Supporting Information for

Mechanistic Insights into Catalytic Oxidations of
Organic Compounds by Ruthenium(IV)-Oxo
Complexes with Pyridylamine Ligands

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General.

Chemicals and solvents were used as received from Tokyo Chemical Industry (TCI) Co., Wako chemicals, or Sigma-Aldrich Corp. unless otherwise mentioned. $[\text{RuCl}_2(\text{DMSO})_4]^1$ and di-(2-pyridyl)methane amine² were prepared by literature procedures. N4Py was synthesized with a modified procedure reported in a literature (*vide infra*).² UV-vis spectra were collected on a Shimadzu UV-3600 spectrophotometer. ¹H NMR spectra were recorded on a JEOL EX-270 spectrometer at room temperature and the chemical shifts of signals were determined with respect to residual proton signals of deuterated solvents. Electrochemical measurements were performed with an AUTOLAB PGSTAT12 potentiometer in Britton-Robinson (B.-R.) buffer (pH = 2~12) at room temperature. pH measurements were made with a Horiba pH-Meter F-51. The sample solution of complex **6** for resonance Raman scattering measurements was prepared with a 2.0 mM solution of **3** (50 mL) in H₂¹⁶O or H₂¹⁸O, which was oxidized by addition of a 20 mM aqueous solution of CAN (20 mL).

Synthesis of *N,N*-bis(2-pyridylmethyl)-*N*-(bis-2-pyridylmethyl)amine (N4Py).² Di-(2-pyridyl)methane amine² (0.90 g, 4.9 mmol) and picolyl chloride hydrochloride (1.67 g, 10.1 mmol) were condensed in NaOH (aq) (10 M, 1 mL) and the mixture was stirred for overnight. The mixture was extracted with CHCl₃, and then the solvent of the combined organic phase was evaporated. The obtained oil was purified by column chromatography as a stationary state eluted with CHCl₃ : MeOH = 7 : 3 mixed solvent, and then the solvent of the collected fraction was evaporated under vacuum. Yield: 0.49 g (1.3 mmol, 27 %). ¹H NMR (CDCl₃): δ 3.69 (s, 4H, CH₂), 5.34 (s, 1H, CH), 7.13 (m, 4H, Py), 7.62 (m, 8H, Py), 8.48 (d, J = 7 Hz, 2H, Py), 8.57 (d, J = 7 Hz, 2H, Py).

Synthesis of $[\text{Ru}^{\text{II}}\text{Cl}(\text{N4Py})](\text{PF}_6)$. N4Py (0.37 g, 1.0 mmol) and $[\text{RuCl}_2(\text{DMSO})_4]^1$ (0.54 g, 1.0 mmol) were charged in a three-necked flask under Ar atmosphere. 2-Propanol (50 mL) was added with a syringe, and then the solution was refluxed for 14 h. After cooling to room temperature, the solvent was completely removed and the resultant dark-brown material was dissolved in water. Upon addition of excess NH₄PF₆, red-brownish solid emerged and the precipitate was filtered and washed with diethyl ether to remove organic materials. The obtained $[\text{Ru}^{\text{II}}\text{Cl}(\text{N4Py})](\text{PF}_6)^3$ (0.43 g, 0.80 mmol) was purified by column chromatography on Al₂O₃ as a stationary state eluted with a CHCl₃/MeOH (9 : 1 v/v) mixed solvent, and then the solvent of the collected fraction was evaporated under vacuum. The residual solid was recrystallized from MeOH, filtered, washed with diethyl ether and then dried in vacuo. Yield: 0.22 g (43%). ESI-MS (methanol): m/z = 504 ($[\text{M} - \text{PF}_6]^+$). ¹H NMR (D₂O): δ 4.31 and 4.47 (ABq, J_{AB} = 18, 4 Hz, 4H, N-CH₂-Py), 6.44 (s, 1H, N-CH-Py₂), 7.01 (d, J = 8 Hz, 2H, H3 of Py₂-CH-N), 7.21 (dd, J = 7, 2 Hz, 2H, H5 of Py₂-CH-N), 7.32 (dd, J = 6, 2 Hz, 2H, H5 of

Py-CH₂-N), 7.54 (t, *J* = 8 Hz, 4H, H4 of *Py*₂-CH-N), 7.86-7.89 (m, 4H, H3 of *Py*-CH₂-N and H4 of *Py*-CH₂-N), 8.89 (d, *J* = 6 Hz, 2H, H6 of *Py*-CH₂-N), 9.11 (d, *J* = 6 Hz, 2H, H6 of *Py*₂-CH-N). Elemental analysis calcd (%) for C₂₃H₂₁N₅RuClPF₆: C 42.57, H 3.26, N 10.79; found: C 42.43, H 3.25, N 10.24; UV-Vis (H₂O): λ_{max} [nm] = 448, 366, 246.

Synthesis of [Ru^{II}(N4Py)(OH₂)](PF₆)₂ (3**).** An H₂O solution (80 mL) containing [RuCl(N4Py)](PF₆) (0.27 g, 0.50 mmol) and AgPF₆ (1.26 g, 5.0 mmol) was refluxed for 3 h. Insoluble white solid was removed by a membrane filter (ADVANTEC H100A025A), and then the solvent of the filtrate was concentrated to a 1/10 volume to obtain a yellow precipitate. The yellow precipitate was recrystallized from a minimal amount of warmed H₂O. The resulting yellow powder was filtered, washed with diethyl ether, and then, dried in vacuo. Yield: 0.22 g (56%). ESI-MS (methanol): *m/z* = 234 ([M – H₂O – 2PF₆]²⁺). ¹H NMR (D₂O): δ 4.31 and 4.44 (ABq, *J*_{AB} = 18, 4 Hz, 4H, N-CH₂-Py), 6.44 (s, 1H, N-CH-Py₂), 7.05 (d, *J* = 8 Hz, 2H, H3 of *Py*₂-CH-N), 7.29 (dd, *J* = 7, 2 Hz, 2H, H5 of *Py*₂-CH-N), 7.37 (dd, *J* = 6, 2 Hz, 2H, H5 of *Py*-CH₂-N), 7.59 (t, *J* = 7 Hz, 4H, H4 of *Py*₂-CH-N), 7.90-7.94 (m, 4H, H3 of *Py*-CH₂-N and H4 of *Py*-CH₂-N), 8.83 (d, *J* = 6 Hz, 2H, H6 of *Py*-CH₂-N), 8.93 (d, *J* = 6 Hz, 2H, H6 of *Py*₂-CH-N). Anal.: calcd (%) for C₂₃H₂₃N₅ORuP₂F₁₂: C 35.58, H 2.99, N 9.02; found: C 35.43, H 3.07, N 9.07. UV-Vis (H₂O): λ_{max} [nm] = 439, 361, 246.

Synthesis of [Ru^{IV}(O)(N4Py)](PF₆)₂ (6**).** The oxo complex **6** was formed by the treatment of **3** (0.77 mg, 1.0 μmol) with 10 equiv of CAN (5.4 mg, 10 μmol) in D₂O (0.5 mL) for ¹H NMR spectroscopy. In addition, ¹⁸O-labeled oxo complex **6** was produced by the reaction of **3** (0.77 mg, 1.0 μmol) with an excess amount of CAN (5.4 mg, 10 μmol) in H₂¹⁸O (0.5 mL) for ESI-MS spectrometry. ESI-MS (H₂O): *m/z* = 242.5 ([M – 2PF₆]²⁺); (H₂¹⁸O): *m/z* = 243.5 ([M – 2PF₆]²⁺). ¹H NMR (D₂O): δ 5.26 and 5.68 (ABq, *J*_{AB} = 17, 4 Hz, 4H, N-CH₂-Py), 7.35 (d, *J* = 8 Hz, 2H, H3 of *Py*₂-CH-N), 7.41 (s, 1H, N-CH-Py₂), 7.50 (dd, *J* = 7, 2 Hz, 2H, H5 of *Py*₂-CH-N), 7.63 (dd, *J* = 6, 2 Hz, 2H, H5 of *Py*-CH₂-N), 7.82 (dd, *J* = 8, 4 Hz, 4H, H4 of *Py*₂-CH-N), 8.10-8.22 (m, 4H, H3 of *Py*-CH₂-N and H4 of *Py*-CH₂-N), 8.43 (d, *J* = 6 Hz, 2H, H6 of *Py*-CH₂-N), 9.00 (d, *J* = 6 Hz, 2H, H6 of *Py*₂-CH-N). UV-vis (H₂O): λ_{max} [nm] = 264. Resonance Raman: ν [cm⁻¹] = 801 (H₂¹⁶O); ν [cm⁻¹] = 761 (H₂¹⁸O).

Iodometry of CAN. The concentration of Na₂S₂O₃ (aq) was determined to be 22.3 mM by titration of the solution into 3.4 mM aqueous KIO₃ solution in the presence of excess KI and starch, where the completion was indicated by disappearance of the purple color of the I₃-starch complex. Then, into CAN (122.7 mg) solution in H₂O (10 mL), to which was added excess KI and starch, was titrated the above Na₂S₂O₃ solution. The purity of CAN was determined to be 95 %.

Reduction potential of CAN.⁴ Reduction potential of CAN was obtained by differential pulse voltammograms (DPV) in 0.1 M NaClO₄ (aq) (pH = 5.68) at room temperature, where the concentration of CAN was 1.0 mM and the scan rate was 0.1 V/s. The reduction potential was determined to be +1.1 V relative to SCE.

Resonance Raman Spectroscopy. The resonance Raman spectra of complex **6** were measured as follows: To a 50 μ L of H₂¹⁶O or H₂¹⁸O solution of **3** (2.0 mM), was added a 20 μ L aqueous solution of CAN (20 mM). Resonance Raman scattering was excited at 363.8 nm from an Ar⁺ laser (Spectra Physics, 2080-25), dispersed by a single polychromator (Ritsu Oyo Kogaku, MC-100DG) and detected by a liquid-nitrogen-cooled CCD detector (Roper Scientific, LNCCD-1100-PB). The resonance Raman measurements were carried out at 22 °C using a spinning NMR tube (outer diameter = 5 mm, wall thickness = 0.24 mm) at 135° back-scattering geometry.

X-ray Crystallography on [Ru(N4Py)(OH₂)](PF₆)₂ (3**).** A single crystal of [Ru(N4Py)(OH₂)](PF₆)₂ (**3**) was obtained by recrystallization with slow evaporation of the CH₂Cl₂ solution in the presence of octane as a poor solvent. All measurements were performed at 120 K on a Bruker APEXII Ultra diffractometer. The structure was solved by a direct method (SIR-97) and expanded with differential Fourier technique. All non-hydrogen atoms were refined anisotropically and the refinement was carried out with full matrix least squares on *F*. All calculations were performed using the Yadokari-XG crystallographic software package.⁵ Crystallographic data: C₂₃H₂₃N₅ORu·2PF₆·CH₂Cl₂, FW = 902.86, red-orange, monoclinic, *P*2₁/*n*, *a* = 11.3277(18) Å, *b* = 18.234(3) Å, *c* = 15.932(3) Å, β = 97.573(2)°, *V* = 3262.1(9) Å³, *Z* = 4, *D*_{calcd} = 1.838 g cm⁻³, *F*₀₀₀ = 1792, *R*1 = 0.0512 (*I* > 2.00σ(*I*)), *wR* = 0.1407 (all data), GOF = 1.033. CCDC-875411 contains the supplementary crystallographic data. This data can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

Reference and Notes.

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Table S1 Selected bond lengths (Å) and angles (°) of **3**.

Bond lengths (Å)			
		O1-Ru-N4	97.48(19)
Ru–O1	2.172(5)	O1-Ru-N5	98.97(18)
Ru–N1	1.967(5)	N1-Ru-N2	82.22(17)
Ru–N2	2.057(4)	N1-Ru-N3	81.91(17)
Ru–N3	2.052(4)	N1-Ru-N4	83.16(18)
Ru–N4	2.061(4)	N1-Ru-N5	83.33(17)
Ru–N5	2.060(5)	N2-Ru-N3	85.62(17)
		N2-Ru-N4	165.38(18)
Bond angles (°)		N2-Ru-N5	90.69(17)
O1-Ru-N1	177.63(18)	N3-Ru-N4	92.25(17)
O1-Ru-N2	97.12(18)	N3-Ru-N5	165.14(19)
O1-Ru-N3	95.77(19)	N4-Ru-N5	87.68(17)

Fig. S1 Spectral changes during pH titration of **3** in B.-R. buffer (sample concentration: 0.1 mM) at room temperature. The spectral changes in the pH range (a) of 0.4–3.8 and (b) of 6.2–14.2 and the plot of the absorbance change at 470 nm relative to pH in the range (c) of pH = 0.4–3.8 and (d) of pH = 6.2–14.2.

Fig. S2 (a) Cyclic and (b) differential pulse voltammograms of **3** (black lines) and $[\text{Ru}^{\text{II}}(\text{bpy})_3]\text{Cl}_2$ (red lines) in B.-R. buffer (1.0 mM) at pH 2.72 at room temperature. Scan rate for CV: 0.1 V s^{-1} .

Fig. S3 Spectral changes of every 7.5 min during the electrochemical oxidation of a) **1**, b) **2**, and c) **3** in B.-R. buffer (pH 1.8, sample concentration: 0.5 mM) at room temperature. The left graphs indicate the changes during the reactions for the first 30 min (0–30 min) and the right ones show the changes for the latter 30 min (30–60 min). The initial, the medium and the final spectra of each complex are indicated as the blue, purple and red lines in the graphs, respectively.

Fig. S4 ESI-TOF MS spectra of **6** a) in H_2^{16}O and b) in H_2^{18}O and their computer simulations (c and d).

Fig. S5 ESI-TOF MS spectra of **6** a) in H_2O and b) in $H_2^{18}O$. The asterisk denotes non-assignable Ru-species and the crosses (x) indicate signals due to organic species.

Fig. S6 ESI-MS spectra of electrochemically generated (a) **4**, (b) **5** and (c) **6** in H₂O (left, and right above) and their computer simulation (right below). *: non-assignable Ru-species, x: organic species.

Fig. S7 Resonance Raman spectra of **6** generated in H₂¹⁶O (801 cm⁻¹, red line) and in H₂¹⁸O (761 cm⁻¹, blue line), and the differential spectrum: (black) = (red) – (blue); **3** (2 mmol) and CAN (20 mmol) in water (70 μL).

Fig. S8 Spectral changes during the chemical oxidation of **3** (sample concentration: 0.05 mM) a) in neutral water and b) in B.-R. buffer (pH 1.8) at room temperature. The initial spectrum of each graph is indicated as the blue red lines, respectively. The purple and red lines in each graph indicate the spectra at the stages upon addition of 5 eq and 10 eq of CAN for graph a, and those upon addition of 1 eq and 2 eq for graph b, respectively.

Fig. S9 ^1H ^1H COSY spectrum of complex **6** in D_2O at room temperature.

Fig. S10 1D NOE measurements on complex **6** in D₂O at room temperature with irradiation at 7.4 ppm: The spectrum a) without irradiation; b) with irradiation at 7.4 ppm; c) the differential spectrum (a – b).

Fig. S11 ^1H NMR spectra in D_2O of catalytic oxidations of a) 4-methylbenzyl alcohol, b) 1-propanol, c) 2-propanol, d) sodium 4-ethylbenzene-sulfonate as substrates in the presence of **1** as a catalyst and CAN as the oxidant. Arrows indicate the signals of each product. *: DSS as an internal reference, x: ammonium ion of CAN.

Fig. S12 ^1H NMR spectra of the control experiments without catalysts for the oxidation of a) 4-methylbenzyl alcohol, b) 1-propanol, c) 2-propanol, d) sodium 4-ethylbenzene-sulfonate as substrates in the presence of CAN as the oxidant. Arrows indicate the signals of each product. *: DSS as an internal reference, x: ammonium ion of CAN.

Fig. S13 Time courses of the oxidation reactions of 1-propanol with 0.025 M (blue), 0.05 M (green), 0.10 M (purple), 0.15 M (red) in the presence of complexes a) **4**, b) **5** and c) **6** as oxidants (0.5 mM) at 301 K.

Fig. S14 Pseudo first-order kinetic analysis for oxidation of 1-propanol with complexes (a) **4**, and (b) **5** as oxidants (0.5 mM) in B.-R. buffer (pH 1.8) at 308 K (red), 301 K (purple), 288 K (green), 280 K (blue).

Fig. S15 Eyring (a, b and c) and van't Hoff plots (d, e and f) for oxidation of 1-propanol with complexes **4** (a, d), **5** (b, e) and **6** (c, f) as oxidants.

Fig. S16 Pseudo-first-order kinetic analysis for oxidation of sodium 4-ethylbenzene-sulphonate with complexes (a) **4**, (b) **5** and (c) **6** as oxidants (0.5 mM) in B.-R. buffer (pH 1.8) at 295 K.

Fig. S17. Pseudo-first-order kinetic analysis for oxidation reactions of CH_3OH (blue line), CD_3OH (red line) and CH_3OD (green line) with complexes (a) **4** and (b) **5** as oxidants at 297 K. CH_3OH and the deuterated derivatives (CD_3OH and CH_3OD) were used as substrates.