

Threading/Folding Recognition Modes of Phosphodiester by a *p*-Nitrophenylamide Cyclodextrin Derivative

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A cyclodextrin derivative **1** possessing multiple *p*-nitrophenylamide groups, which is a strong hydrogen-bond donor, encapsulates phosphodiester anions in two recognition modes. One is the ‘threading’ mode, in which the phosphodiester passes through the cyclodextrin cavity. The other is the ‘folding’ mode, in which the included phosphodiester is bent inside the cyclodextrin cavity. The tendency between the two recognition modes depends on the substituents on the phosphodiester.

Keywords: Amide, Cyclodextrin, Anion receptor

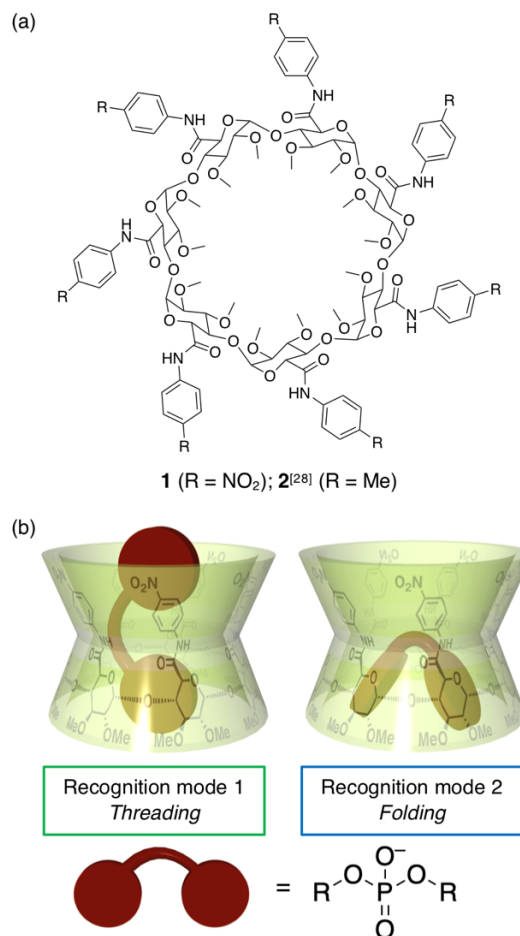
The precise molecular recognition by proteins is realized by multiple hydrogen bonds utilizing amino acid side chains and backbone amide groups which are accumulated inside the recognition pocket.^[1–5] Inspired by nature, many artificial receptors whose interaction moieties are closely arranged to each other have been reported.^[6–13] In particular, amide groups have been widely used for such receptors, because they can be easily introduced and ubiquitously found in biological molecules.^[14–21]

Fixing the position, orientation, and conformation of the encapsulated molecules via multiple non-covalent bonds is important for the precise recognition and catalytic functions.^[22–24] Phosphodiester groups are found ubiquitously in biological molecules such as DNA and phospholipids. To understand and mimic the functions of phosphodiesterases, the artificial host molecules that recognize or hydrolyze diaryl phosphates as model compounds have been investigated.^[25–27] Host molecules that recognize a phosphodiester in a specific conformation would contribute to the development of selective phosphate transporters and artificial enzymes.

We have previously reported a cyclodextrin derivative **2** whose amide groups are directly attached to the pyranose units.^[28] **2** possesses seven *p*-tolylamide groups. Although they are in close proximity, they formed only partial intramolecular hydrogen bonds to each other because of the conformational restriction by the cyclodextrin framework and the steric hindrance of the accumulated aromatic rings. This contributes to the effective intermolecular hydrogen bonds between the amides and the substrate included in the cavity. It is notable that the selective recognition of hydrogen phosphonate anions has been achieved by utilizing chemically equivalent amide groups both as a hydrogen-bond donor and acceptor.

It is known that the introduction of electron-withdrawing groups on aromatic amide or urea units leads to a greater superior hydrogen-bonding ability by lowering the electron density of the N-H bond.^[6,15] We have now

synthesized a novel cyclodextrin derivative **1** possessing seven *p*-nitrophenyl groups with the objective of producing a unique recognition ability for anions (Figure 1a). It was revealed that the receptor **1** has a double-conical-shaped recognition pocket consisting of two distinct cavities on each side of the central amide moieties. The ‘first’ one is the normal cyclodextrin’s cavity surrounded by the pyranose units. The ‘second’ one is on its other side, and is surrounded by the electron-deficient aromatic rings. With this feature, the receptor **1** exhibited two unique recognition modes for the inclusion of phosphodiester with different conformations, that is, a ‘threading’ mode and a ‘folding’ mode (Figure 1b).



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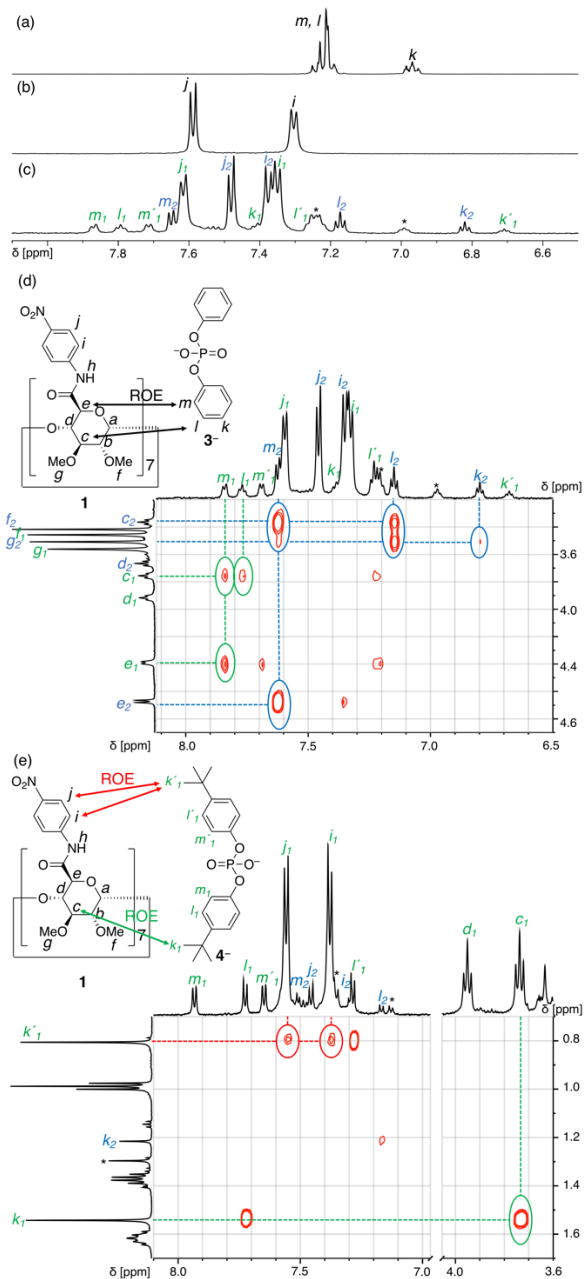
Figure 1. (a) Structures of aromatic-amide cyclodextrin derivatives. (b) Schematic representation of the two recognition modes of phosphodiester by **1**.

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1 Receptor **1** was synthesized by the condensation
 2 reaction of a per(5-carboxy-5-dehydroxymethyl)- β -
 3 cyclodextrin derivative^[29] and *p*-nitroaniline (Scheme S1,
 4 Figures S1–S7). A ¹H NMR signal of the amide proton of **1**
 5 was observed at 9.00 ppm (CD₃CN), which is a good
 6 indicator for its participation in the hydrogen bonding.

7 We first briefly investigated whether **1** exhibits a
 8 stronger binding of anions than **2**. ¹H NMR titration
 9 experiments confirmed that **1** (0.80 mM) quantitatively
 10 encapsulated *p*-toluenesulfonate, acetate, and chloride in a
 11 1:1 manner ($K_a > 10^4$ M⁻¹, CDCl₃) (Figures S8–S10), which
 12 is in contrast to the weak or no binding of the three anions by
 13 **2** ($K_a < 10^2$ M⁻¹)^[28]. This can be attributed to the superior
 14 hydrogen-bonding ability of receptor **1**.

15 The interaction between the diphenyl phosphate anion
 16 (**3**⁻) and **1** was then investigated. The diphenyl phosphate
 17 anion showed no binding toward **2** in a previous study.^[28] A
 18 ¹H NMR titration experiment of tetrabutylammonium
 19 diphenyl phosphate ((*n*-Bu₄N)**3**) against **1** in a CD₃CN
 20 solution (0.80 mM) was performed (Figure 2a–c, Figure S11).
 21 Upon the addition of 1 equivalent of (*n*-Bu₄N)**3**, the signals
 22 of the free host molecule **1** disappeared, and two new sets of
 23 signals of the host-guest complexes were observed. The ¹H
 24 signals of the amide N-H were shifted from 9.00 to 9.98 ppm
 25 and 10.00 ppm. This downfield shift indicated the formation
 26 of hydrogen bonds between the amides and the phosphate.
 27 During the course of the titration experiment, the ratio of
 28 these two complexes was constant (58%/42%) irrespective of
 29 the amount of **3**⁻. The integration ratio of the ¹H signals **1** and
 30 **3**⁻ revealed that both species are 1:1 complexes. For the
 31 major complex (58 %), two phenyl groups of **3**⁻ were
 32 separately observed by ¹H NMR, and ROE correlations to the
 33 pyranose unit of the cyclodextrin were observed for only one
 34 of the two phenyls (Figure 2d, Figure S24). For the minor
 35 complex (42%), two phenyl groups are chemically equivalent
 36 in the ¹H NMR spectrum, and they showed ROE correlations
 37 to the sugar unit. These results suggest that the major species
 38 (58%) is a complex in which the guest is threaded through the
 39 cyclodextrin cavity (Recognition mode 1 in Figure 1b), while
 40 the minor species (42%) is a complex in which the guest is
 41 folded in the cyclodextrin's cavity (Recognition mode 2 in
 42 Figure 1b). Chemical shifts of the ³¹P NMR signals of the
 43 two complexes are different by more than 5 ppm (−5.61 and
 44 −10.68 ppm, respectively), which also supports the two
 45 distinctive recognition modes of the phosphate (Figure S23).
 46 The two species are separately observed by ¹H NMR even at
 47 high temperature (343 K), which shows a slow conversion
 48 and high energy barrier between these complexes (Figure
 49 S26).




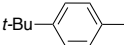
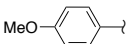
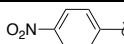
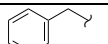
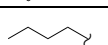
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51 **Figure 2.** Recognition of phosphodiester by **1**. (a)–(c) ¹H NMR spectra
 52 of aromatic region (600 MHz, CD₃CN). (a) (*n*-Bu₄N)(PhO-PO₂⁻OPh)
 53 ((*n*-Bu₄N)**3**). (b) **1** (0.8 mM). (c) **1** and (*n*-Bu₄N)**3** (1.3 eq). Protons of
 54 the host-guest complex in the ‘threading’ mode are denoted by green
 55 letters with subscript 1. Protons of the phenyl group of **3**⁻ in the
 56 nitrophenyl group’s cavity are denoted with primed letters, and those in
 57 the cyclodextrin’s cavity are denoted with letters without prime. Protons
 58 of the host-guest complex in the ‘folding’ mode are denoted by blue
 59 letters with subscript 2. * indicates **3**⁻ that is not encapsulated in **1**. (d)
 60 and (e) ¹H–¹H ROESY spectra (600 MHz, CD₃CN). (d) ROE
 61 correlations between aromatic regions and sugar regions for the host-
 62 guest complex between **1** and (*n*-Bu₄N)**3** (1.3 eq). (e) ROE correlations
 63 with *tert*-butyl group for the host-guest complex between **1** and (*n*-
 64 Bu₄N)(*p*-*t*-BuC₆H₄O-PO₂⁻O(*p*-*t*-BuC₆H₄)) ((*n*-Bu₄N)**4**) (1.3 eq). Red
 65 circles represent ROEs between *p*-nitrophenyl group of **1** and *t*-Bu group

of **4**⁻, and a green circle represents that between the sugar moiety of **1** and *t*-Bu group of **4**⁻.

Table 1 summarizes the ratio of the two recognition modes for the various phosphodiester, together with the 1:1 binding constants K_a [M⁻¹] (the stoichiometry has been confirmed by the integral ratio of ¹H NMR signals of the inclusion complexes; see also Figures S11–S21). For **4**⁻, which has a bulky *t*-Bu group at the *p*-position of the benzene ring, the equilibrium was biased toward the ‘threading’ mode (‘threading’/‘folding’ = 87%/13%). Steric hindrance of the *t*-Bu group would have destabilized the folded conformation of **4**⁻ inside the cyclodextrin’s cavity. In the ‘threading’ mode, a ¹H NMR signal assigned to one of the *t*-Bu groups was shifted downfield (from 1.29 ppm to 0.81 ppm) as the result of the shielding effects from the surrounding *p*-nitrophenyl groups. The structure of the ‘threading’ mode was further supported by the ROE correlation between the *t*-Bu group of **4**⁻ and the nitrophenyl groups of receptor **1** (Figure 2e, Figure S25). For **5**⁻, to which a methoxy group is introduced at the *p*-position of the benzene ring, the recognition mode was also biased toward the ‘threading’ mode (72%/28%). Meanwhile, the ‘folding’ mode was slightly favored in the case of the electron-deficient *p*-nitrophenyl phosphodiester **6**⁻ (47%/53%). The comparison of **3**⁻, **5**⁻, and **6**⁻ suggested that the electron-rich substituent on the phosphodiester might have stabilized the ‘threading’ mode through the intermolecular interactions with the electron-deficient *p*-nitrophenyl groups of **1**. Interestingly, for the non-phenolic derivatives, dibenzyl phosphate **7** and dibutyl phosphate **8**⁻, the recognition modes were biased toward the ‘folding’ mode (41%/59%, 10%/90%, respectively). These results can be explained as the result of the easiness of the flexible diphosphate molecules to take a folded conformation in the cyclodextrin’s cavity. The binding constants K_a of **1** and all of the phosphodiester were high (log K_a ~ 4–6), among which the highest affinity was observed for the dibutyl phosphate **8**⁻ (log K_a = 6.2).

Table 1. Effect of substituents on the ratio of two recognition modes and binding constants K_a [M⁻¹] between **1** and phosphodiester RO-PO₂⁻-OR (CH₃CN or CD₃CN, 298 K).

Guest anions ^a	Substituents (R)	Recognition mode 1 ^b (Threading)	Recognition mode 2 ^b (Folding)	Binding constant ^c log K_a
3 ⁻		58%	42%	4.8
4 ⁻		87%	13%	4.5
5 ⁻		72%	28%	5.4
6 ⁻		47%	53%	4.4
7 ⁻		41%	59%	4.6
8 ⁻		10%	90%	6.2

^a *n*-Bu₄N⁺ salts except for **6**⁻. **6**⁻ was used as an *i*-Pr₂EtNH⁺ salt prepared by mixing the corresponding free phosphoric acid and 1.5 equivalents of *i*-Pr₂EtN. ^b Determined by ¹H NMR measurements. ^c Determined by UV-vis titration experiments except for **6**⁻, which was determined by ¹H NMR.

Regardless of the employed phosphodiester (**3**⁻–**8**⁻), the ¹H NMR signals of *a* and *e* (1 and 5 positions of pyranose) were observed at 5.14–5.21 ppm and 4.42–4.63 ppm for the ‘threading’ mode, respectively (Figure S22). They appeared in a different region for the ‘folding’ mode, that is, 5.03–5.08 ppm for proton *a* and 4.61–4.77 ppm for proton *e*. A similar trend was also observed for the ³¹P NMR signals of **3**–**5**⁻ included within **1**. The signals were observed at –5.61 – –4.90 ppm for the ‘threading’ mode, but at –10.68 – –9.78 ppm for the ‘folding’ mode (Figure S23). These regularities indicated that the structures of both the ‘threading’ and ‘folding’ modes are fixed to some extent irrespective of the substituents on the phosphodiester.

Figure 3 shows the calculated structures of receptor **1** encapsulating diphenyl phosphate **3**⁻ for each recognition mode (semi-empirical calculation (PM6)). In the ‘threading’ mode, one phenyl group of **3**⁻ is included in the cyclodextrin’s ‘first’ cavity, while the other one is placed in the ‘second’ cavity surrounded by *p*-nitrophenyl groups (Figure 3a). The calculated electrostatic potential map of **1** in the conformation of the ‘threading’ mode suggests that many *p*-nitrophenyl groups are in the standing conformation to allow the inclusion of the guest, and the ‘second’ cavity on the electron-deficient *p*-nitrophenyl side has a partial positive charge (Figure 3c). Thus, the model supports that receptor **1** has two distinctive cavities on both sides of the amide groups, and that the recognition pocket of **1** is regarded to have a double-conical shape. The *para*-proton of the phenyl group of **3**⁻ at the ‘second’ pocket (*k*₁ in Figure 2c) is close to the *p*-nitrophenyl rings of **1**, which is consistent with the upfield shift of the corresponding NMR signal. In the ‘folding’ mode, two phenyl groups are placed adjacent to each other in the cyclodextrin’s cavity (Figure 3b). This bent conformation of **3**⁻ is consistent with the downfield shift of the NMR signal of the *ortho*-proton of **3**⁻ (*o*₂ in Figure 2c), which can be explained by the deshielding effect from the opposite phenyl group. The calculated electrostatic potential map of **1** for the ‘folding’ mode suggests that the *p*-nitrophenyl groups filled the upper space, and some tilted *p*-nitrophenyl groups constituted the positively-charged ceiling of the cyclodextrin’s cavity (Figure 3d). Phosphate groups formed multiple hydrogen bonds with the amides in the cavity for both structures. Thus, interesting inclusion modes of the phosphodiester are observed utilizing the uniquely-restricted aromatic-amide cyclodextrin framework.

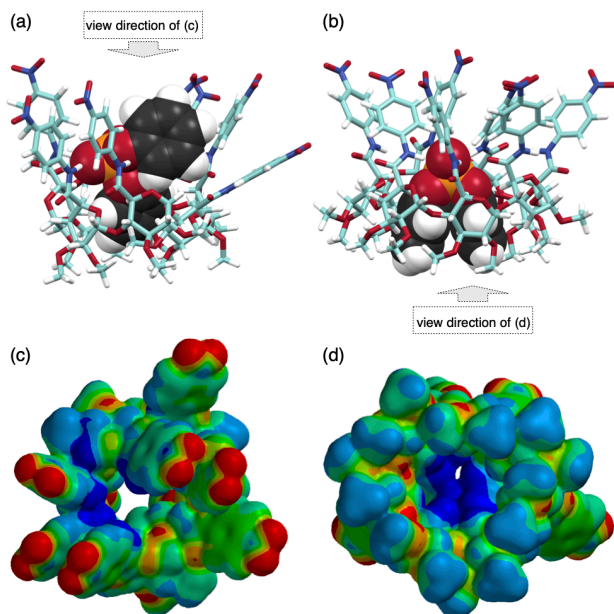


Figure 3. Structures of the two recognition modes of diphenyl phosphate (3^-) by **1** obtained by semi-empirical calculations (PM6). (a) and (c) 'Threading' mode. (b) and (d) 'Folding' mode. (a) and (b) **1**, stick model; diphenyl phosphate, space-filling model. H, white; C in **1**, light green; C in diphenyl phosphate, black; N, blue; O, red; P, orange. (c) and (d) Calculated electrostatic potential maps of **1** for the conformation of each mode (3^- has been removed). (c) View from the top. (d) View from the bottom.

In conclusion, we have synthesized a novel cyclodextrin receptor **1** possessing seven *p*-nitrophenylamide groups that shows a strong hydrogen-bonding ability. Receptor **1** is a double-conical-shaped host molecule that has two distinct cavities, and recognizes phosphodiester in two different recognition modes, i.e., the 'threading' mode and the 'folding' mode. The equilibrium is biased toward the 'threading' mode for the guests with bulky or electron-rich substituents, while the 'folding' mode is favored for the flexible guests. Further applications of **1** to utilize the precise recognition ability of a series of phosphate anions are now being investigated.

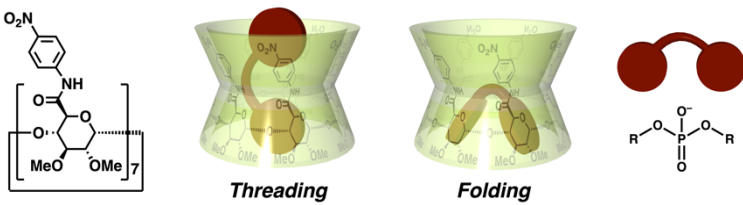
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Supporting Information is available on http://dx.doi.org/10.1246/cl.*****.

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Graphical Abstract	
Textual Information	
A brief abstract	A cyclodextrin derivative possessing multiple <i>p</i> -nitrophenylamide groups encapsulates phosphodiester anions in two recognition modes. One is the ‘threading’ mode, in which the phosphodiester passes through the cyclodextrin cavity. The other is the ‘folding’ mode, in which the included phosphodiester is bent inside the cyclodextrin cavity. The tendency between the two recognition modes depends on the substituents on the phosphodiesters.
Title	Threading/Folding Recognition Modes of Phosphodiesters by a <i>p</i> -Nitrophenylamide Cyclodextrin Derivative
Authors’ Names	Sota Yonemura, Takashi Nakamura, and Tatsuya Nabeshima
Graphical Information	
 <p>The graphical information section contains four chemical diagrams. From left to right: 1) The chemical structure of the cyclodextrin derivative, which is a cyclodextrin ring with seven units, each substituted with a <i>p</i>-nitrophenylamide group. 2) A 3D model of the 'Threading' mode, where a phosphodiester anion (represented by a red sphere) passes through the cyclodextrin cavity. 3) A 3D model of the 'Folding' mode, where the phosphodiester anion is bent and fits inside the cyclodextrin cavity. 4) The chemical structure of a phosphodiester anion, $\text{R-O-P(=O)(O}^-\text{)-O-R}$.</p>	