1	DHEA administration activates local bioactive androgen metabolism in cancellous site of tibia
2	of ovariectomized rats
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13	Running title: DHEA and local androgen metabolism in bone tissue
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1	Abstract It is not known whether local androgen metabolism is involved in the mechanisms
2	underlying the dehydroepiandrosterone (DHEA) administration-induced improvement of bone
3	mineral density (BMD) in an estrogen deficiency state. The aim of the present study was to
4	clarify whether DHEA administration would improve local androgen metabolism and BMD in
5	cancellous site of tibia of ovariectomized (OVX) rats. Twenty-two female rats, 6 weeks old,
6	were randomized into three groups: sham-operated rats, OVX control rats, and OVX rats that
7	received DHEA treatment. DHEA was administered intraperitoneally at 20 mg DHEA/kg body
8	weight for 8 weeks. The concentrations of free testosterone and dihydrotestosterone (DHT) in
9	cancellous site of tibia did not change as a result of ovariectomy, while the DHT concentration
10	increased following DHEA administration. We revealed that DHEA administration improved the
11	reduction of 17β- and 3β-hydroxysteroid dehydrogenases and clearly reversed the reduction of
12	5α -reductase type 1 and 2 and androgen receptor in the cancellous site of tibia of OVX rats.
13	DHEA administration suppressed estrogen deficiency relative to the decrease in the cancellous
14	BMD, which was positively associated with local DHT concentration. These findings indicate
15	that DHEA administration enhances local bioactive androgen metabolism in the cancellous tibia
16	of young OVX rat, suggesting that local DHT may play a part in the DHEA
17	administration-induced improvement of cancellous BMD.

18 Key words: estrogen deficiency, BMD, DHEA, androgen metabolism

1 Introduction

 $\mathbf{2}$ Before natural menopause, primary ovarian insufficiency defined as the premature cessation of ovarian function or menstrual dysfunction due to disordered eating or the female athlete triad 3 4 syndrome is associated with severe risks for bone health [1, 2]. The ovariectomized (OVX) rat is a useful animal model for examining actions of sex steroids on bone. In young aged rats, $\mathbf{5}$ 6 ovariectomy induces the increases in both longitudinal and radial growth, as well as significant 7loss of trabecular bone [3, 4]. Although it is well known that the trabecular bone loss caused by 8 ovariectomy is attributable to estrogen deficiency, it is not clear what actions other gonadal 9 hormones have for bone metabolism in the estrogen deficiency state. 10 Dehydroepiandrosterone (DHEA), secreted mainly from the adrenal gland and ovary, 11 plays a critical physiological role for maintaining steroidogenesis by being used as the available 12precursors converted to testosterone and estrogens in the various peripheral tissues such as the bones, liver, brain, skeletal muscles, and so on [5]. DHEA concentration in the blood decreases 1314 following ovariectomy in animals [6, 7]. On the other hand, DHEA replacement improves bone mineral density (BMD) in OVX animals [6, 7]. However, the mechanisms by which DHEA 15induces the BMD increase are largely unknown. 1617In osteoblast cells (OBs), DHEA can be converted to testosterone and further metabolized to dihydrotestosterone (DHT), which has the physiological strong function [8-10] and is known 18

1	to be the most powerful androgen because its higher affinity for androgen receptor (AR) than
2	that of testosterone [11]. DHEA also increases the transcription of AR in OBs [7]. In particular,
3	DHT prevents cancellous bone loss in OVX rats [12], and AR knockout female mice have a
4	decreased amount of cancellous bone as well [13]. Thus, a decrease in circulating DHEA level
5	caused by ovariectomy could deactivate the transcription of local androgen-related steroid
6	enzymes in bone tissue, probably leading to decreased cancellous bone mass. However, it is not
7	clear whether DHEA administration would activate the local androgen-related steroid enzymes
8	or hormones in vivo during a period of rapid bone loss caused by estrogen deficiency.
9	We previously showed that estrogen deficiency reduced the cancellous BMD of tibia but
10	not the cortical BMD in OVX rats [14]. Moreover, DHEA administration apparently improved
11	the cancellous bone osteopenia in OVX rats [6]. Accordingly, we hypothesized that DHEA
12	administration would activate the local androgen-related steroidogenesis with lessening the
13	reduction of cancellous BMD of tibia caused by estrogen deficiency. In the present study, we
14	examined androgen levels of free testosterone and DHT, steroidogenesis-related enzymes of
15	17 β -hydroxysteroid dehydrogenase (17 β -HSD), 3 β -HSD, 5 α -reductase type 1 and 2, and AR,
16	and BMD in a cancellous site of the tibia in OVX rats after 8 wks of DHEA administration.
17	

18 Materials and methods

1	Experimental animals and feeding protocol. Twenty-two female Sprague-Dawley rats, 6
2	weeks old, were randomized into three groups: sham-operated rats (Sham Control; SC, n=5),
3	OVX control rats (OC, n=8), and OVX rats with DHEA treatment (OD, n=9). The OVX groups
4	were ovariectomized via the dorsal route, and the sham-operated rats were operated on without
5	the ovaries being removed. During the period of the experiment, all rats were fed a diet with
6	1.05% calcium and 1.01% phosphate purchased from CLEA Japan (CE-2, CLEA Japan, Inc.,
7	Japan). DHEA dissolved in sesame oil was administered to the OD group intraperitoneally at 20
8	mg DHEA/kg body weight for 8 weeks beginning one week after ovariectomy while the SC and
9	OC groups were treated with vehicle only (sesame oil, 0.5 ml) [15]. Rats were not treated with
10	DHEA or vehicle every fourth day (i.e., they were treated for 3 consecutive days). The rats were
11	kept in individual cages ($15 \times 25 \times 19.5$ cm) and allowed access to food and distilled water <i>ad</i>
12	libitum. Food consumption and body weight gain were measured every second day. The room
13	temperature was kept at 24±1 $^{\circ}$ C, the humidity at 50±5%. Fluorescent lights were on from 8:00
14	a.m. to 8:00 p.m Animal care and experimental procedures were approved by the Animal
15	Experimental Committee of the University of Tsukuba.

17 Measurement of bone mineral density. The left tibia of each rat was isolated by dissection and 18 freed from any muscle and connective tissue. Thereafter, bone mineral content (BMC) and bone

1	mineral density (BMD) values for the whole tibia were measured by Dual-energy X-ray
2	absorptiometry (DXA; Aloka DCS-600R instrument). We studied the proximal one-fifth of the
3	tibia, including the epimetaphyseal region representing the cancellous sites, and middle
4	one-fifth of the tibia representing the cortical diaphyseal region [16].

 $\mathbf{5}$

6 Immunoblot analysis. Western blot analysis of the cancellous site (proximal one-fifth of the 7tibia) of the right tibia of each rat was performed according to the procedure described in our previous paper with minor modifications [17]. After completely removing marrow and blood, 8 9 the tibia fragment was homogenized by grinding the tissue in a mortar on dry ice with 10 10 volumes of 20 mM Tris-HCL (pH 7.8), 2 mM EDTA, 300 mM NaCl, 2 nM DTT, 2% NP 40, 11 0.2% SDS, 0.2% sodium deoxycholate, 0.5 mM PMSF, 60 µg/ml aprotinin, and 1 µg/ml leupeptin. The homogenate was gently rotated for 30 min at 4 $^{\circ}$ C and then centrifuged at 12,000 1213g for 15 min at 4 °C. The protein concentration of the resulting supernatant was determined by 14protein assay rapid kit (Wako Pure Chemical Industries, Ltd., Osaka, Japan). Samples (50 µg protein) were subjected to heat denaturation at 96 $^{\circ}$ C for 7 min with Laemmli buffer. Western 15blot analysis for the detection of 17 β - and 3 β -HSD, 5 α -reductase type 1 and 2, and AR was 1617performed according to the procedure described in our previous report [17, 18]. Briefly, each sample was separated on an SDS-polyacrylamide gel (10 %) and then transferred to a 18

1	polyvinylidene difluoride (PVDF, Millipore, Billerica, MA, USA) membrane. The membrane
2	was then incubated in blocking buffer of 3% skimmed milk in phosphate-buffered saline
3	containing 0.1% Tween-20 (PBS-T) for 1 h at room temperature followed by incubation with
4	primary antibodies, a polyclonal anti-17β-HSD and anti-3β-HSD (1:1000 dilution, Santa Cruz
5	Biotechnology), a polyclonal anti-5 α -reductase type 1 and 2 (1:1000 dilution, Abnova
6	Corporation, Taipei, Taiwan), a polyclonal anti-AR (1:1000 dilution, Santa Cruz Biotechnology),
7	and a monoclonal anti- β -actin (1:1000 dilution, Santa Cruz Biotechnology), for 12 h at 4 $^\circ\!\mathbb{C}.$
8	Subsequently, the membrane was washed with PBS-T three times, and incubated for 1 h at room
9	temperature with a horseradish peroxidase (HRP)-conjugated secondary antibody, either an
10	anti-rabbit IgG (1:2000 dilution, Amersham Biosciences, Piscataway, NJ, USA) or an
11	anti-mouse IgG (1:2000 dilution, Amersham Biosciences) After this reaction, the membrane
12	was washed with PBS-T five times. Finally, binding was detected by chemiluminescence with
13	the ECL Plus system (Amersham Life Sciences), and the membrane was exposed to Hyper film
14	(Amersham Biosciences).

16 Sandwich-enzyme immunoassay (EIA). Concentrations of E_2 and DHEA in serum and free 17 testosterone and DHT in bone tissue were determined using a sandwich EIA kit (free 18 testosterone: IBL International GMBH, Flughafenstrasse, Hamburg, Germany; E_2 : Cayman

1	Chemical, Ann Arbor, MI, U.S.A.; DHEA: Assay Designs, Inc., Ann Arbor, MI, U.S.A.; DHT:
2	IBL International GMBH, Flughafenstrasse, Hamburg, Germany). The protein samples of bone
3	tissue for the EIA analysis were the same as those for the western blot analysis. All techniques
4	and materials used in this analysis were in accordance with the manufacturers' protocols. The
5	sensitivities of intra- and inter-assay coefficients of variation for the hormone assays were as
6	follows: at dose of free testosterone (62 pg/ml for intra-assay precision; 76 pg/ml for inter-assay
7	precision), 4.7% and 9.3%, respectively; at dose of E_2 (102 pg/ml for intra- and inter-assay
8	precision), 13.0% and 8.2%, respectively; at dose of DHEA (1039 pg/ml for intra-assay
9	precision; 1093 pg/ml for inter-assay precision), 5.8% and 6.5%, respectively; at dose of DHT
10	(237 pg/ml for intra-assay precision; 281 pg/ml for inter-assay precision), 11.4% and 12.1%,
11	respectively.
11 12	respectively.
	respectively. Statistics. All the data are expressed as mean \pm SE. Statistical analysis was carried out by
12	
12 13	Statistics. All the data are expressed as mean \pm SE. Statistical analysis was carried out by
12 13 14	Statistics. All the data are expressed as mean \pm SE. Statistical analysis was carried out by analysis of variance (ANOVA) followed by Fisher's F-test for multiple comparisons. A
12 13 14 15	Statistics. All the data are expressed as mean \pm SE. Statistical analysis was carried out by analysis of variance (ANOVA) followed by Fisher's F-test for multiple comparisons. A significance level of $p < 0.05$ was used for all comparisons. Association between BMD and

<The position of Table 1>

2 **Results**

3	Initial body weight did not differ among the three groups. The final body weight, body
4	weight gain, and food intake of the OVX groups were significantly higher than those of the SC
5	group (Table 1). The final body weight and body weight gain of the OD group were
6	significantly lower than those of the OC group, but there were no differences in food intake. The
7	serum concentration of DHEA and E_2 in the OC group was found to be significantly lower
8	compared to that of the SC group and significantly higher in the OD group than in the OC group
9	(Table 1). There were no differences in serum free testosterone concentration between the SC
10	and OC groups, but the concentration was significantly higher in the OD group than in the OC
11	group.
12	<i><the 1="" figure="" of="" position=""></the></i>
13	The BMD of the tibial proximal metaphysis (cancellous site) in the OC group was found

15 The Diric of the usual proximal metaphysis (cancendous site) in the OC group was round 14 to be significantly lower compared to that in the SC group, but this decrease was significantly 15 attenuated in the OD group. On the other hand, there were no differences among the 3 groups in 16 the BMD of the tibial diaphysis (cortical-abundant region) (Fig. 1). 17 *< The position of Figure 2>* 18 In the tibial proximal metaphysis, free testosterone concentration did not differ among the

19 3 groups (Fig. 2). On the other hand, local DHT concentration was significantly higher in the

1 OD group than in the OC group, whereas there was no difference between the SC and OC 2 groups (Fig. 2).

3	The local concentration of DHT was also significantly correlated with the BMD of the tibial
4	proximal metaphysis in the OVX groups ($r = 0.706$, $p < 0.05$). The local DHT concentration
5	was significantly associated with the serum concentration of DHEA ($r = 0.936$, $p < 0.001$), free
6	testosterone ($r = 0.933$, $p < 0.001$), or E ₂ ($r = 0.817$, $p < 0.01$) in the OVX groups. Also, there
7	was a significant correlation between the BMD of tibial proximal metaphysis and the serum
8	concentration of DHEA ($r = 0.741$, $p < 0.01$), free testosterone ($r = 0.754$, $p < 0.01$), or E ₂ ($r =$
9	0.685, $p < 0.05$) in the OVX groups. In the correlation analysis, we used only 11 samples (n=6
10	for the OC group and n=5 for the OD group) because of hemolysis in blood samples.
11	The protein expressions of 17 β - and 3 β -HSD, 5 α -reductase type 1 and 2, and AR were
12	significantly lower in the OC group than in the SC group and significantly higher in the OD
13	group than in the OC group (Fig. 3).
14	<the 3="" figure="" of="" position=""></the>
15	Discussion
16	The present study results confirmed that DHEA administration improved the decrease in
17	the cancellous BMD of tibia caused by estrogen deficiency. The present study demonstrated that
18	DHEA administration increased the local DHT concentration in the cancellous site of tibia of

1	OVX rats. We also revealed that DHEA administration improved the reduction of 17 β - and
2	3 β -hydroxysteroid dehydrogenases and clearly reversed the reduction of 5 α -reductase type 1
3	and 2 and androgen receptor in the cancellous site of tibia of OVX rats. Moreover, the
4	cancellous BMD of tibia was positively correlated with the local DHT concentration in OVX
5	rats. These results suggest that local bioactive androgen metabolism plays a role, at least in part,
6	in the mechanism underlying DHEA administration-induced improvement of cancellous bone
7	loss caused by estrogen deficiency.
8	The present study demonstrated that estrogen deficiency clearly reduced the activation of
9	the androgen-related steroidogenic enzymes. An increase in DHEA enhances the transcriptions
10	of the androgen-related steroidogenic enzymes in OBs [7-9]. We previously also demonstrated
11	that androgen-related hormones could be locally synthesized by improving the transcriptions of
12	the androgen-related steroidogenic enzymes from DHEA in muscle tissues as well [17]. Thus,
13	we assumed in the present study that the decrease in circulating DHEA level by OVX might
14	have partly deactivated the transcriptions of the androgen-related steroidogenic enzymes in the
15	bone tissue. However, it is unclear whether estrogen deficiency reduced the activation of the
16	androgen-related steroidogenic enzymes through DHEA, directly or indirectly or both.
17	In vitro studies using OBs demonstrated that DHEA can be metabolized to testosterone by
18	17 β - and 3 β -HSD and further metabolized to DHT by the 5 α -reducatase type 1 and 2 enzymes,

1	binding to the AR [7-9]. These results mostly corresponded to the results of the present study in
2	a cancellous site of the tibia in vivo that DHEA administration increased the DHT concentration
3	and the steroidogenesis-related enzymes in 17 β - and 3 β -HSD, the 5 α -reducatase type 1 and 2,
4	and AR. However, in the present study, DHEA administration did not induce any increase in the
5	local free testosterone concentration. Free testosterone is metabolized to DHT only through the
6	5α -reductase enzymes [11, 19]. Actually, in the present study, the expression of 5α -reductase
7	type 1 and 2 was greatly increased to the level of sham-operated rats. Moreover, testosterone
8	can be metabolized not only to DHT but E_2 by p450arom enzyme activity [7]. Thus, the
9	transient increase in testosterone might have occurred, but the testosterone was soon
10	metabolized to synthesize DHT and/or E_{2} .
10 11	metabolized to synthesize DHT and/or E_{2} . The present study results showed that DHEA administration clearly reversed the reduction
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11 12	The present study results showed that DHEA administration clearly reversed the reduction of AR induced by estrogen deficiency as well. Wang et al. [7] reported that DHEA increases the
11 12 13	The present study results showed that DHEA administration clearly reversed the reduction of AR induced by estrogen deficiency as well. Wang et al. [7] reported that DHEA increases the transcription of AR in OBs. AR knockout mice have a decreased amount of cancellous bone
11 12 13 14	The present study results showed that DHEA administration clearly reversed the reduction of AR induced by estrogen deficiency as well. Wang et al. [7] reported that DHEA increases the transcription of AR in OBs. AR knockout mice have a decreased amount of cancellous bone [13]. DHT also has the capacity to prevent ovariectomy-induced cancellous bone loss [12]. On
 11 12 13 14 15 	The present study results showed that DHEA administration clearly reversed the reduction of AR induced by estrogen deficiency as well. Wang et al. [7] reported that DHEA increases the transcription of AR in OBs. AR knockout mice have a decreased amount of cancellous bone [13]. DHT also has the capacity to prevent ovariectomy-induced cancellous bone loss [12]. On the other hand, the inhibitor of 5α -reductase does not appear to have a bone-sparing effect [20].

1	The present study showed the significant correlation between the BMD and local DHT
2	concentration in the site of cancellous bone loss caused by estrogen deficiency. Especially, the
3	marked increase in serum free testosterone concentration by DHEA administration was found
4	and the serum free testosterone concentration was highly correlated with the local DHT
5	concentration. Thus, we cautiously considered that the increase in the serum free
6	testosterone concentration by DHEA administration may, at least in part, induce the
7	increase in local DHT level, resulting in the increase in the cancellous BMD. On the
8	other hand, the present study also demonstrated that DHEA administration resulted in a
9	substantial rise in serum E_2 concentration which was also positively correlated with the
10	cancellous BMD of tibia. The increase in serum E_2 concentration prevents severe bone loss
11	caused by estrogen deficiency [7]. Thus, it could not be excluded that the increase in serum E_2
12	concentration by DHEA administration would have a sufficient bone sparing effect irrespective
13	of the local DHT level. Therefore, to evaluate if parts of the DHEA effect was dependent on
14	local DHT level, a further study is strongly needed to demonstrate if the DHEA effect would be
15	reduced by co-treatment with androgen receptor antagonist or if the DHEA effect would be
16	sustained by co-treatment with estrogen receptors or aromatase antagonist.
17	Although the present study results confirmed that DHEA prevented bone loss caused by
18	estrogen deficiency and improved local bioactive androgen metabolism, DHEA concentration in

1	serum was approximately 13 times higher in OVX rats treated with DHEA than in
2	sham-operated control rats. Future study should focus on the dose-dependent effect of DHEA
3	administration on the relation between BMD and local bioactive androgen metabolism.
4	Furthermore, the present study results confirmed that the estrogen deficiency reduced the
5	cancellous BMD but not the cortical BMD as reported previously [14]. Moreover, DHEA
6	administration prevented the cancellous BMD loss but did not affect the cortical BMD.
7	However, the BMD measurement by DXA in the present study did not allow the details of bone
8	parameters such as cortical width and periosteal or endosteal circumferences. Thus, the
9	parameters should be examined in detail using by the methods of pQCT, μ CT, or bone
10	morphometry in further studies.
10 11	morphometry in further studies. In conclusion, the present study results demonstrated for the first time that in a cancellous
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11 12	In conclusion, the present study results demonstrated for the first time that in a cancellous site of the tibia, DHEA administration increased the DHT concentration and improved the
11 12 13	In conclusion, the present study results demonstrated for the first time that in a cancellous site of the tibia, DHEA administration increased the DHT concentration and improved the reduction of the androgen steroidogenesis-related enzymes caused by estrogen deficiency.
11 12 13 14	In conclusion, the present study results demonstrated for the first time that in a cancellous site of the tibia, DHEA administration increased the DHT concentration and improved the reduction of the androgen steroidogenesis-related enzymes caused by estrogen deficiency. Furthermore, DHEA administration suppressed the decrease in the cancellous BMD of tibia
 11 12 13 14 15 	In conclusion, the present study results demonstrated for the first time that in a cancellous site of the tibia, DHEA administration increased the DHT concentration and improved the reduction of the androgen steroidogenesis-related enzymes caused by estrogen deficiency. Furthermore, DHEA administration suppressed the decrease in the cancellous BMD of tibia induced by estrogen deficiency, which was significantly associated with the local DHT

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5	wo	ork. The authors have no relevant financial interest in this article (DISCLOSURES: NONE).
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1 Figure legends

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Figure 1. Bone mineral density of tibial proximal metaphysis (A) and diaphysis (B).
The proximal one-fifth of the tibia, including the epimetaphyseal region representing the
cancellous sites, and middle one-fifth of the tibia, representing the cortical diaphyseal
region. SC, Sham-operated control group; OC, Ovariectomy-operated control group;
OD, Ovariectomy group with DHEA treatment. *** p < 0.001 vs. SC; ## p < 0.01 vs.
OC. Values are expressed as the means ± SE.
Figure 2. Concentrations of free testosterone (A) and DHT (B) in the proximal

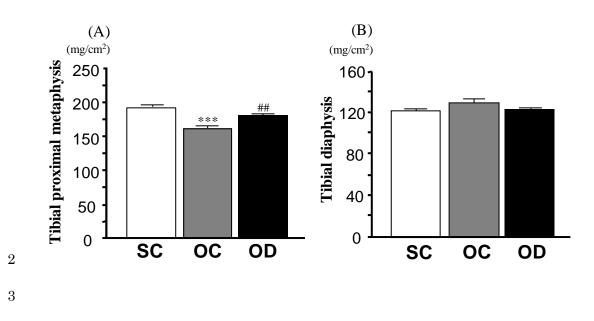
metaphysis of the tibia (trabecular bone). The tissue concentrations were normalized for total protein. SC, Sham-operated control group; OC, Ovariectomy-operated control group; OD, Ovariectomy group with DHEA treatment. ^{###} p < 0.001 vs. OC. Values are expressed as the means \pm SE.

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Figure 3. Concentrations of 17 β -HSD protein (A), 3 β -HSD protein (B), 5 α -reductase type 1 protein (C), 5 α -reductase type 2 protein (D), and androgen receptor protein (E) in the proximal metaphysis of the tibia (trabecular bone). The tissue concentrations were

1	normalized for	or total	protein.	SC,	Sham-opera	nted	control	group;	OC,
2	Ovariectomy-op	perated con	trol group;	OD,	Ovariectomy	group	with DI	HEA treat	ment.
3	*** p < 0.001 v	vs. SC; ### p	< 0.001 vs.	OC.	Values are exp	oressed	l as the m	neans ± SE	
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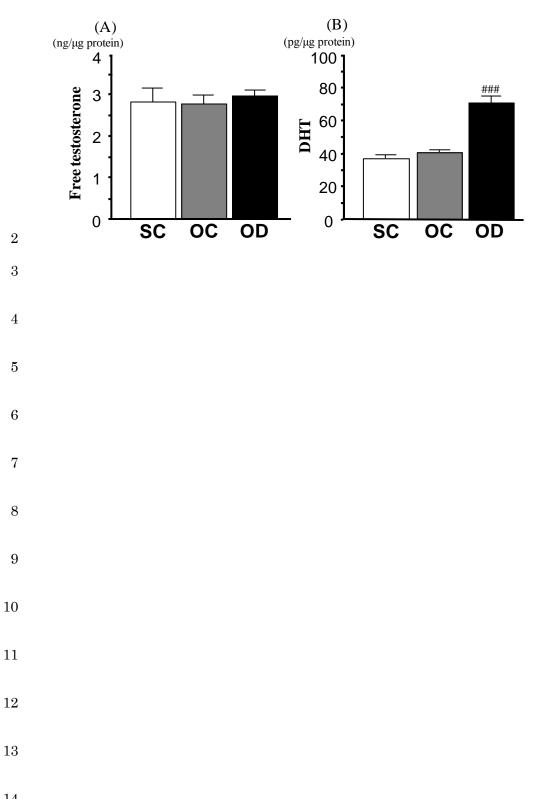
1 Figure 1.



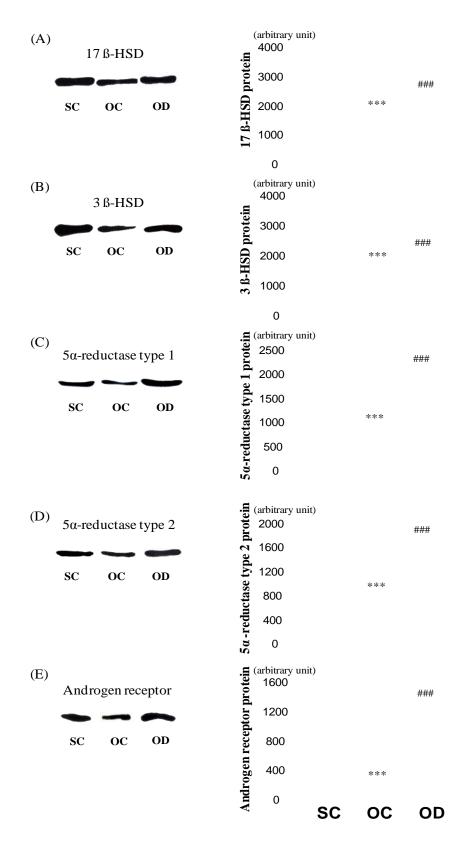
 $\mathbf{5}$

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1 Figure 2.



1 Figure 3.



 $\mathbf{2}$

Table 1. Body weight, food intake, and steroid hormones in serum

	-						
	Final body weight I	Body weight gain	Food intake	DHEA	E ₂	Free testosterone	
	(g)	(g/day)	(g/day)	(ng/ml)	(pg/ml)	(pg/ml)	
SC	274.8 ± 5.8	2.0 ± 0.1	15.4 ± 0.4	2.7 ± 0.2	35.8 ± 5.9	2.9 ± 0.2	
OC	$340.5 \pm 6.3^{***}$	$3.2 \pm 0.1^{***}$	$17.7 \pm 0.2^{***}$	$1.1 \pm 0.2^{**}$	$13.4 \pm 1.3^*$	1.2 ± 0.1	
OD	$309.1 \pm 6.8^{\#}$	$2.6 \pm 0.1^{\#}$	17.4 ± 0.5	$35.0 \pm 0.6^{\#\#}$	$53.1 \pm 10.0^{\#}$	$^{\#\#}$ 71.7 ± 3.1 $^{\#\#\#}$	

Data show mean ±SE. SC, Sham-operated control group; OC, Ovariectomy-operated control group; OD, Ovariectomy-operated

group with DHEA treatment. * p < 0.05, ** p < 0.01, *** p < 0.001 vs. SC; #p < 0.05, ##p < 0.01, ###p < 0.001 vs. OC.