

Threading/Folding Recognition Modes of Phosphodiester by a *p*-Nitrophenylamide Cyclodextrin Derivative

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1 A cyclodextrin derivative **1** possessing multiple *p*-
2 nitrophenylamide groups, which is a strong hydrogen-bond
3 donor, encapsulates phosphodiester anions in two recognition
4 modes. One is the 'threading' mode, in which the
5 phosphodiester passes through the cyclodextrin cavity. The
6 other is the 'folding' mode, in which the included
7 phosphodiester is bent inside the cyclodextrin cavity. The
8 tendency between the two recognition modes depends on the
9 substituents on the phosphodiester.

10 **Keywords:** Amide, Cyclodextrin, Anion receptor

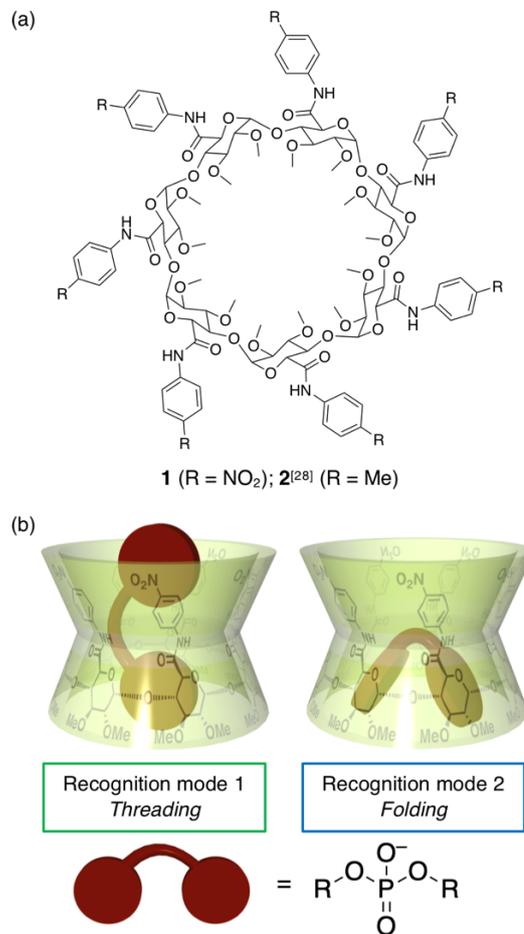
11 The precise molecular recognition by proteins is
12 realized by multiple hydrogen bonds utilizing amino acid side
13 chains and backbone amide groups which are accumulated
14 inside the recognition pocket.^[1-5] Inspired by nature, many
15 artificial receptors whose interaction moieties are closely
16 arranged to each other have been reported.^[6-13] In particular,
17 amide groups have been widely used for such receptors,
18 because they can be easily introduced and ubiquitously found
19 in biological molecules.^[14-21]

20 Fixing the position, orientation, and conformation of the
21 encapsulated molecules via multiple non-covalent bonds is
22 important for the precise recognition and catalytic
23 functions.^[22-24] Phosphodiester groups are found
24 ubiquitously in biological molecules such as DNA and
25 phospholipids. To understand and mimic the functions of
26 phosphodiesterases, the artificial host molecules that
27 recognize or hydrolyze diaryl phosphates as model
28 compounds have been investigated.^[25-27] Host molecules that
29 recognize a phosphodiester in a specific conformation would
30 contribute to the development of selective phosphate
31 transporters and artificial enzymes.

32 We have previously reported a cyclodextrin derivative
33 **2** whose amide groups are directly attached to the pyranose
34 units.^[28] **2** possesses seven *p*-tolylamide groups. Although
35 they are in close proximity, they formed only partial
36 intramolecular hydrogen bonds to each other because of the
37 conformational restriction by the cyclodextrin framework
38 and the steric hindrance of the accumulated aromatic rings.
39 This contributes to the effective intermolecular hydrogen
40 bonds between the amides and the substrate included in the
41 cavity. It is notable that the selective recognition of hydrogen
42 phosphonate anions has been achieved by utilizing
43 chemically equivalent amide groups both as a hydrogen-bond
44 donor and acceptor.

45 It is known that the introduction of electron-
46 withdrawing groups on aromatic amide or urea units leads to
47 a greater superior hydrogen-bonding ability by lowering the
48 electron density of the N-H bond.^[6,15] We have now

49 synthesized a novel cyclodextrin derivative **1** possessing
50 seven *p*-nitrophenyl groups with the objective of producing a
51 unique recognition ability for anions (Figure 1a). It was
52 revealed that the receptor **1** has a double-conical-shaped
53 recognition pocket consisting of two distinct cavities on each
54 side of the central amide moieties. The 'first' one is the
55 normal cyclodextrin's cavity surrounded by the pyranose
56 units. The 'second' one is on its other side, and is surrounded
57 by the electron-deficient aromatic rings. With this feature,
58 the receptor **1** exhibited two unique recognition modes for the
59 inclusion of phosphodiester with different conformations,
60 that is, a 'threading' mode and a 'folding' mode (Figure 1b).



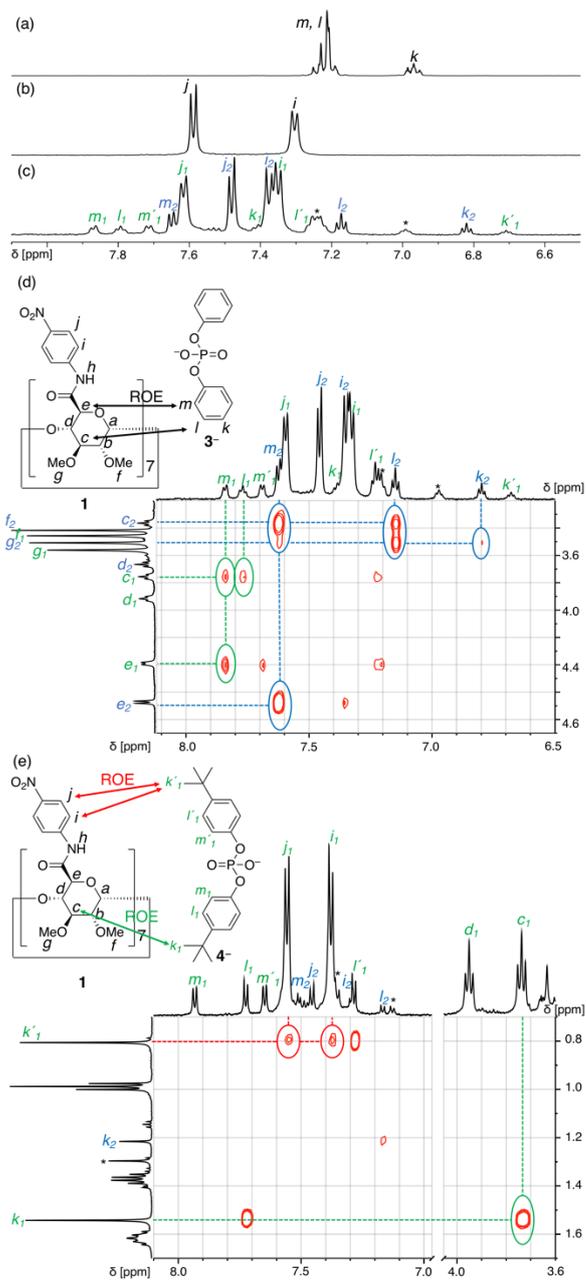
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62 **Figure 1.** (a) Structures of aromatic-amide cyclodextrin derivatives.
63 (b) Schematic representation of the two recognition modes of
64 phosphodiester by **1**.

1 Receptor **1** was synthesized by the condensation
 2 reaction of a per(5-carboxy-5-dehydroxymethyl)- β -
 3 cyclodextrin derivative^[29] and *p*-nitroaniline (Scheme S1,
 4 Figures S1–S7). A ¹H NMR signal of the amide proton of **1**
 5 was observed at 9.00 ppm (CD₃CN), which is a good
 6 indicator for its participation in the hydrogen bonding.

7 We first briefly investigated whether **1** exhibits a
 8 stronger binding of anions than **2**. ¹H NMR titration
 9 experiments confirmed that **1** (0.80 mM) quantitatively
 10 encapsulated *p*-toluenesulfonate, acetate, and chloride in a
 11 1:1 manner ($K_a > 10^4 \text{ M}^{-1}$, CDCl₃) (Figures S8–S10), which
 12 is in contrast to the weak or no binding of the three anions by
 13 **2** ($K_a < 10^2 \text{ M}^{-1}$)^[28]. This can be attributed to the superior
 14 hydrogen-bonding ability of receptor **1**.

15 The interaction between the diphenyl phosphate anion
 16 (**3⁻**) and **1** was then investigated. The diphenyl phosphate
 17 anion showed no binding toward **2** in a previous study.^[28] A
 18 ¹H NMR titration experiment of tetrabutylammonium
 19 diphenyl phosphate ((*n*-Bu₄N)**3**) against **1** in a CD₃CN
 20 solution (0.80 mM) was performed (Figure 2a–c, Figure S11).
 21 Upon the addition of 1 equivalent of (*n*-Bu₄N)**3**, the signals
 22 of the free host molecule **1** disappeared, and two new sets of
 23 signals of the host-guest complexes were observed. The ¹H
 24 signals of the amide N-H were shifted from 9.00 to 9.98 ppm
 25 and 10.00 ppm. This downfield shift indicated the formation
 26 of hydrogen bonds between the amides and the phosphate.
 27 During the course of the titration experiment, the ratio of
 28 these two complexes was constant (58%/42%) irrespective of
 29 the amount of **3⁻**. The integration ratio of the ¹H signals **1** and
 30 **3⁻** revealed that both species are 1:1 complexes. For the
 31 major complex (58 %), two phenyl groups of **3⁻** were
 32 separately observed by ¹H NMR, and ROE correlations to the
 33 pyranose unit of the cyclodextrin were observed for only one
 34 of the two phenyls (Figure 2d, Figure S24). For the minor
 35 complex (42%), two phenyl groups are chemically equivalent
 36 in the ¹H NMR spectrum, and they showed ROE correlations
 37 to the sugar unit. These results suggest that the major species
 38 (58%) is a complex in which the guest is threaded through the
 39 cyclodextrin cavity (Recognition mode 1 in Figure 1b), while
 40 the minor species (42%) is a complex in which the guest is
 41 folded in the cyclodextrin's cavity (Recognition mode 2 in
 42 Figure 1b). Chemical shifts of the ³¹P NMR signals of the
 43 two complexes are different by more than 5 ppm (–5.61 and
 44 –10.68 ppm, respectively), which also supports the two
 45 distinctive recognition modes of the phosphate (Figure S23).
 46 The two species are separately observed by ¹H NMR even at
 47 high temperature (343 K), which shows a slow conversion
 48 and high energy barrier between these complexes (Figure
 49 S26).



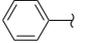
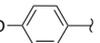
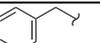
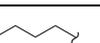
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51 **Figure 2.** Recognition of phosphodiester by **1**. (a)–(c) ¹H NMR spectra
 52 of aromatic region (600 MHz, CD₃CN). (a) (*n*-Bu₄N)(PhO-PO₂-OPh)
 53 ((*n*-Bu₄N)**3**). (b) **1** (0.8 mM). (c) **1** and (*n*-Bu₄N)**3** (1.3 eq). Protons of
 54 the host-guest complex in the ‘threading’ mode are denoted by green
 55 letters with subscript 1. Protons of the phenyl group of **3⁻** in the
 56 nitrophenyl group’s cavity are denoted with primed letters, and those in
 57 the cyclodextrin’s cavity are denoted with letters without prime. Protons
 58 of the host-guest complex in the ‘folding’ mode are denoted by blue
 59 letters with subscript 2. * indicates **3⁻** that is not encapsulated in **1**. (d)
 60 and (e) ¹H–¹H ROESY spectra (600 MHz, CD₃CN). (d) ROE
 61 correlations between aromatic regions and sugar regions for the host-
 62 guest complex between **1** and (*n*-Bu₄N)**3** (1.3 eq). (e) ROE correlations
 63 with *tert*-butyl group for the host-guest complex between **1** and (*n*-
 64 Bu₄N)(*p*-*t*-BuC₆H₄O-PO₂-O(*p*-*t*-BuC₆H₄)) ((*n*-Bu₄N)**4**) (1.3 eq). Red
 65 circles represent ROEs between *p*-nitrophenyl group of **1** and *t*-Bu group

1 of **4**⁻, and a green circle represents that between the sugar moiety of **1**
2 and *t*-Bu group of **4**⁻.

3
4 Table 1 summarizes the ratio of the two recognition
5 modes for the various phosphodiester, together with the 1:1
6 binding constants K_a [M^{-1}] (the stoichiometry has been
7 confirmed by the integral ratio of ¹H NMR signals of the
8 inclusion complexes; see also Figures S11–S21). For **4**⁻,
9 which has a bulky *t*-Bu group at the *p*-position of the benzene
10 ring, the equilibrium was biased toward the ‘threading’ mode
11 (‘threading’/‘folding’ = 87%/13%). Steric hindrance of the
12 *t*-Bu group would have destabilized the folded conformation
13 of **4**⁻ inside the cyclodextrin’s cavity. In the ‘threading’ mode,
14 a ¹H NMR signal assigned to one of the *t*-Bu groups was
15 shifted downfield (from 1.29 ppm to 0.81 ppm) as the result
16 of the shielding effects from the surrounding *p*-nitrophenyl
17 groups. The structure of the ‘threading’ mode was further
18 supported by the ROE correlation between the *t*-Bu group of
19 **4**⁻ and the nitrophenyl groups of receptor **1** (Figure 2e, Figure
20 S25). For **5**⁻, to which a methoxy group is introduced at the
21 *p*-position of the benzene ring, the recognition mode was also
22 biased toward the ‘threading’ mode (72%/28%). Meanwhile,
23 the ‘folding’ mode was slightly favored in the case of the
24 electron-deficient *p*-nitrophenyl phosphodiester **6**⁻
25 (47%/53%). The comparison of **3**⁻, **5**⁻, and **6**⁻ suggested that
26 the electron-rich substituent on the phosphodiester might
27 have stabilized the ‘threading’ mode through the
28 intermolecular interactions with the electron-deficient *p*-
29 nitrophenyl groups of **1**. Interestingly, for the non-phenolic
30 derivatives, dibenzyl phosphate **7** and dibutyl phosphate **8**⁻,
31 the recognition modes were biased toward the ‘folding’ mode
32 (41%/59%, 10%/90%, respectively). These results can be
33 explained as the result of the easiness of the flexible
34 diphosphate molecules to take a folded conformation in the
35 cyclodextrin’s cavity. The binding constants K_a of **1** and all
36 of the phosphodiester were high ($\log K_a \sim 4$ –6), among which
37 the highest affinity was observed for the dibutyl phosphate
38 **8**⁻ ($\log K_a = 6.2$).

39 **Table 1.** Effect of substituents on the ratio of two recognition modes
40 and binding constants K_a [M^{-1}] between **1** and
41 phosphodiester RO-PO₂⁻-OR (CH₃CN or CD₃CN, 298 K).

Guest anions ^a	Substituents (R)	Recognition mode 1 ^b (Threading)	Recognition mode 2 ^b (Folding)	Binding constant ^c $\log K_a$
3 ⁻		58%	42%	4.8
4 ⁻	<i>t</i> -Bu- 	87%	13%	4.5
5 ⁻	MeO- 	72%	28%	5.4
6 ⁻	O ₂ N- 	47%	53%	4.4
7 ⁻		41%	59%	4.6
8 ⁻		10%	90%	6.2

42 ^a *n*-Bu₄N⁺ salts except for **6**⁻. **6**⁻ was used as an *i*-Pr₂EtNH⁺ salt prepared
43 by mixing the corresponding free phosphoric acid and 1.5 equivalents of
44 *i*-Pr₂EtN. ^b Determined by ¹H NMR measurements. ^c Determined by UV-
45 vis titration experiments except for **6**⁻, which was determined by ¹H
46 NMR.

47
48 Regardless of the employed phosphodiester (**3**⁻–**8**⁻), the
49 ¹H NMR signals of *a* and *e* (1 and 5 positions of pyranose)
50 were observed at 5.14–5.21 ppm and 4.42–4.63 ppm for the
51 ‘threading’ mode, respectively (Figure S22). They appeared
52 in a different region for the ‘folding’ mode, that is, 5.03–5.08
53 ppm for proton *a* and 4.61–4.77 ppm for proton *e*. A similar
54 trend was also observed for the ³¹P NMR signals of **3**–**5**⁻
55 included within **1**. The signals were observed at –5.61 –
56 –4.90 ppm for the ‘threading’ mode, but at –10.68 –
57 –9.78 ppm for the ‘folding’ mode (Figure S23). These regularities
58 indicated that the structures of both the ‘threading’ and
59 ‘folding’ modes are fixed to some extent irrespective of the
60 substituents on the phosphodiester.

61 Figure 3 shows the calculated structures of receptor **1**
62 encapsulating diphenyl phosphate **3**⁻ for each recognition
63 mode (semi-empirical calculation (PM6)). In the ‘threading’
64 mode, one phenyl group of **3**⁻ is included in the
65 cyclodextrin’s ‘first’ cavity, while the other one is placed in
66 the ‘second’ cavity surrounded by *p*-nitrophenyl groups
67 (Figure 3a). The calculated electrostatic potential map of **1**
68 in the conformation of the ‘threading’ mode suggests that
69 many *p*-nitrophenyl groups are in the standing conformation
70 to allow the inclusion of the guest, and the ‘second’ cavity on
71 the electron-deficient *p*-nitrophenyl side has a partial positive
72 charge (Figure 3c). Thus, the model supports that receptor **1**
73 has two distinctive cavities on both sides of the amide groups,
74 and that the recognition pocket of **1** is regarded to have a
75 double-conical shape. The *para*-proton of the phenyl group
76 of **3**⁻ at the ‘second’ pocket (*k*₁’ in Figure 2c) is close to the
77 *p*-nitrophenyl rings of **1**, which is consistent with the upfield
78 shift of the corresponding NMR signal. In the ‘folding’ mode,
79 two phenyl groups are placed adjacent to each other in the
80 cyclodextrin’s cavity (Figure 3b). This bent conformation of
81 **3**⁻ is consistent with the downfield shift of the NMR signal of
82 the *ortho*-proton of **3**⁻ (*o*₂ in Figure 2c), which can be
83 explained by the deshielding effect from the opposite phenyl
84 group. The calculated electrostatic potential map of **1** for the
85 ‘folding’ mode suggests that the *p*-nitrophenyl groups filled
86 the upper space, and some tilted *p*-nitrophenyl groups
87 constituted the positively-charged ceiling of the
88 cyclodextrin’s cavity (Figure 3d). Phosphate groups formed
89 multiple hydrogen bonds with the amides in the cavity for
90 both structures. Thus, interesting inclusion modes of the
91 phosphodiester are observed utilizing the uniquely-
92 restricted aromatic-amide cyclodextrin framework.

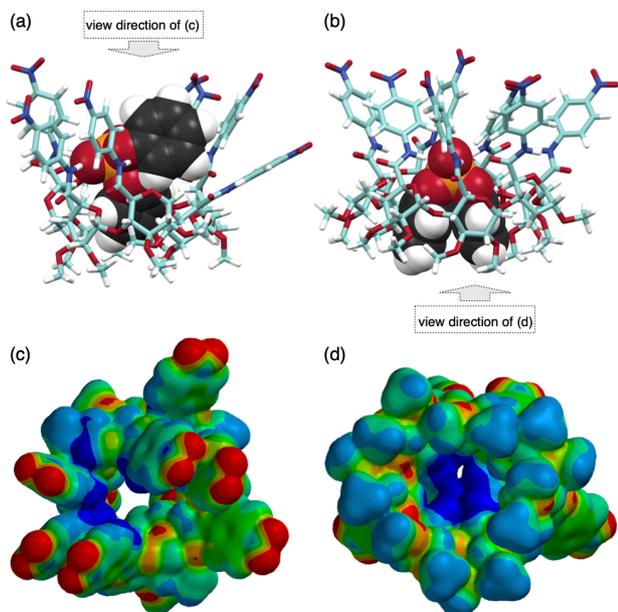


Figure 3. Structures of the two recognition modes of diphenyl phosphate (**3**) by **1** obtained by semi-empirical calculations (PM6). (a) and (c) ‘Threading’ mode. (b) and (d) ‘Folding’ mode. (a) and (b) **1**, stick model; diphenyl phosphate, space-filling model. H, white; C in **1**, light green; C in diphenyl phosphate, black; N, blue; O, red; P, orange. (c) and (d) Calculated electrostatic potential maps of **1** for the conformation of each mode (**3** has been removed). (c) View from the top. (d) View from the bottom.

In conclusion, we have synthesized a novel cyclodextrin receptor **1** possessing seven *p*-nitrophenylamide groups that shows a strong hydrogen-bonding ability. Receptor **1** is a double-conical-shaped host molecule that has two distinct cavities, and recognizes phosphodiester in two different recognition modes, i.e., the ‘threading’ mode and the ‘folding’ mode. The equilibrium is biased toward the ‘threading’ mode for the guests with bulky or electron-rich substituents, while the ‘folding’ mode is favored for the flexible guests. Further applications of **1** to utilize the precise recognition ability of a series of phosphate anions are now being investigated.

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Graphical Abstract

Textual Information

A brief abstract

A cyclodextrin derivative possessing multiple *p*-nitrophenylamide groups encapsulates phosphodiester anions in two recognition modes. One is the 'threading' mode, in which the phosphodiester passes through the cyclodextrin cavity. The other is the 'folding' mode, in which the included phosphodiester is bent inside the cyclodextrin cavity. The tendency between the two recognition modes depends on the substituents on the phosphodiesters.

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