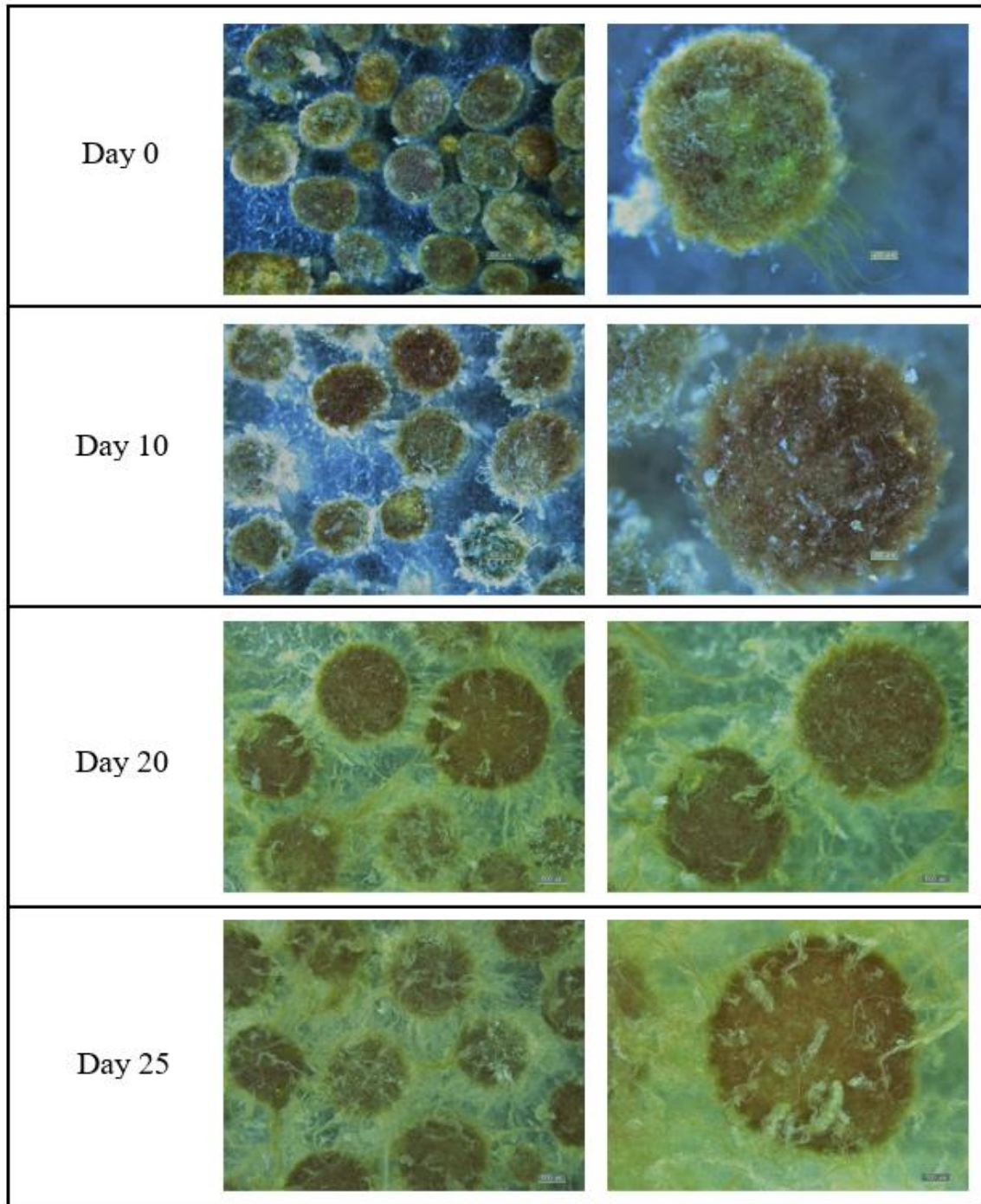


Fig. S1. Morphological changes of the algal-bacterial AGS in the two reactors on day 0, 10, 20, and 25, respectively.

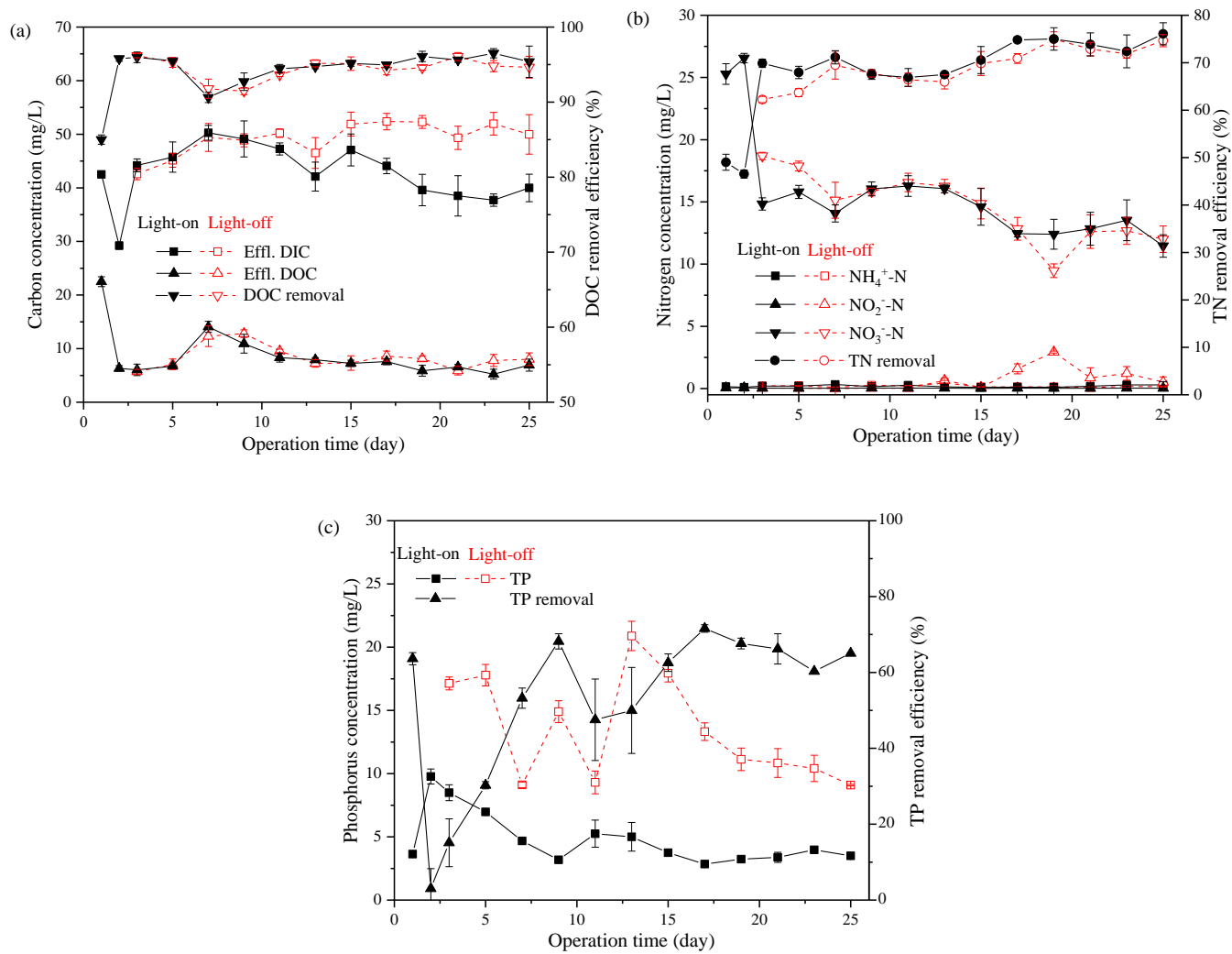


Fig. S2. Changes in nutrients profiles during the operation: carbon (a), nitrogen (b), and phosphorus (c).

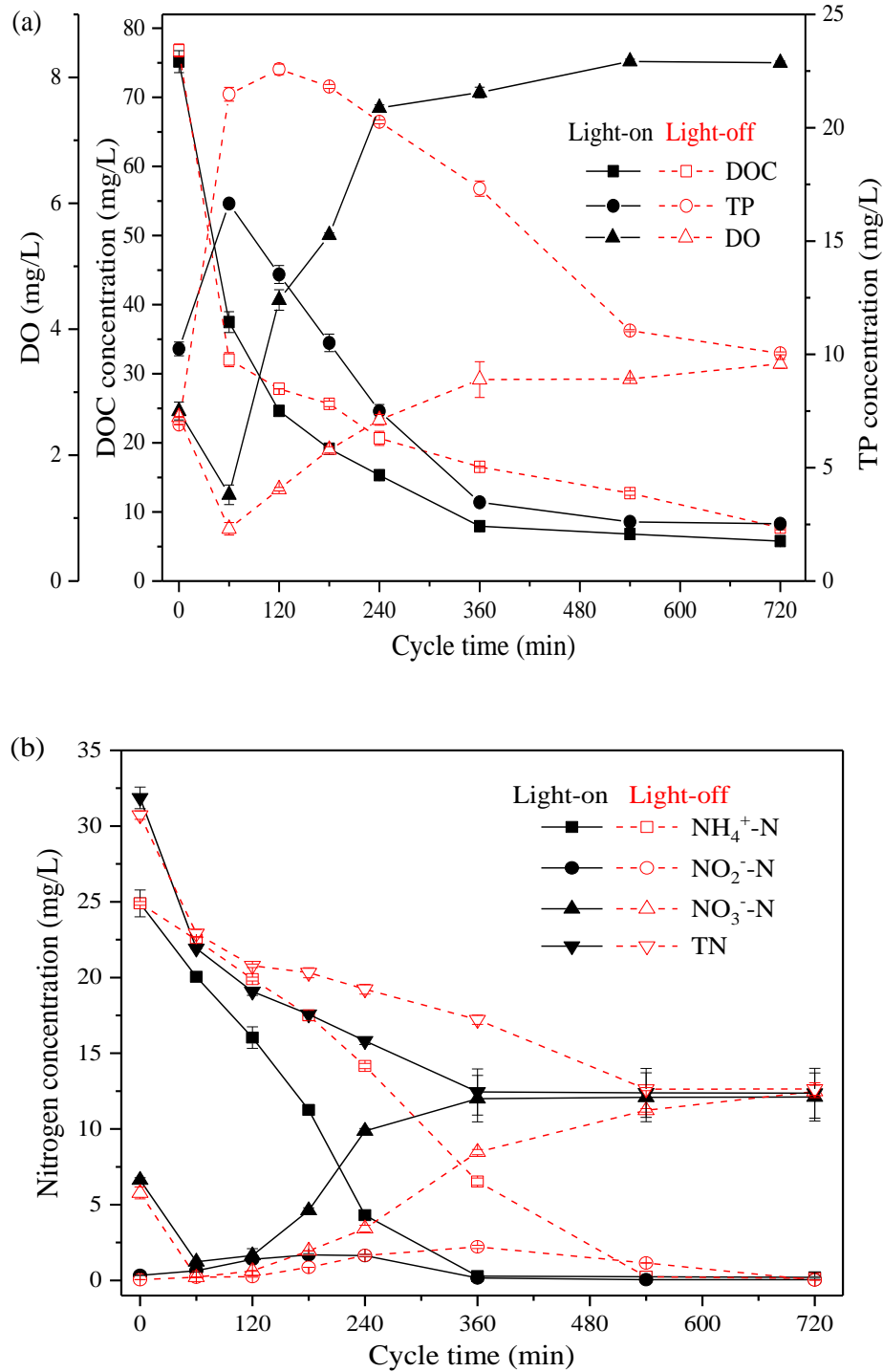


Fig. S3. Variations of DO, DOC, and TP (a), and N species (b) in the typical light-on and light-off cycles on day 20.

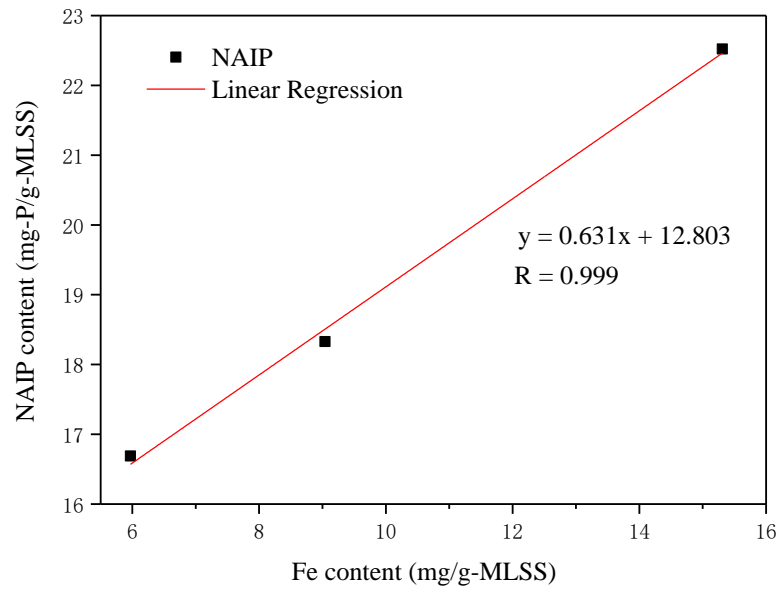


Fig. S4. The correlation of Fe content and NAIP content in the granules.