

# Mechanism of Rapid Nuclear Factor-E2-Related Factor 2 (Nrf2) Activation via Membrane-Associated Estrogen Receptors: Roles of NADPH Oxidase 1, Neutral Sphingomyelinase 2 and Epidermal Growth Factor Receptor (EGFR)

著者 (英)	Tetsuro Ishii, Eiji WARABI
journal or publication title	Antioxidants
volume	8
number	3
page range	69
year	2019-03
権利	(C) 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license ( <a href="http://creativecommons.org/licenses/by/4.0/">http://creativecommons.org/licenses/by/4.0/</a> ).
URL	<a href="http://hdl.handle.net/2241/00160232">http://hdl.handle.net/2241/00160232</a>

doi: 10.3390/antiox8030069



Review

# Mechanism of Rapid Nuclear Factor-E2-Related Factor 2 (Nrf2) Activation via Membrane-Associated Estrogen Receptors: Roles of NADPH Oxidase 1, Neutral Sphingomyelinase 2 and Epidermal Growth Factor Receptor (EGFR)

Tetsuro Ishii \* and Eiji Warabi

Faculty of Medicine, University of Tsukuba, Tsukuba Ibaraki 305-8575, Japan; warabi-e@md.tsukuba.ac.jp

\* Correspondence: ishiitetsuro305@gmail.com

Received: 21 January 2019; Accepted: 13 March 2019; Published: 18 March 2019



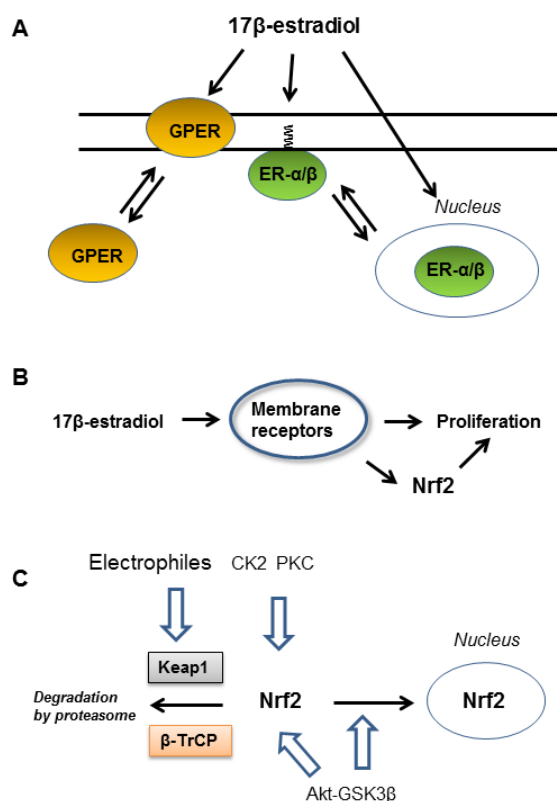
**Abstract:** Membrane-associated estrogen receptors (ER)- $\alpha$ 36 and G protein-coupled estrogen receptor (GPER) play important roles in the estrogen's rapid non-genomic actions including stimulation of cell proliferation. Estrogen via these receptors induces rapid activation of transcription factor nuclear factor-E2-related factor 2 (Nrf2), a master regulator of detoxification and antioxidant systems, playing a key role in the metabolic reprogramming to support cell proliferation. This review highlights the possible mechanism underlying rapid Nrf2 activation via membrane-associated estrogen receptors by estrogen and phytoestrogens. Stimulation of ER- $\alpha$ 36-GPER signaling complex rapidly induces Src-mediated transactivation of epidermal growth factor receptor (EGFR) leading to a kinase-mediated signaling cascade. We propose a novel hypothesis that ER- $\alpha$ 36-GPER signaling initially induces rapid and temporal activation of NADPH oxidase 1 to generate superoxide, which subsequently activates redox-sensitive neutral sphingomyelinase 2 generating the lipid signaling mediator ceramide. Generation of ceramide is required for Ras activation and ceramide-protein kinase C  $\zeta$ -casein kinase 2 (CK2) signaling. Notably, CK2 enhances chaperone activity of the Cdc37-Hsp90 complex supporting activation of various signaling kinases including Src, Raf and Akt (protein kinase B). Activation of Nrf2 may be induced by cooperation of two signaling pathways, (i) Nrf2 stabilization by direct phosphorylation by CK2 and (ii) EGFR-Ras-PI 3 kinase (PI3K)-Akt axis which inhibits glycogen synthase kinase 3 $\beta$  leading to enhanced nuclear transport and stability of Nrf2.

**Keywords:** estrogen; MCF-7; Nrf2; ER- $\alpha$ 36; GPER; EGFR; NOX1; neutral sphingomyelinase 2; ceramide; ERK; p38; PKC $\delta$ ; PKC $\zeta$ ; CK2; Akt; PI3K; phytoestrogen

## 1. Introduction

Estrogens have long been known as important regulators of the female reproductive functions. However, it is now commonly accepted that estrogens are locally synthesized and play important roles in the cardiovascular protection, bone preservation, and neuroprotection in both female and male (reviewed in [1–3]). Estrogens transduce signals through two signaling pathways via nuclear receptors estrogen receptor (ER)- $\alpha$  and ER- $\beta$ , and plasma membrane-associated receptors [4–6]. The G-protein coupled estrogen receptor GPER/GPR30 is a 7-transmembrane receptor implicated in rapid estrogen signaling [7–9]. However, estrogen signaling is complex because ER- $\alpha$  and ER- $\beta$  receptors can be palmitoylated to localize the cell membrane [5,10] and ER- $\alpha$  has three isoforms termed ER- $\alpha$ 66, ER- $\alpha$ 46 and ER- $\alpha$ 36, derived from different promoter usage and alternative splicing [6]. Additionally, GPER undergoes constitutive retrograde trafficking from the plasma membrane to the endoplasmic

reticulum and its surface expression is limited and controlled by intrinsic factors [7]. These estrogen receptors are expressed in cell type-specific manners increasing variation of signaling pathways as reviewed in [11–14] (Figure 1A).



**Figure 1.** 17β-estradiol activates nuclear factor-E2-related factor 2 (Nrf2) and enhances cell proliferation. (A) Part of G protein-coupled estrogen receptor (GPER) and estrogen receptors α and β (ER-α/β) are associated with plasma membrane and transduce mitogenic signals elicited by 17β-estradiol. (B) 17β-Estradiol acting through membrane-associated estrogen receptors stimulates proliferation of human breast cancer MCF-7 and other cells. Rapid Nrf2 activation by 17β-estradiol enhances proliferation. (C) Nrf2 is unstable due to Kelch-like ECH-associated protein 1 (Keap1)- and β-TrCP (transducing repeat-containing protein)-mediated ubiquitylation and following degradation by proteasome. Rapid activation of Nrf2 is regulated by both stabilization and facilitation of nuclear import of Nrf2. Electrophiles react with Keap1 leading to stabilization of Nrf2. Protein kinase C (PKC) and casein kinase 2 (CK2) phosphorylate/stabilize Nrf2. Glycogen synthase kinase 3β (GSK3β) phosphorylates Nrf2 to induce β-TrCP-mediated degradation and also suppresses nuclear import of Nrf2. Akt (protein kinase B) phosphorylates/inactivates GSK3β to enhance Nrf2 activation.

One of the biological functions of estrogen is stimulation of cell proliferation. ER-α is the major driver of breast cancer and multiple ER targeting drugs are routinely used clinically to treat patients with ER positive breast cancer [15–17]. Accumulating evidence indicates non-genomic rapid estrogen signaling via membrane-associated estrogen receptors such as ER-α36 and GPER play important roles in mitogenic estrogen signaling [6,7]. Transactivation of epidermal growth factor receptor (EGFR) via membrane-associated estrogen receptors plays a key role for the promotion of cell proliferation [6,7]. A rapid cellular response following mitogenic stimulation is activation of transcription factor nuclear factor-E2-related factor 2 (Nrf2), which regulates the expression of a large number of genes, including detoxification enzymes, antioxidant proteins and glutathione (GSH) synthesis system via the antioxidant responsive element (ARE) in a cell type specific manner [18–20]. Additionally, Nrf2 upregulates metabolic genes involved in cell proliferation and differentiation [21]. Nrf2 promotes proliferation of various cancer cells and enhances resistance to anticancer drugs [22–25].

For example, stimulation of human breast cancer MCF-7 cells with nM levels of 17 $\beta$ -estradiol rapidly activates Nrf2 via the PI3K dependent manner [26]. However, precise signaling cascades downstream of estrogen receptors leading to rapid activation of Nrf2 are currently not clear (Figure 1B).

Rapid Nrf2 activation can be induced by both stabilization of Nrf2 and enhanced nuclear import of Nrf2 protein. Nrf2 protein is unstable under normal conditions because the cytosolic Nrf2 binding protein Kelch-like ECH-associated protein 1 (Keap1) facilitates degradation of Nrf2 by proteasome [27–29]. It is well known that electrophilic compounds chemically react with sulfhydryl residues in Keap1 and inhibit degradation of Nrf2 [27–29]. In addition to Keap1, another ubiquitin ligase adaptor  $\beta$ -transducin repeat-containing protein ( $\beta$ -TrCP) also destabilizes Nrf2 when glycogen synthase kinase 3 $\beta$  (GSK3 $\beta$ ) phosphorylates serine residues in Nrf2 [30,31]. It is also proposed that GSK3 $\beta$  suppresses nuclear import of Nrf2 [32,33]. High glucose or diabetic conditions accompanies upregulation of GSK3 $\beta$  activity and facilitates degradation of Nrf2 [34]. Therefore, inhibition of GSK3 $\beta$  by protein kinase Akt (protein kinase B) secures activation of Nrf2 [30,33]. Additionally, some protein kinases such as protein kinase C (PKC) [35] and casein kinase 2 (CK2) [36–38] directly phosphorylate and protect Nrf2 from Keap1-dependent degradation (Figure 1C).

This review highlights the signaling pathways leading to rapid activation of Nrf2 by 17 $\beta$ -estradiol and some estrogen mimetics via membrane-associated estrogen receptors. Nrf2 activation via hormone-receptor signaling cascades differs from that induced by toxic electrophiles which directly target Keap1. We recently proposed that neurotrophins can activate Nrf2 by a mechanism dependent on the common neurotrophin receptor p75<sup>NTR</sup> (neurotrophin receptor) [39–41]. This hypothesis is based on the facts that nerve growth factor activates CK2 by a p75<sup>NTR</sup>-dependent manner [42] and that CK2 directly phosphorylates Nrf2 to stabilize/activate it [36,37]. Notably, stimulation of p75<sup>NTR</sup> induces generation of the lipid signal mediator ceramide via activation of neutral sphingomyelinase (nSMase) [43]. As ceramide activates PKC $\zeta$  at low concentrations [44] and PKC $\zeta$  can phosphorylate/activate CK2 [45], we hypothesized that neurotrophins activate Nrf2 via p75<sup>NTR</sup>-ceramide-PKC $\zeta$ -CK2 pathway [39], although mechanism of nSMase activation via p75<sup>NTR</sup> is currently not known. We here propose novel signaling pathways downstream of membrane-associated estrogen receptors leading to Nrf2 activation with a special interest in the regulation of proliferation of estrogen-dependent breast cancer cell MCF-7.

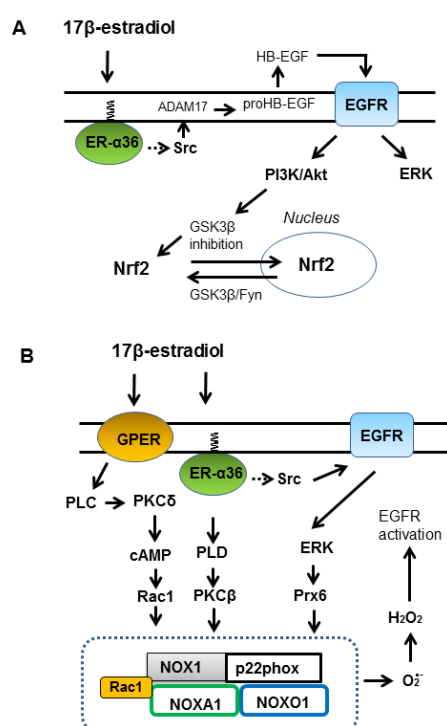
## 2. Role of ER- $\alpha$ 36 in EGFR-Dependent Activation of Nrf2

ER- $\alpha$ 36 is widely expressed in estrogen sensitive cancer cells, and high expression levels correlates with a poor survival prognosis for breast cancer patients [46]. MCF-7 cells express ER- $\alpha$ 66 as the major ER- $\alpha$ , but also express ER- $\alpha$  isoforms, ER- $\alpha$ 46 and ER- $\alpha$ 36 [6]. ER- $\alpha$ 36 differs from the ER- $\alpha$ 66 by lacking N- and C-terminal transcriptional activation domains but retaining the central DNA-binding and dimerization domains, and partial ligand-binding domains. It also possesses an extra, unique 27 amino acid domain to replace the last 138 amino acids of the ER- $\alpha$ 66 [6]. All three ER- $\alpha$  isoforms contain the cysteine residue in the ligand binding domain for S-palmitoylation, which is a reversible post-translational lipid modification important for membrane anchoring [47].

ER- $\alpha$ 36 localizes in the cytoplasm and at the plasma membrane, mediating rapid estrogen signaling in human breast cancer cells [6] and plays a key role in estrogen-dependent proliferation [48]. Post-translational palmitoylation is suspected to play a crucial role in anchoring ER- $\alpha$ 36 to the cell membrane and interaction with caveolin-1 [10]. Activation of ER- $\alpha$ 36 leads to activation of phospholipase D and PKC in breast cancer cells [49]. Stimulation of membrane-associated estrogen receptors accompanies transactivation of epidermal growth factor receptor (EGFR), inducing rapid kinase cascades that influence both transcriptional and non-transcriptional actions of estrogen [50].

EGFR is expressed in various cell types and plays a key role in the regulation of cell proliferation, survival and differentiation. In pathological settings, mostly in lung and breast cancer and in glioblastoma, EGFR is overexpressed or mutated [51–53]. Activation of EGFR can be caused by the release of heparin-binding EGF-like growth factor (HB-EGF) from membrane-anchored form (proHB-EGF) by a metalloprotease ADAM17. Although intracellular Ca<sup>2+</sup> elevation is required for the

activation of tyrosine kinase Src which lead to activation of ADAM17 [54], precise mechanism of Src activation is not known. EGFR signaling activates Ras/RAF/MEK/ERK and Ras/PI3K/PTEN/Akt pathways [55,56]. Notably, treatment of MCF-7 cells with 1 nM 17 $\beta$ -estradiol induced rapid association of ER- $\alpha$ 36 with Src at the plasma membrane and activation of ERK in 5 min [57], and activation/phosphorylation of Akt and inhibition/phosphorylation of GSK3 $\beta$  in 15 min [26]. As GSK3 $\beta$  destabilizes Nrf2 and suppresses Nrf2 nuclear import, activation of Akt plays a key role in the activation of Nrf2-ARE-mediated target gene expression [30–33]. Wu et al. [26] showed that a PI3K-specific inhibitor wortmannin significantly inhibited Nrf2 activation by 17 $\beta$ -estradiol in MCF-7 cells [26]. These studies suggest EGFR-PI3K-Akt-GSK3 $\beta$  signaling downstream of ER- $\alpha$ 36 largely contributes Nrf2 nuclear import and activation of gene transcription via ARE elements at physiological concentrations of 17 $\beta$ -estradiol (Figure 2A). However, Wu et al. [26] also showed that a kinase inhibitor LY294002, which inhibits both PI3K and CK2, also inhibited Nrf2 activation by 17 $\beta$ -estradiol in MCF-7 cells more efficiently than wortmannin [26].



**Figure 2.** Activation of epidermal growth factor receptor (EGFR) and NADPH oxidase 1 via membrane-associated estrogen receptors. (A) Stimulation of ER- $\alpha$ 36 induces transactivation of EGFR receptor (EGFR) leading to activation of Ras/RAF/MEK/ERK and Ras/PI3K/PTEN/Akt pathways. Activated Akt inhibits GSK3 $\beta$  and enhances nuclear import of Nrf2 and stability of Nrf2. HB-EGF (heparin-binding EGF-like growth factor), ADAM17, PLC (phospholipase C), PLD (phospholipase D), Prx6 (peroxiredoxin 6), NOXA1, NOXO1. (B) GPER-mediated signaling cooperate with ER- $\alpha$ 36-EGFR signaling leading to activation of NADPH oxidase 1 (NOX1), which generates superoxide. Superoxide is metabolized to H $_2$ O $_2$ , which enhances EGFR signaling. Activation of Rho family GTPase Rac1, PKC $\beta$  and ERK can induce NOX1 activation.

ER- $\alpha$ 36 is expressed in rat and human brain especially high levels in cerebral cortex and plays a role in protection of neurons under oxygen and glucose deprivation [58]. Estrogen synthetic enzyme aromatase is expressed in neurons under basal conditions and is upregulated in reactive astrocytes after injury [59]. Zhu et al. [60] showed intravitreal injection of 17 $\beta$ -estradiol induced rapid activation of Nrf2 in retina's cell layers and protected the tissue from light-induced degeneration [60]. After the 17 $\beta$ -estradiol treatment, Nrf2 protein levels significantly increased in 4 h and peaked in 12 h. The kinase inhibitor LY294002 effectively blocked Nrf2 activation in retina's cell layers [60].

### 3. Functions of Membrane-Associated GPER

Cloned GPER cDNA predicts GPER is a 375 amino acid protein with the structural signature of the Class A Rhodopsin family 7-transmembrane receptor with about 30% amino acid identity to angiotensin II (AngII) and interleukin 8 receptors [9]. Compared to ER- $\alpha$ 36, GPER is expressed ubiquitously and has now become recognized as the major mediator of estrogen's rapid cellular effects throughout the body. It is not involved in reproduction but GPER deficiency results in multiple physiological alterations including obesity, cardiovascular dysfunction, insulin resistance and glucose intolerance (reviewed in [61]). Activation of GPER can mediate multiple salutary effects on the cardiovascular system and GPER-selective agonist G1 can be considered a non-feminizing estrogenic compound and thus of potential therapeutic use in both women and men.

GPER associates with  $G\alpha_s$  protein and activates phospholipase C (PLC) to catalyze the hydrolysis of membrane-bound phosphatidylinositol 4,5-bisphosphate (PIP<sub>2</sub>) into inositol 1,4,5-triphosphate (IP<sub>3</sub>) and diacylglycerol (DAG) [62]. DAG activates PKC $\delta$  to augment adenylyl cyclase activity generating cAMP [45]. Additionally, GPER has two notable functions, which may not necessarily be beneficial for health. First, it regulates Ca<sup>2+</sup> signaling via interaction with plasma membrane Ca<sup>2+</sup>-ATPase (PMCA) [63]. PMCA is essential for removal of cytoplasmic Ca<sup>2+</sup> and for shaping the time courses of Ca<sup>2+</sup>-dependent activity. The authors show that GPER physically associates with PMCA and suppresses PMCA activity. GPER agonist G1 further inhibits PMCA activity through tyrosine phosphorylation of the pump resulting in an increase in intracellular Ca<sup>2+</sup> levels. Second, GPER enhances expression of NADPH oxidase 1 (NOX1) [64,65]. NOX1 generates superoxide which is instantly metabolized to hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) by superoxide dismutase. Meyer et al. [64] observed that genetic ablation of GPER in mice prevented cardiovascular pathologies associated with aging by reducing superoxide formation through downregulation of NOX1 expression. Blocking GPER activity with specific antagonist G36 decreased NOX1 abundance and decreased superoxide production and arterial hypertension in mice chronically infused with AngII (0.7 mg/kg body weight/day) [64,65].

In addition to vascular cells [66,67], NOX1 is expressed in various cells including neurons and cancer cells [68]. Activation of NOX1 and generated superoxide and H<sub>2</sub>O<sub>2</sub> serve as co-stimulatory signals for cell proliferation in many normal and cancer cells [69–73]. NOX1-derived ROS promoted phosphorylation of ERK1/2 and expression of cyclin D1 and Fos family genes during the immediate early gene response [74]. As important consequences of GPER activation are the regulation of cell growth, migration, and apoptotic cell death [75], GPER may contribute to NOX1 activation in addition to upregulation of NOX1 expression.

### 4. Cooperation of ER- $\alpha$ 36 and GPER in NADPH Oxidase 1 Activation

Activation of NOX1 requires association with its regulatory components p22phox, NOX organizer 1 (NOXO1), NOX activator 1 (NOXA1) and Rho family small GTPase Rac1 [68]. Choi et al. [76] ectopically over-expressed AngII type 1 receptor (AT<sub>1</sub>R), NOX1, NOXO1 and NOXA1 using virus vectors into HEK293 and CHO cells and characterized mechanism and kinetics of NOX1 activation by AngII [76]. The activation of NOX1 by AngII was very fast and the production of superoxide peaked at around 10 min but decayed faster. The earliest response of AngII-AT<sub>1</sub>R signaling is  $G\alpha_q$ -mediated activation of phospholipase- $\beta$ , which is followed by Ca<sup>2+</sup> mobilization and activation of PKC, and G protein-dependent and -independent activation of Src family kinases [77]. The authors observed that extracellularly added EGF also induces NOX1 activation in the model cells, which had additionally expressed EGFR, with a similar kinetics but generated superoxide was lower compared to that by AngII-AT<sub>1</sub>R-NOX1 under the experimental conditions [76]. This result suggests that once EGFR is activated it will enhance NOX1 activation. Notably, single addition of PKC agonist phorbol 12-myristate 13-acetate at 0.3  $\mu$ M induced NOX1 activation and superoxide generation in the model cells at similar extent as that by 1  $\mu$ M AngII [76]. Notably, activation of Src is required for AngII induced NOX1 activation, and inhibition of PI3K by wortmannin or LY294002 attenuated both Rac activation and ROS generation in vascular endothelial cells [78].



These studies suggest that ER- $\alpha$ 36 and GPER respectively facilitate NOX1 activation. NOX1 phosphorylation at Thr429 by PKC $\beta$  facilitates the association of NOX1 with the NOXA1 in vascular smooth muscle cells [79] and GPER-mediated increases in cAMP to activate Rac1 via cAMP dependent Epac1/Rap1 signaling [80]. Moreover, in tumor colon epithelial cells, activated Rac1 interacts with NOXA1 to enhance NOX1 activity [81]. GPER-mediated Ca<sup>2+</sup> mobilization may activate PKC and contribute in NOX1 activation like the phorbol ester (Figure 2B). However, currently there is no report showing 17 $\beta$ -estradiol at low levels activates NOX1 in MCF-7 or other cells. However, Lei et al. [82] showed that stimulation of MCF-7 cells with bisphenol F (a estrogenic pollutant, 10–100 nM) induced rapid but marginal increases in both intracellular Ca<sup>2+</sup> and ROS levels in an ER- $\alpha$  and GPER dependent manner [82]. Bisphenol F is one of the alternatives to an estradiol mimetic bisphenol A and induces proliferation of MCF-7 cells similar to 17 $\beta$ -estradiol [83].

GPER forms a complex with ER- $\alpha$ 36 and also with p65 component of NF- $\kappa$ B, acting as suppressor of NF- $\kappa$ B in human primary monocytes [84]. Additionally, stimulation of GPER by 1 nM 17 $\beta$ -estradiol induces rapid gene expression of ER- $\alpha$ 36 by an AP-1 dependent manner in SK-BR-3 breast cancer cells [85] and by PKA/CREB pathway in seminoma-like TCam-2 cells [86]. In these cancer cell lines, upregulation of ER- $\alpha$ 36 is required for cell proliferation. Expression of EGFR is also upregulated in TCam-2 cells by the 17 $\beta$ -estradiol treatment [86]. Interestingly, GPER-EGFR-ERK-c-Fos-AP-1 signaling also upregulates GPER expression in MCF-7 cells [83] and spermatogonial GC-1 cells [87]. These studies suggest existence of a functional cooperation between GPER, ER- $\alpha$ 36, EGFR and NOX1 in supporting cell proliferation following stimulation with 17 $\beta$ -estradiol or estrogen mimetics (Figure 2B).

Once NOX1 is activated, superoxide and H<sub>2</sub>O<sub>2</sub> will be generated. NOX1 transfers an electron from intracellular NADPH across the cell membrane to molecular oxygen to generate superoxide in the extracellular space or within a signaling endosome if the NOX1 is endocytosed with the membrane receptors [88]. Importantly, extracellular or intra-endosomal superoxide can penetrate cell membranes through a chloride channel-3 (CIC-3) [88]. Cellular superoxide dismutase converts superoxide to H<sub>2</sub>O<sub>2</sub>, which can traverse the cell membrane through aquaporin channels [89]. Therefore, both superoxide and H<sub>2</sub>O<sub>2</sub> may interact with specific cellular targets to initiate intracellular signaling. Recent studies show that 17 $\beta$ -estradiol upregulates expression of CIC-3 channels by an ER- $\alpha$ -dependent manner in MCF-7 cells [89] and that the CIC-3 channel activity contributes proliferation of MCF-7 and MDA-MB-231 breast cancer cells [90].

As low levels of H<sub>2</sub>O<sub>2</sub> inhibit activity of protein tyrosine phosphatases which result in the activation of receptor tyrosine kinase [91], activation of NOX1 will contribute to maintain/enhance EGFR signaling. At the same time EGFR activation may help NOX1 activation as demonstrated in the model cells [76]. Association of NOX1 to plasma membrane requires the presence of phosphatidylinositol lipids [92]. It is produced by calcium-independent phospholipase A<sub>2</sub> or peroxiredoxin 6, which is activated/phosphorylated by ERK and p38 [93]. Thus, once NOX1 and EGFR are activated, ER- $\alpha$ 36-GPER-EGFR-NOX1 seems to form a positive feedback loop to further enhance EGFR-mediated signaling cascade (Figure 2B). However, we think superoxide generated by NOX1 plays another important function to support EGFR-mediated kinase cascade.

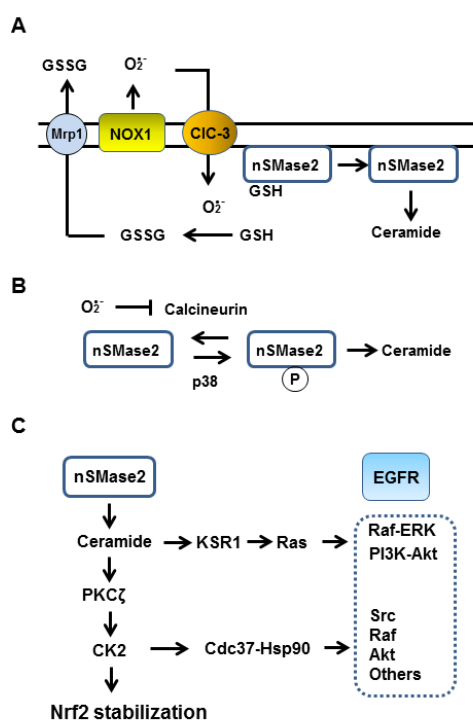
## 5. Superoxide Dependent Activation of Neutral Sphingomyelinase 2

We suggest that GPER-ER- $\alpha$ 36-NOX1 signaling could rapidly induce activation of neutral sphingomyelinase 2 (nSMase2), which hydrolyzes sphingomyelin to generate ceramide. Ceramide is a pleiotropic signal mediator of cellular responses including differentiation, proliferation, cell cycle arrest and apoptosis [94].

A notable characteristic of nSMase2 is that enzyme activity can be reversibly inhibited by GSH. Liu and Hannun [95] partially purified nSMase2 from MCF-7 cells and observed that reduced GSH at 1 to 3 mM almost linearly inhibited the activity from about 0 to 93% with a 50% inhibition at around 1.7 mM. These concentrations of GSH have physiological relevance as cultured cells have a few mM GSH depending on cell types and culture conditions [96]. Interestingly, the authors showed that

dilution of the pre-incubated medium containing the enzyme plus GSH induced rapid recovery of the enzyme activity suggesting the inhibition by GSH is reversible. They also showed that reducing agents, dithiothreitol and  $\beta$ -mercaptoethanol, neither inhibit nSMase activity nor disturb the inhibition by GSH, suggesting reduction of disulfide(s) may not play a role in the GSH-mediated enzyme inhibition. These results clearly suggest that nSMase2 has a low affinity GSH binding site and that GSH acts as a ligand to inhibit nSMase2 activity by a non-covalent and reversible manner. The inhibition of nSMase2 by GSH may be a feedback response to maintain cellular GSH levels as low levels of ceramide could activate Nrf2 leading to upregulation of GSH synthesis via expression of the cystine transporter and  $\gamma$ -glutamyl-cysteine synthetase [39–41]. The idea that GSH acts as a non-covalent ligand is already known in case of feedback inhibition by GSH of  $\gamma$ -glutamyl-cysteine synthetase, the rate limiting enzyme for GSH synthesis [97].

Taken together, nSMase2 activity seems to be suppressed in cells under normal conditions with high GSH and that the enzyme activation requires downregulation of cellular GSH. Notably, superoxide but not  $H_2O_2$  rapidly and effectively oxidizes GSH to generate disulfide GSSG [98], which can be exported from cells [99]. Importantly, the complete or significant depletion of whole cellular GSH is not required for the superoxide dependent transient activation of nSMase2 as membrane estrogen receptors, NOX1, nSMase2 and EGFR may be closely located in the membrane and NOX1-derived superoxide would rapidly down-regulate GSH in a cellular micro-compartment or micro-environment and facilitate activation of closely located nSMase2 (Figure 3A).



**Figure 3.** Role of neutral sphingomyelinase-2 (nSMase2) in Nrf2 activation and transactivation of EGFR. (A) NOX1 extracellularly generates superoxide, which can pass cell membrane through chloride channel-3 (CIC-3) to oxidize cellular glutathione (GSH) to GSSG. As GSSG can rapidly be exported from cells via multidrug resistant protein 1 (Mrp1), activation of NOX1 leads to temporal decrease in cellular GSH levels. GSH non-covalently binds neutral sphingomyelinase-2 (nSMase2) to inhibit the enzyme activity. Decrease in GSH induces nSMase2 activation and generation of lipid signaling molecule ceramide. (B) Superoxide inactivates calcineurin, which is bound to nSMase2 to inhibit from activation by p38. (C) Ceramide plays a crucial role for the transactivation of EGFR. Ceramide is required for KSR1 (kinase suppressor of Ras 1)-mediated activation of Ras, and CK2 plays an important role in the activation of Cdc37-Hsp90 molecular chaperone which mediates activation of signaling protein kinases, including Src, Raf and Akt. Ceramide-PKC $\zeta$ -CK2 axis stabilizes/activates Nrf2 [39].



The activation of nSMase2 is also modulated by protein kinases. PKC $\delta$  mediates translocation of nSMase2 from the Golgi to the plasma membrane [100]. Phosphorylation by p38 mitogen-activated protein kinase is important for the activation of nSMase2 in cigarette smoke-treated human airway epithelial cells [101] and in rat hippocampal astrocytes following cerebral ischemia [102]. Filosto et al. [103] showed that calcineurin (protein phosphatase 2B) directly associates with nSMase2 and suppresses phosphorylation/activation of nSMase2. Calcineurin is a serine/threonine phosphatase involved in a wide range of cellular responses to calcium mobilizing signals. Calcineurin has a Fe-Zn active center which becomes sensitive to superoxide in its calcium-activated conformation [104], and copper/zinc superoxide dismutase protects calcineurin [105,106]. Therefore, it is possible that p38 may phosphorylate/activate nSMase2 when superoxide is generated to inactivate calcineurin (Figure 3B). As Src can activate p38 [107] and PKC $\delta$  [108], once Src is stably activated, nSMase2 activity may be enhanced or maintained while superoxide is generated. Taken together, generation of superoxide could cause activation of nSMase2 in at least two ways, via oxidation of GSH and inactivation of calcineurin (Figure 3A,B).

In fact, it was shown that superoxide can activate nSMase in cultured cells [109,110]. Sawada et al. [109] examined effects of DNA damaging agent etoposide on ceramide-dependent apoptosis of human glioma cells and found long lasting nSMase activation which was dependent on prolonged generation of superoxide rather than H<sub>2</sub>O<sub>2</sub>. In case of amyloid- $\beta$ -induced neuronal death, nSMase activation occurred by a NADPH oxidase-superoxide-H<sub>2</sub>O<sub>2</sub> dependent manner [110]. These cases suggest that superoxide is involved in nSMase activation and that long lasting ceramide generation or higher levels of ceramide induces cell apoptosis rather than protection. Notably, ceramide effectively activates PKC $\zeta$  only at concentrations between 1 and 60 nM, and higher concentrations above 100 nM hardly enhance autophosphorylation/activation of PKC $\zeta$  [44]. Therefore, mild and temporal increases in ceramide levels via NOX1-nSMase2 axis can induce PKC- $\zeta$ -CK2-mediated phosphorylation/stabilization of Nrf2 (Figure 3C).

## 6. Ceramide Activates EGFR-Mediated Signaling Kinase Cascade

Ceramide generation by nSMase2 seems to be required for the EGFR mediated signaling cascade. This idea comes from the two facts; (i) Ras scaffolding protein KSR1 is a ceramide-binding protein requiring low levels of ceramide for membrane translocation to trigger the Ras signaling via EGFR [111,112] and (ii) nSMase inhibitor GW4869 thoroughly abrogates activation of ERK and p38 MAP kinases MAPKs in rat glioma C6 cells stimulated by phytochemicals, pent-acetyl geniposide [113] or caffeic acid phenethyl ester (CAPE) [114]. These compounds respectively induce rapid activation of nSMase (peaking at 15 min) and ERK and p38 phosphorylation/activation within 30 min in C6 cells, noting these authors suggested ceramide generation is required for activation of MAPKs [113,114].

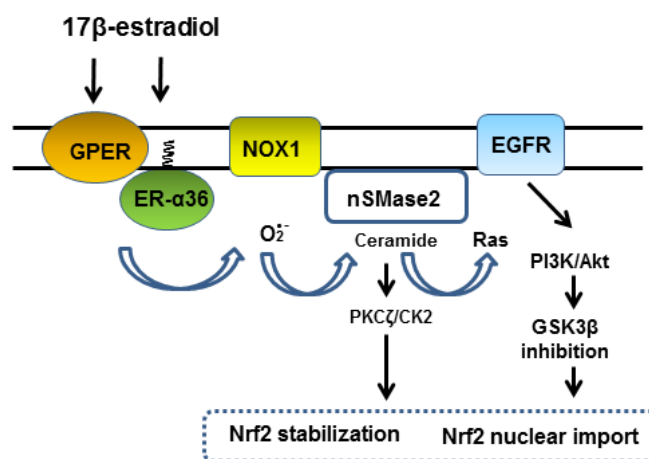
Geniposide, a major iridoid component of *Gardenia* fruit, is an agonist for G-protein coupled glucagon-like peptide-1 (GLP-1) receptor [115] and GLP-1 is known to transactivate EGFR [116]. Therefore, GLP-1 receptor functions like GPER in transactivation of EGFR. GLP-1 induces pancreatic  $\beta$ -cell proliferation and induces Nrf2 activation [117]. Geniposide and its structurally-related compounds are known to activate Nrf2 [118,119]. Peng et al. [113] showed that treatment of rat glioma C6 cells with pent-acetyl geniposide induced rapid and temporal activation of nSMase with intracellular ceramide levels increasing three to four fold at 2 to 3 h. Under the experimental conditions, these authors observed a rapid and temporal decrease in intracellular total GSH levels in C6 cells. The GSH levels decreased by about 30% in 0.5 to 1 h, thereafter the GSH levels gradually recovered in 2–3 h. They did not detect any peroxidized lipid metabolites in the cells, suggesting limited oxidative cell damage occurred under the experimental conditions. Although the authors did not measure superoxide generation in the C6 cells in the early time phase of pent-acetyl geniposide treatment, we speculate that superoxide was generated to oxidize GSH to GSSG, which was rapidly exported from cells via multidrug resistance protein Mrp1 [99] leading to temporal decrease in cellular GSH.

CAPE is an active ingredient of coffee, vegetables and beehive propolis and shown to induce Nrf2 activation in renal epithelial cells [120] and in mice brain [121]. As CAPE has a weak affinity for estrogen receptors ER $\alpha$  and ER $\beta$ , it may also interact with membrane-associated estrogen receptors [122]. Although these authors used the compounds at a high concentration (50  $\mu$ M), the results suggest a possibility that GPER/GLP-1R-NOX1-nSMase2-ceramide-KSR1 signaling is necessary for Ras-PI3K/ERK signaling downstream of Src-activated EGFR in C6 cells (Figure 3C).

In addition to ceramide-KSR1 signaling, ceramide-PKC $\zeta$ -CK2 signaling also plays a key role in enhancing kinase signaling via interaction of CK2 with molecular chaperone Cdc37 [123]. Activation of CK2 drastically changes cell metabolism [124] partly through interaction with Cdc37 [125]. Cdc37 is known as a cochaperone of Hsp90 interacting with many signaling protein kinases including Src, Raf-1 and Akt [123,125,126]. Miyake and Nishida [123] showed that CK2 associates with Cdc37 and phosphorylates Ser13 leading to increase in its chaperone activity resulting in activation of associated kinases. Therefore, nSMase-ceramide-CK2 signaling contributes to activation of Src to enhance ADAM17 mediated EGFR activation and in kinase signaling cascade downstream of EGFR (Figure 3C).

## 7. Conclusions and Future Perspectives

We propose that the stimulation of membrane-associated estrogen receptors initially activates NOX1. NOX1-derived superoxide activates nSMase2 to generate ceramide which subsequently activates Ras and PKC $\zeta$ -CK2 axis. The CK2-Cdc37-Hsp90 axis supports EGFR-mediated signaling kinase cascade. Activation of Nrf2 depends on both PI3K-Akt-GSK3 $\beta$  and PKC $\zeta$ -CK2 signaling pathways (Figure 4). As PI3K seems to be required for NOX1 activation [78], the kinase inhibitor LY294002, which inhibits both PI3K and CK2 activities, could significantly suppress Nrf2-ARE signaling as was observed in MCF-7 cells following treatment with 10 nM 17 $\beta$ -estradiol [26].



**Figure 4.** Summary of signal transduction mediated by membrane-associated estrogen receptors leading to activation of Nrf2. 17 $\beta$ -Estradiol via GPER and ER- $\alpha$ 36 receptors initially activates Src and NOX1 to generate superoxide, which in turn activates nSMase2 to generate the signal mediator ceramide. Ceramide plays a key role for the activation of CK2 and the EGFR-mediated kinase cascade. Direct modification of Nrf2 by CK2 and enhanced nuclear import of Nrf2 by Akt/GSK3 $\beta$  achieve rapid and effective Nrf2-ARE mediated gene expression. Inhibition of GSK3 $\beta$  also suppresses  $\beta$ -TrCP dependent Nrf2 degradation.

The importance of superoxide in Nrf2 activation has been demonstrated in vascular endothelial cells exposed to vasoprotective unidirectional laminar shear stress [127]. Fluid shear stress regulates endothelial cell function but the precise mechanism involved in mechanotransduction remains unclear. Czarny et al. [128,129] showed that nSMase is concentrated at the endothelial cell surface in caveolae and is activated to produce ceramide in an acute and transient manner in response

to increased flow rate and pressure in rat lung vasculature. We speculate that the shear stress activates NOX1 which subsequently induces nSMase2-ceramide signaling leading to Nrf2 activation. The authors showed that extracellularly supplemented cell-permeable ceramide analogs induced activation of Src and ERK kinases, and also the Akt-dependent endothelial nitric oxide synthase (eNOS) pathway [129]. These studies suggest that an unidentified sensor for mechanical stress induces rapid activation of NOX1-nSMase2-ceramide axis, which then leads to PKC $\zeta$ -CK2 and PI3K-Akt signaling in vascular endothelial cells by similar signaling pathways shown in Figure 3; Figure 4. It is noted that the estrogen mimetic isoflavone Equol via membrane-associated receptors rapidly activates eNOS/Hsp90 by an ERK and Akt dependent manner in human endothelial cells [130]. Taken together, NOX1, nSMase2 and EGFR seems to compose a common platform in membrane caveolae to mediate various receptor-mediated rapid signaling cascades. Probably membrane-associated estrogen receptors share this signaling platform to supplement or boost various signaling pathways including mechanotransduction in vascular endothelial cells and GLP-1R signaling in  $\beta$ -cell. This idea could explain why estrogen signaling is important in the cardiovascular protection, bone preservation, neuroprotection, metabolism and proliferation of various cell types.

Rapid activation of Nrf2 by low levels of 17 $\beta$ -estradiol will favor proliferation of MCF-7 cancer cells [26] by facilitating metabolic reprogramming through upregulation of enzymes for glucose metabolism and at the same time suppresses generation of ROS to protect cells from oxidative damages. The membrane-associated estrogen receptors are targets for phytoestrogens and endocrine-disrupting substances [8,131]. In addition to the phytoestrogen genistein, chemically synthesized compounds bisphenol A and nonylphenol have high affinities for ER- $\alpha$ , ER- $\beta$  and GPER [8,131]. Cadmium chloride activates GPER-mediated signaling at concentrations 5 to 500 nM and promotes cell proliferation like 17 $\beta$ -estradiol in human breast cancer, lung adenocarcinoma and thyroid cancer cells [132–134]. These compounds may increase cancer risks partly through promotion of cell proliferation [135,136]. Further studies on NOX1-nSMase2-ceramide axis will help to find novel strategies for hormone replacement and cancer therapies.

**Funding:** This work is supported by Japanese Society for Promotion of Sciences KAKENHI Grant Numbers JP21500386 (T.I.) and JP19790178 (E.W.).

**Acknowledgments:** We thank Giovanni E. Mann, King's College London, UK for valuable discussion.

**Conflicts of Interest:** The authors declare no conflict of interest.

## Abbreviations

Ang II	angiotensin II
ARE	antioxidant responsive element
AT <sub>1</sub> R	angiotensin II type 1 receptor
CIC-3	chloride channel 3
CK2	casein kinase 2
DAG	diacylglycerol
EGFR	epidermal growth factor receptor
eNOS	endothelial nitric oxide synthase
GLP-1	G-protein coupled glucagon-like peptide-1
GPER	G-protein coupled estrogen receptor
GSK3 $\beta$	glycogen synthase 3 $\beta$
Keap1	Kelch-like ECH-associated protein 1
Mrp1	multidrug resistance protein 1
NOXA1	NOX activator 1
NOX1	NADPH oxidase 1
NOXO1	NOX organizer 1
Nrf2	nuclear factor-E2-related factor 2
nSMase2	neutral sphingomyelinase 2
PLC	phospholipase C
PMCA	plasma membrane Ca <sup>2+</sup> -ATPase

## References

1. Menazza, S.; Murphy, E. The expanding complexity of estrogen receptor signaling in the cardiovascular system. *Circ. Res.* **2016**, *118*, 994–1007. [[CrossRef](#)]
2. Khosla, S.; Oursler, M.J.; Monroe, D.G. Estrogen and the skeleton. *Trends Endocrinol. Metab.* **2012**, *23*, 576–581. [[CrossRef](#)] [[PubMed](#)]
3. Behl, C. Oestrogen as a neuroprotective hormone. *Nat. Rev. Neurosci.* **2002**, *3*, 433–442. [[CrossRef](#)] [[PubMed](#)]
4. Arnal, J.F.; Lenfant, F.; Metivier, R.; Flouriot, G.; Henrion, D.; Adlanmerini, M.; Fontaine, C.; Gourdy, P.; Chambon, P.; Katzenellenbogen, B.; et al. Membrane and nuclear estrogen receptor alpha actions: From tissue specificity to medical implications. *Physiol. Rev.* **2017**, *97*, 1045–1087. [[CrossRef](#)]
5. Li, L.; Haynes, M.P.; Bender, J.R. Plasma membrane localization and function of the estrogen receptor alpha variant (ER46) in human endothelial cells. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 4807–4812. [[CrossRef](#)] [[PubMed](#)]
6. Wang, Z.Y.; Yin, L. Estrogen receptor alpha-36 (ER- $\alpha$ 36): A new player in human breast cancer. *Mol. Cell. Endocrinol.* **2015**, *418*, 193–206. [[CrossRef](#)] [[PubMed](#)]
7. Gaudet, H.M.; Cheng, S.B.; Christensen, E.M.; Filardo, E.J. The G-protein coupled estrogen receptor, GPER: The inside and inside-out story. *Mol. Cell. Endocrinol.* **2015**, *418 Pt 3*, 207–219. [[CrossRef](#)]
8. Prossnitz, E.R.; Arterburn, J.B. International union of basic and clinical pharmacology. XC VII. G protein-coupled estrogen receptor and its pharmacologic modulators. *Pharmacol. Rev.* **2015**, *67*, 505–540. [[CrossRef](#)]
9. Barton, M.; Filardo, E.J.; Lolait, S.J.; Thomas, P.; Maggiolini, M.; Prossnitz, E.R. Twenty years of the G protein-coupled estrogen receptor GPER: Historical and personal perspectives. *J. Steroid Biochem. Mol. Biol.* **2018**, *176*, 4–15. [[CrossRef](#)]
10. Acconcia, F.; Bocedi, A.; Ascenzi, P.; Marino, M. Does palmitoylation target estrogen receptors to plasma membrane caveolae? *IUBMB Life* **2003**, *55*, 33–35. [[CrossRef](#)]
11. Manavathi, B.; Kumar, R. Steering estrogen signals from the plasma membrane to the nucleus: Two sides of the coin. *J. Cell. Physiol.* **2006**, *207*, 594–604. [[CrossRef](#)] [[PubMed](#)]
12. Heldring, N.; Pike, A.; Andersson, S.; Matthews, J.; Cheng, G.; Hartman, J.; Tujague, M.; Ström, A.; Treuer, E.; Warner, M.; et al. Estrogen receptors: How do they signal and what are their targets. *Physiol. Rev.* **2007**, *87*, 905–931. [[CrossRef](#)] [[PubMed](#)]
13. Mann, G.E.; Rowlands, D.J.; Li, F.Y.L.; de Winter, P.; Siow, R.C.M. Activation of endothelial nitric oxide synthase by dietary isoflavones: Role of NO in Nrf2-mediated antioxidant gene expression. *Cardiovasc. Res.* **2007**, *75*, 261–274. [[CrossRef](#)] [[PubMed](#)]
14. Vrtačnik, P.; Ostanek, B.; Mencej-Bedrač, S.; Marc, J. The many faces of estrogen signaling. *Biochem. Med.* **2014**, *24*, 329–342. [[CrossRef](#)]
15. Huang, B.; Warner, M.; Gustafsson, J.A. Estrogen receptors in breast carcinogenesis and endocrine therapy. *Mol. Cell. Endocrinol.* **2015**, *418*, 240–244. [[CrossRef](#)] [[PubMed](#)]
16. Kulkoyluoglu, E.; Madak-Erdogan, Z. Nuclear and extranuclear-initiated estrogen receptor signaling crosstalk and endocrine resistance in breast cancer. *Steroids* **2016**, *114*, 41–47. [[CrossRef](#)] [[PubMed](#)]
17. Siersbæk, R.; Kumar, S.; Carroll, J.S. Signaling pathways and steroid receptors modulating estrogen receptor  $\alpha$  function in breast cancer. *Genes Dev.* **2018**, *32*, 1141–1154. [[CrossRef](#)] [[PubMed](#)]
18. Itoh, K.; Chiba, T.; Takahashi, S.; Ishii, T.; Igarashi, K.; Katoh, Y.; Oyake, T.; Hayashi, N.; Satoh, K.; Hatayama, I.; Yamamoto, M.; et al. An Nrf2/small Maf heterodimer mediates the induction of phase II detoxifying enzyme genes through antioxidant response elements. *Biochem. Biophys. Res. Commun.* **1997**, *236*, 313–322. [[CrossRef](#)] [[PubMed](#)]
19. Ishii, T.; Itoh, K.; Takahashi, S.; Sato, H.; Yanagawa, T.; Katoh, Y.; Bannai, S.; Yamamoto, M. Transcription factor Nrf2 coordinately regulates a group of oxidative stress-inducible genes in macrophages. *J. Biol. Chem.* **2000**, *275*, 16023–16029. [[CrossRef](#)]
20. Mann, G.E.; Forman, H.J. Introduction to Special Issue on ‘Nrf2 regulated redox signaling and metabolism in physiology and medicine’. *Free Radic. Biol. Med.* **2015**, *88*, 91–92.
21. Murakami, S.; Motohashi, H. Roles of Nrf2 in cell proliferation and differentiation. *Free Radic. Biol. Med.* **2015**, *88*, 168–178. [[CrossRef](#)] [[PubMed](#)]

22. Homma, S.; Ishii, Y.; Morishima, Y.; Yamadori, T.; Matsuno, Y.; Haraguchi, N.; Kikuchi, N.; Satoh, H.; Sakamoto, T.; Hizawa, N.; et al. Nrf2 enhances cell proliferation and resistance to anticancer drugs in human lung cancer. *Clin. Cancer Res.* **2009**, *15*, 3423–3432. [[CrossRef](#)] [[PubMed](#)]
23. Lister, A.; Nedjadi, T.; Kitteringham, N.R.; Campbell, F.; Costello, E.; Lloyd, B.; Copple, I.M.; Williams, S.; Owen, A.; Neoptolemos, J.P.; et al. Nrf2 is overexpressed in pancreatic cancer: Implications for cell proliferation and therapy. *Mol. Cancer* **2011**, *10*, 37. [[CrossRef](#)]
24. Suzuki, T.; Motohashi, H.; Yamamoto, M. Toward clinical application of the Keap1-Nrf2 pathway. *Trends Pharmacol. Sci.* **2013**, *34*, 340–346. [[CrossRef](#)] [[PubMed](#)]
25. Syu, J.P.; Chi, J.T.; Kung, H.N. Nrf2 is the key to chemotherapy resistance in MCF7 breast cancer cells under hypoxia. *Oncotarget* **2016**, *7*, 14659–14672. [[CrossRef](#)] [[PubMed](#)]
26. Wu, J.; Williams, D.; Walter, G.A.; Thompson, W.E.; Sidell, N. Estrogen increases Nrf2 activity through activation of the PI3K pathway in MCF-7 breast cancer cells. *Exp. Cell Res.* **2014**, *328*, 351–360. [[CrossRef](#)] [[PubMed](#)]
27. Itoh, K.; Wakabayashi, N.; Katoh, Y.; Ishii, T.; Igarashi, K.; Engel, J.D.; Yamamoto, M. Keap1 represses nuclear activation of antioxidant responsive elements by Nrf2 through binding to the amino-terminal Neh2 domain. *Genes Dev.* **1999**, *13*, 76–86. [[CrossRef](#)]
28. Itoh, K.; Mimura, J.; Yamamoto, M. Discovery of the negative regulator of Nrf2, Keap1: A historical overview. *Antioxid. Redox Signal.* **2010**, *13*, 1665–1678. [[CrossRef](#)] [[PubMed](#)]
29. Kobayashi, M.; Li, L.; Iwamoto, N.; Nakajima-Takagi, Y.; Kaneko, H.; Nakayama, Y.; Eguchi, M.; Wada, Y.; Kumagai, Y.; Yamamoto, M. The antioxidant defense system Keap1-Nrf2 comprises a multiple sensing mechanism for responding to a wide range of chemical compounds. *Mol. Cell. Biol.* **2009**, *29*, 493–502. [[CrossRef](#)]
30. Rada, P.; Rojo, A.I.; Evrard-Todeschi, N.; Innamorato, N.G.; Cotte, A.; Jaworski, T.; Tobón-Velasco, J.C.; Devijver, H.; García-Mayoral, M.F.; Van Leuven, F.; et al. Structural and functional characterization of Nrf2 degradation by the glycogen synthase kinase 3/ $\beta$ -TrCP. *Mol. Cell. Biol.* **2012**, *32*, 3486–3499. [[CrossRef](#)]
31. Hayes, J.D.; Chowdhry, S.; Dinkova-Kostova, A.T.; Sutherland, C. Dual regulation of transcription factor Nrf2 by Keap1 and by the combined actions of  $\beta$ -TrCP and GSK-3. *Biochem. Soc. Trans.* **2015**, *43*, 611–620. [[CrossRef](#)]
32. Jain, A.K.; Jaiswal, A.K. GSK-3 $\beta$  acts upstream of Fyn kinase in regulation of nuclear export and degradation of NF-E2 related factor 2. *J. Biol. Chem.* **2007**, *282*, 16502–16510. [[CrossRef](#)] [[PubMed](#)]
33. Kaspar, J.W.; Jaiswal, A.K. Tyrosine phosphorylation controls nuclear export of Fyn, allowing Nrf2 activation of cytoprotective gene expression. *FASEB J.* **2011**, *25*, 1076–1087. [[CrossRef](#)] [[PubMed](#)]
34. Mathur, A.; Pandey, V.K.; Kakkar, P. Activation of GSK3 $\beta$ / $\beta$ -TrCP axis via PHLPP1 exacerbates Nrf2 degradation leading to impairment in cell survival pathway during diabetic nephropathy. *Free Radic. Biol. Med.* **2018**, *120*, 414–424. [[CrossRef](#)] [[PubMed](#)]
35. Huang, H.C.; Nguyen, T.; Pickett, C.B. Phosphorylation of Nrf2 at Ser-40 by protein kinase C regulates antioxidant response element-mediated transcription. *J. Biol. Chem.* **2002**, *277*, 42769–42774. [[CrossRef](#)] [[PubMed](#)]
36. Pi, J.; Bai, Y.; Reece, J.M.; Williams, J.; Liu, D.; Freeman, M.L.; Fahl, W.E.; Shugar, D.; Liu, J.; Qu, W.; et al. Molecular mechanism of human Nrf2 activation and degradation: Role of sequential phosphorylation by protein kinase CK2. *Free Radic. Biol. Med.* **2007**, *42*, 1797–1806. [[CrossRef](#)]
37. Apopa, P.L.; He, X.; Ma, Q. Phosphorylation of Nrf2 in the transcription activation domain by casein kinase 2 (CK2) is critical for the nuclear translocation and transcription activation function of Nrf2 in IMR-32 neuroblastoma cells. *J. Biochem. Mol. Toxicol.* **2008**, *22*, 63–76. [[CrossRef](#)] [[PubMed](#)]
38. Afonyuskin, T.; Oskolkova, O.V.; Binder, B.R.; Bochkov, V.N. Involvement of CK2 in activation of electrophilic genes in endothelial cells by oxidized phospholipids. *J. Lipid Res.* **2011**, *52*, 98–103. [[CrossRef](#)] [[PubMed](#)]
39. Ishii, T.; Warabi, E.; Mann, G.E. Circadian control of p75 neurotrophin receptor leads to alternate activation of Nrf2 and c-Rel to reset energy metabolism in astrocytes via brain-derived neurotrophic factor. *Free Radic. Biol. Med.* **2018**, *119*, 34–44. [[CrossRef](#)]
40. Ishii, T.; Mann, G.E. When and how does brain-derived neurotrophic factor activate Nrf2 in astrocytes and neurons? *Neural Regen. Res.* **2018**, *13*, 803–804. [[CrossRef](#)] [[PubMed](#)]
41. Ishii, T.; Warabi, E.; Mann, G.E. Circadian control of BDNF-mediated Nrf2 activation in astrocytes protects dopaminergic neurons from ferroptosis. *Free Radic. Biol. Med.* **2018**. [[CrossRef](#)]



42. Arevalo, M.A.; Rodriguez-Tébar, A. Activation of casein kinase II and inhibition of phosphatase and tensin homologue deleted on chromosome 10 phosphatase by nerve growth factor/p75<sup>NTR</sup> inhibit glycogen synthase kinase-3 $\beta$  and stimulate axonal growth. *Mol. Biol. Cell* **2006**, *17*, 3369–3377. [[CrossRef](#)]
43. Dobrowsky, R.T.; Werner, M.H.; Castellino, A.M.; Chao, M.V.; Hannun, Y.A. Activation of the sphingomyelin cycle through the low-affinity neurotrophin receptor. *Science* **1994**, *265*, 1596–1599. [[CrossRef](#)]
44. Müller, G.; Ayoub, M.; Storz, P.; Rennecke, J.; Fabbro, D.; Pfizenmaier, K. PKC $\zeta$  is a molecular switch in signal transduction of TNF- $\alpha$ , bifunctionally regulated by ceramide and arachidonic acid. *EMBO J.* **1995**, *14*, 1961–1969. [[CrossRef](#)]
45. Lee, Y.H.; Park, J.W.; Bae, Y.S. Regulation of protein kinase CK2 catalytic activity by protein kinase C and phospholipase D2. *Biochimie* **2016**, *121*, 131–139. [[CrossRef](#)]
46. Chamard-Jovenin, C.; Jung, A.C.; Chesnel, A.; Abecassis, J.; Flament, S.; Ledrappier, S.; Macabre, C.; Boukhobza, T.; Dumond, H. From ER $\alpha$ 66 to ER $\alpha$ 36: A generic method for validating a prognosis marker of breast tumor progression. *BMC Syst. Biol.* **2015**, *9*, 28. [[CrossRef](#)]
47. Acconcia, F.; Ascenzi, P.; Fabozzi, G.; Visca, P.; Marino, M. S-palmitoylation modulates human estrogen receptor- $\alpha$  functions. *Biochem. Biophys. Res. Commun.* **2004**, *316*, 878–883. [[CrossRef](#)]
48. Gu, W.; Dong, N.; Wang, P.; Shi, C.; Yang, J.; Wang, J. Tamoxifen resistance and metastasis of human breast cancer cells were mediated by the membrane-associated estrogen receptor ER- $\alpha$ 36 signaling in vitro. *Cell. Biol. Toxicol.* **2017**, *33*, 183–195. [[CrossRef](#)]
49. Schwartz, N.; Chaudhri, R.A.; Hadadi, A.; Schwartz, Z.; Boyan, B.D. 17 $\beta$ -estradiol promotes aggressive laryngeal cancer through membrane-associated estrogen receptor- $\alpha$  36. *Horm. Cancer* **2014**, *5*, 22–32. [[CrossRef](#)]
50. Levin, E.R. Bidirectional signaling between the estrogen receptor and the epidermal growth factor receptor. *Mol. Endocrinol.* **2003**, *17*, 309–317. [[CrossRef](#)]
51. Kadioglu, O.; Cao, J.; Saeed, M.E.; Greten, H.J.; Efferth, T. Targeting epidermal growth factor receptors and downstream signaling pathways in cancer by phytochemicals. *Target. Oncol.* **2015**, *10*, 337–353. [[CrossRef](#)]
52. Kyriakopoulou, K.; Kefali, E.; Piperigkou, Z.; Bassiony, H.; Karamanos, N.K. Advances in targeting epidermal growth factor receptor signaling pathway in mammary cancer. *Cell Signal.* **2018**, *51*, 99–109. [[CrossRef](#)]
53. Sigismund, S.; Avanzato, D.; Lanzetti, L. Emerging function of the EGFR in cancer. *Mol. Oncol.* **2018**, *12*, 3–20. [[CrossRef](#)]
54. Frank, G.D.; Mifune, M.; Inagami, T.; Ohba, M.; Sasaki, T.; Higashiyama, S.; Dempsey, P.J.; Eguchi, S. Distinct mechanisms of receptor and nonreceptor tyrosine kinase activation by reactive oxygen species in vascular smooth muscle cells: Role of metalloprotease and protein kinase C- $\delta$ . *Mol. Cell. Biol.* **2003**, *23*, 1581–1589. [[CrossRef](#)]
55. McCubrey, J.A.; Steelman, L.S.; Chappell, W.H.; Abrams, S.L.; Wong, E.W.; Chang, F.; Lehmann, B.; Terrian, D.M.; Milella, M.; Tafuri, A.; et al. Roles of the Raf/MEK/ERK pathway in cell growth, malignant transformation and drug resistance. *Biochem. Biophys. Acta* **2007**, *1773*, 1263–1284. [[CrossRef](#)]
56. Zhang, X.T.; Ding, L.; Kang, L.G.; Wang, Z.Y. Involvement of ER- $\alpha$ 36, Src, EGFR and STAT5 in the biphasic estrogen signaling of ER-negative breast cancer cells. *Oncol. Rep.* **2012**, *27*, 2057–2065.
57. Omarjee, S.; Jacquemetton, J.; Poulard, C.; Rochel, N.; Dejaegere, A.; Chebaro, Y.; Treilleux, I.; Marangoni, E.; Corbo, L.; Romancer, M.L. The molecular mechanisms underlying the ER $\alpha$ -36-mediated signaling in breast cancer. *Oncogene* **2017**, *36*, 2503–2514. [[CrossRef](#)]
58. Zou, W.; Fang, C.; Ji, X.; Liang, X.; Liu, Y.; Han, C.; Huang, L.; Zhang, Q.; Li, H.; Zhang, Y.; et al. Estrogen receptor (ER)- $\alpha$ 36 is involved in estrogen- and tamoxifen-induced neuroprotective effects in ischemic stroke models. *PLoS ONE* **2015**, *10*, e0140660. [[CrossRef](#)]
59. Garcia-Ovejero, D.; Azcoitia, I.; DonCarlos, L.L.; Melcangi, R.C.; Garcia-Segura, L.M. Glia-neuron crosstalk in the neuroprotective mechanisms of sex steroid hormones. *Brain Res. Rev.* **2005**, *48*, 273–286. [[CrossRef](#)]
60. Zhu, C.; Wang, S.; Wang, B.; Du, F.; Hu, C.; Li, H.; Feng, Y.; Zhu, R.; Mo, M.; Cao, Y.; et al. 17 $\beta$ -Estradiol up-regulates Nrf2 via PI3K/AKT and estrogen receptor signaling pathways to suppress light-induced degeneration in rat retina. *Neuroscience* **2015**, *304*, 328–339. [[CrossRef](#)]
61. Prossnitz, E.R.; Hathaway, H.J. What have we learned about GPER function in physiology and disease from knockout mice? *J. Steroid Biochem. Mol. Biol.* **2015**, *153*, 114–126. [[CrossRef](#)]
62. Vail, G.; Roepke, T.A. Membrane-initiated estrogen signaling via Gq-coupled GPCR in the central nervous system. *Steroids* **2018**. [[CrossRef](#)]

63. Tran, Q.K.; VerMeer, M.; Burgard, M.A.; Hassan, A.B.; Giles, J. Hetero-oligomeric complex between the G protein-coupled estrogen receptor 1 and the plasma membrane  $\text{Ca}^{2+}$ -ATPase 4b. *J. Biol. Chem.* **2015**, *290*, 13293–13307. [[CrossRef](#)]
64. Meyer, M.R.; Fredette, N.C.; Daniel, C.; Sharma, G.; Amann, K.; Arteburn, J.B.; Barton, M.; Prossnitz, E.R. Obligatory role for GPER in cardiovascular aging and disease. *Sci. Signal.* **2016**, *9*, ra105. [[CrossRef](#)]
65. Meyer, M.R.; Barton, M. GPER blockers as Nox downregulators: A new drug class to target chronic non-communicable diseases. *J. Steroid Biochem. Mol. Biol.* **2018**, *176*, 82–87. [[CrossRef](#)]
66. Brandes, R.P.; Schröder, K. Composition and functions of vascular nicotinamide adenine dinucleotide phosphate oxidases. *Trends Cardiovasc. Med.* **2008**, *18*, 15–19. [[CrossRef](#)]
67. Frey, R.S.; Ushio-Fukai, M.; Malik, A.B. NADPH oxidase-dependent signaling in endothelial cells: Role in physiology and pathophysiology. *Antioxid. Redox Signal.* **2009**, *11*, 791–810. [[CrossRef](#)]
68. Belarbi, K.; Cuvelier, E.; Destée, A.; Gressier, B.; Chartier-Harlin, M.-C. NADPH oxidases in Parkinson's disease: A systematic review. *Mol. Neurodegener.* **2017**, *12*, 84. [[CrossRef](#)]
69. Park, H.S.; Lee, S.H.; Park, D.; Ryu, S.H.; Rhee, S.G.; Bae, Y.S. Sequential activation of phosphatidylinositol 3-kinase, beta Pix, Rac1, and Nox1 in growth factor-induced production of  $\text{H}_2\text{O}_2$ . *Mol. Cell. Biol.* **2004**, *24*, 4384–4394. [[CrossRef](#)]
70. Kamata, T. Roles of Nox1 and other Nox isoforms in cancer development. *Cancer Sci.* **2009**, *100*, 1382–1388. [[CrossRef](#)]
71. Juhasz, A.; Markel, S.; Gaur, S.; Liu, H.; Lu, J.; Jiang, G.; Wu, X.; Antony, S.; Wu, Y.; Melillo, G.; et al. NADPH oxidase 1 supports proliferation of colon cancer cells by modulating reactive oxygen species-dependent signal transduction. *J. Biol. Chem.* **2017**, *292*, 7866–7887. [[CrossRef](#)] [[PubMed](#)]
72. Yamamoto, T.; Nakano, H.; Shiomi, K.; Wanibuchi, K.; Masui, H.; Takahashi, T.; Urano, Y.; Kamata, T. Identification and characterization of a novel NADPH oxidase 1 (Nox1) inhibitor that suppresses proliferation of colon and stomach cancer cells. *Biol. Pharm. Bull.* **2018**, *41*, 419–426. [[CrossRef](#)] [[PubMed](#)]
73. Wang, X.; Sun, Z. Thyroid hormone induces artery smooth muscle cell proliferation: Discovery of a new TRalpha1-Nox1 pathway. *J. Cell. Mol. Med.* **2010**, *14*, 368–380. [[CrossRef](#)] [[PubMed](#)]
74. Ranjan, P.; Anathy, V.; Burch, P.M.; Weirather, K.; Lambeth, J.D.; Heintz, N.H. Redox-dependent expression of cyclin D1 and cell proliferation by Nox1 in mouse lung epithelial cells. *Antioxid. Redox Signal.* **2006**, *8*, 1447–1459. [[CrossRef](#)] [[PubMed](#)]
75. Feldman, R.D.; Limbird, L.E. GPER (GPR30): A nongenomic receptor (GPER) for steroid hormones with implications for cardiovascular disease and cancer. *Annu. Rev. Pharmacol. Toxicol.* **2017**, *57*, 567–584. [[CrossRef](#)] [[PubMed](#)]
76. Choi, H.; Leto, T.L.; Hunyady, L.; Catt, K.J.; Bae, Y.S.; Rhee, S.G. Mechanism of angiotensin II-induced superoxide production in cells reconstituted with angiotensin type 1 receptor and the components of NADPH oxidase. *J. Biol. Chem.* **2008**, *283*, 255–267. [[CrossRef](#)]
77. Seta, K.; Nanamori, M.; Modrall, J.G.; Neubig, R.R.; Sadoshima, J. AT1 receptor mutant lacking heterotrimeric G protein coupling activates the Src-Ras-ERK pathway without nuclear translocation of ERKs. *J. Biol. Chem.* **2002**, *277*, 9268–9277. [[CrossRef](#)]
78. Seshiah, P.N.; Weber, D.S.; Rocic, P.; Valppu, L.; Taniyama, Y.; Griendling, K.K. Angiotensin II stimulation of NAD(P)H oxidase activity: Upstream mediators. *Circ. Res.* **2002**, *91*, 406–413. [[CrossRef](#)]
79. Streeter, J.; Schickling, B.M.; Jiang, S.; Stanic, B.; Thiel, W.H.; Gakhar, L.; Houtman, J.C.; Miller, F.J., Jr. Phosphorylation of Nox1 regulates association with NoxA1 activation domain. *Circ. Res.* **2014**, *115*, 911–918. [[CrossRef](#)]
80. Zaldua, N.; Gastineau, M.; Hoshino, M.; Lezoualch, F.; Zugaza, J.L. Epac signaling pathway involves STEF, a guanine nucleotide exchange factor for Rac, to regulate APP processing. *FEBS Lett.* **2007**, *581*, 5814–5818. [[CrossRef](#)]
81. Nishimoto, Y.; Tsubouchi, R.; Diebold, B.A.; Qiao, S.; Ogawa, H.; Ohara, T.; Tamura, M. Activation of NADPH oxidase 1 in tumor colon epithelial cells. *Biochem. J.* **2008**, *415*, 57–65. [[CrossRef](#)] [[PubMed](#)]
82. Lei, B.; Huan, Y.; Liu, Y.; Xu, J.; Sun, S.; Zhang, X.; Xu, G.; Wu, M.; Yu, Y.; Feng, C. Low-concentration BPF induced cell biological responses by the  $\text{ER}\alpha$  and GPER1-mediated signaling pathways in MCF-7 breast cancer cells. *Ecotoxicol. Environ. Saf.* **2018**, *165*, 144–152. [[CrossRef](#)] [[PubMed](#)]

83. Lei, B.; Sun, S.; Zhang, X.; Feng, C.; Xu, J.; Wen, Y.; Huang, Y.; Wu, M.; Yu, Y. Bisphenol AF exerts estrogenic activity in MCF-7 cells through activation of Erk and PI3K/Akt signals via GPER signaling pathway. *Chemosphere* **2019**, *220*, 362–370. [[CrossRef](#)] [[PubMed](#)]
84. Pelekanou, V.; Kampa, M.; Kiagiadaki, F.; Deli, A.; Theodoropoulos, P.; Agrogiannis, G.; Patsouris, E.; Tsapis, A.; Castanas, E.; Notas, G. Estrogen anti-inflammatory activity on human monocytes is mediated through cross-talk between estrogen receptor ER $\alpha$ 36 and GPR30/GPER1. *J. Leukoc. Biol.* **2016**, *99*, 333–347. [[CrossRef](#)]
85. Kang, L.; Zhang, X.; Xie, Y.; Tu, Y.; Wang, D.; Liu, Z.; Wang, Z.Y. Involvement of estrogen receptor variant ER- $\alpha$ 36, not GPR30, in nongenomic estrogen signaling. *Mol. Endocrinol.* **2010**, *24*, 709–721. [[CrossRef](#)] [[PubMed](#)]
86. Wallacides, A.; Chesnel, A.; Ajj, H.; Chillet, M.; Flament, S.; Dumond, H. Estrogen promote proliferation of the seminoma-like TCam-2 cell line through a GPER-dependent ER $\alpha$ 36 induction. *Mol. Cell. Endocrinol.* **2012**, *350*, 61–71. [[CrossRef](#)] [[PubMed](#)]
87. Sheng, Z.G.; Huang, W.; Liu, Y.X.; Zhu, B.Z. Bisphenol A at a low concentration boosts mouse spermatogonial cell proliferation by inducing the G protein-coupled receptor 30 expression. *Toxicol. Appl. Pharmacol.* **2013**, *267*, 88–94. [[CrossRef](#)] [[PubMed](#)]
88. Fisher, A.B. Redox signaling across cell membrane. *Antioxid. Redox Signal.* **2009**, *11*, 1349–1356. [[CrossRef](#)]
89. Yang, H.; Ma, L.; Wang, Y.; Zuo, W.; Li, B.; Yang, Y.; Chen, Y.; Chen, L.; Wang, L.; Zhu, L. Activation of CIC-3 chloride channels by 17 $\beta$ -estradiol relies on the estrogen receptor  $\alpha$  expression in breast cancer. *J. Cell. Physiol.* **2018**, *233*, 1071–1081. [[CrossRef](#)] [[PubMed](#)]
90. Zhou, F.M.; Huang, Y.Y.; Tian, T.; Li, X.Y.; Tang, Y.B. Knockdown of chloride channel-3 inhibits breast cancer growth in vitro and in vivo. *J. Breast Cancer* **2018**, *21*, 103–111. [[CrossRef](#)]
91. Hecht, D.; Zick, Y. Selective inhibition of protein tyrosine phosphatase activities by H<sub>2</sub>O<sub>2</sub> and vanadate in vitro. *Biochem. Biophys. Res. Commun.* **1992**, *188*, 773–779. [[CrossRef](#)]
92. Cheng, G.; Lambeth, J.D. NOXO1, regulation of lipid binding, localization, and activation of Nox1 by the Phox homology (PX) domain. *J. Biol. Chem.* **2004**, *279*, 4737–4742. [[CrossRef](#)]
93. Wu, Y.; Feinstein, S.I.; Manevich, Y.; Chowdhury, I.; Pak, J.H.; Kazi, A.; Dodia, C.; Speicher, D.W.; Fisher, A.B. Mitogen-activated protein kinase-mediated phosphorylation of peroxiredoxin 6 regulates its phospholipase A<sub>2</sub> activity. *Biochem. J.* **2009**, *419*, 669–679. [[CrossRef](#)]
94. Bieberich, E. Ceramide signaling in cancer and stem cells. *Future Lipidol.* **2008**, *3*, 273–300. [[CrossRef](#)]
95. Liu, B.; Hannun, Y.A. Inhibition of the neutral magnesium-dependent sphingomyelinase by glutathione. *J. Biol. Chem.* **1997**, *271*, 16281–16287. [[CrossRef](#)]
96. Bannai, S.; Ishii, T. Transport of cystine and cysteine and cell growth in cultured human diploid fibroblasts: Effect of glutamate and homocysteate. *J. Cell. Physiol.* **1982**, *112*, 265–271. [[CrossRef](#)]
97. Richman, P.G.; Meister, A. Regulation of  $\gamma$ -glutamyl-cysteine synthetase by nonallosteric feedback inhibition by glutathione. *J. Biol. Chem.* **1975**, *250*, 1422–1426.
98. Winterbourn, C.C.; Metodiewa, D. The reaction of superoxide with reduced glutathione. *Arch. Biochem. Biophys.* **1994**, *14*, 284–290. [[CrossRef](#)]
99. Minich, T.; Riemer, J.; Schulz, J.B.; Wielinga, P.; Wijnholds, J.; Dringen, R. The multidrug resistance protein (Mrp1), but not Mrp5, mediates export of glutathione and glutathione disulfide from brain astrocytes. *J. Neurochem.* **2006**, *97*, 373–384. [[CrossRef](#)]
100. Clarke, C.J.; Guthrie, J.M.; Hannun, Y.A. Regulation of neutral sphingomyelinase-2 (nSMase2) by tumor necrosis factor- $\alpha$  involves protein kinase C- $\delta$  in lung epithelial cells. *Mol. Pharmacol.* **2008**, *74*, 1022–1032. [[CrossRef](#)]
101. Chung, S.; Vu, S.; Filosto, S.; Goldkorn, T. Src regulates cigarette smoke-induced ceramide generation via neutral sphingomyelinase 2 in the airway epithelium. *Am. J. Respir. Cell. Mol. Biol.* **2015**, *52*, 738–748. [[CrossRef](#)]
102. Gu, L.; Huang, B.; Shen, W.; Gao, L.; Ding, Z.; Wu, H.; Guo, J. Early activation of nSMase2/ceramide pathway in astrocytes is involved in ischemia-associated neuronal damage via inflammation in rat hippocampi. *J. Neuroinflamm.* **2013**, *10*, 109. [[CrossRef](#)]
103. Filosto, S.; Fry, W.; Knowlton, A.A.; Goldkorn, T. Neutral sphingomyelinase 2 (nSMase2) is a phosphoprotein regulated by calcineurin (PP2B). *J. Biol. Chem.* **2010**, *285*, 10213–10222. [[CrossRef](#)]

104. Ferri, A.; Gabbianelli, R.; Casciati, A.; Paolucci, E.; Rotilio, G.; Carri, M.T. Calcineurin activity is regulated both by redox compounds and by mutant familial amyotrophic lateral sclerosis-superoxide dismutase. *J. Neurochem.* **2000**, *75*, 606–613. [[CrossRef](#)]
105. Wang, X.; Culotta, V.C.; Klee, C.B. Superoxide dismutase protects calcineurin from inactivation. *Nature* **1996**, *383*, 434–437. [[CrossRef](#)]
106. Namgaladze, D.; Shcherbyna, I.; Kienhöfer, J.; Hofer, H.W.; Ullrich, V. Superoxide targets calcineurin signaling in vascular endothelium. *Biochem. Biophys. Res. Commun.* **2005**, *334*, 1061–1067. [[CrossRef](#)]
107. Jang, E.J.; Jeong, H.O.; Park, D.; Kim, D.H.; Choi, Y.J.; Chung, K.W.; Park, M.H.; Yu, B.P.; Chung, H.Y. Src tyrosine kinase activation by 4-hydroxynoneal upregulates p38, ERK/AP-1 signaling and COX-2 expression in YPEN-1 cells. *PLoS ONE* **2015**, *10*, e0129244. [[CrossRef](#)]
108. Benes, C.; Soltoff, S.P. Modulation of PKC $\delta$  tyrosine phosphorylation and activity in salivary and PC-12 cells by Src kinases. *Am. J. Physiol. Cell Physiol.* **2001**, *280*, C1498–C1510. [[CrossRef](#)]
109. Sawada, M.; Nakashima, S.; Kiyono, T.; Nakagawa, M.; Yamada, J.; Yamakawa, H.; Banno, Y.; Shinoda, J.; Nishimura, Y.; Nozawa, Y.; et al. p53 regulates ceramide formation by neutral sphingomyelinase through reactive oxygen species in human glioma cells. *Oncogene* **2001**, *20*, 1368–1378. [[CrossRef](#)]
110. Jana, A.; Pahan, A. Fibrillar amyloid- $\beta$  peptides kill human primary neurons via NADPH oxidase-mediated activation of neutral sphingomyelinase: Implications for Alzheimer's disease. *J. Biol. Chem.* **2004**, *279*, 51451–51459. [[CrossRef](#)]
111. Zhang, Y.; Yao, B.; Delikat, S.; Bayoumy, S.; Lin, X.H.; Basu, S.; McGinley, M.; Chan-Hui, P.Y.; Lichenstein, H.; Kolesnick, R. Kinase suppressor of Ras is ceramide-activated protein kinase. *Cell* **1997**, *89*, 63–72. [[CrossRef](#)]
112. Yin, X.; Zafrullah, M.; Lee, H.; Haimovitz-Friedman, A.; Fuks, Z.; Kolesnick, R. A ceramide-binding C1 domain mediates kinase suppressor of Ras membrane translocation. *Cell. Physiol. Biochem.* **2009**, *24*, 219–230. [[CrossRef](#)]
113. Peng, C.H.; Huang, C.N.; Hsu, S.P.; Wang, C.J. Pent-acetyl geniposide induce apoptosis in C6 glioma cells by modulating the activation of neutral sphingomyelinase-induced p75 nerve growth factor receptor and protein kinase C $\delta$  pathway. *Mol. Pharmacol.* **2006**, *70*, 997–1004. [[CrossRef](#)]
114. Lin, W.L.; Liang, W.H.; Lee, Y.J.; Chuang, S.K.; Tseng, T.H. Antitumor progression potential of caffeic acid phenethyl ester involving p75<sup>NTR</sup> in C6 glioma cells. *Chem. Biol. Interact.* **2010**, *188*, 607–615. [[CrossRef](#)]
115. Liu, J.; Yin, F.; Zheng, X.; Jing, J.; Hu, Y. Geniposide, a novel agonist for GLP-1 receptor, prevents PC12 cells from oxidative damage via MAP kinase pathway. *Neurochem. Int.* **2007**, *51*, 361–369. [[CrossRef](#)]
116. Buteau, J.; Foisy, S.; Joly, E.; Prentki, M. Glucagon-like peptide 1 induces pancreatic  $\beta$ -cell proliferation via transactivation of the epidermal growth factor receptor. *Diabetes* **2003**, *52*, 124–132. [[CrossRef](#)]
117. Fernández-Millán, E.; Martín, M.A.; Goya, L.; Lizárraga-Mollinedo, E.; Escrivá, F.; Ramos, S.; Álvarez, C. Glucagon-like peptide-1 improves  $\beta$ -cell antioxidant capacity via extracellular regulated kinases pathway and Nrf2 translocation. *Free Radic. Biol. Med.* **2016**, *95*, 16–26. [[CrossRef](#)] [[PubMed](#)]
118. Yin, F.; Liu, J.; Zheng, X.; Guo, L.; Xiao, H. Geniposide induces the expression of hemeoxygenase-1 via PI3K/Nrf3-signaling to enhance the antioxidant capacity in primary hippocampal neurons. *Biol. Pharm. Bull.* **2010**, *33*, 1841–1846. [[CrossRef](#)]
119. Jeon, W.K.; Hong, H.Y.; Kim, B.C. Genipin up-regulates heme oxygenase-1 via PI3-kinase-JNK1/2-Nrf2 signaling pathway to enhance the anti-inflammatory capacity in RAW264.7 macrophages. *Arch. Biochem. Biophys.* **2011**, *512*, 119–125. [[CrossRef](#)]
120. Balogun, E.; Hogue, M.; Gong, P.; Killeen, E.; Green, C.J.; Foresti, R.; Alam, J.; Motterini, R. Curcumin activates the haem oxygenase-1 gene via regulation of Nrf2 and the antioxidant-responsive element. *Biochem. J.* **2003**, *371*, 887–895. [[CrossRef](#)]
121. Morroni, F.; Sita, G.; Graziosi, A.; Turrini, E.; Fimognari, C.; Tarozzi, A.; Hrelia, P. Neuroprotective effect of caffeic acid phenethyl ester in a mouse model of Alzheimer's disease involves Nrf2/HO-1 pathway. *Aging Dis.* **2018**, *9*, 605–622. [[CrossRef](#)] [[PubMed](#)]
122. Jung, B.I.; Kim, M.S.; Kim, H.A.; Kim, D.; Yang, J.; Her, S.; Song, Y.S. Caffeic acid phenethyl ester, a component of beehive propolis, is a novel selective estrogen receptor modulator. *Phytother. Res.* **2010**, *24*, 295–300. [[CrossRef](#)]
123. Miyata, Y.; Nishida, E. CK2 controls multiple protein kinases by phosphorylating a kinase-targeting molecular chaperone, Cdc37. *Mol. Cell. Biol.* **2004**, *24*, 4065–4074. [[CrossRef](#)]

124. Al Quobaili, F.; Montenarh, M. CK2 and the regulation of the carbohydrate metabolism. *Metabolism* **2012**, *61*, 1512–1517. [[CrossRef](#)]
125. Hunter, T.; Poon, R.Y.C. Cdc37: A protein kinase chaperone? *Trends Cell Biol.* **1997**, *7*, 157–161. [[CrossRef](#)]
126. MacLean, M.; Picard, D. Cdc37 goes beyond Hsp90 and kinases. *Cell Stress Chaperones* **2003**, *8*, 114–119. [[CrossRef](#)]
127. Warabi, E.; Takabe, W.; Minami, T.; Inoue, K.; Itoh, K.; Yamamoto, M.; Ishii, T.; Kodama, T.; Noguchi, N. Shear stress stabilizes NF-E2-related factor 2 and induces antioxidant genes in endothelial cells: Role of reactive oxygen/nitrogen species. *Free Radic. Biol. Med.* **2007**, *42*, 260–269. [[CrossRef](#)]
128. Czarny, M.; Liu, J.; Oh, P.; Schnitzer, J.E. Transient mechanoactivation of neutral sphingomyelinase in caveolae to generate ceramide. *J. Biol. Chem.* **2003**, *278*, 4424–4430. [[CrossRef](#)]
129. Czaeny, M.; Schnitzer, J.E. Neutral sphingomyelinase inhibitor scyphostatin prevents and ceramide mimics mechanotransduction in vascular endothelium. *Am. J. Physiol. Heart Circ. Physiol.* **2004**, *287*, H1344–H1352.
130. Joy, S.; Siow, R.C.; Rowlands, D.J.; Becker, M.; Wyatt, A.W.; Aaronson, P.I.; Coen, C.W.; Kallo, I.; Jacob, R.; Mann, G.E. The isoflavone Equol mediates rapid vascular relaxation: Ca<sup>2+</sup>-independent activation of endothelial nitric-oxide synthase/Hsp90 involving ERK1/2 and Akt phosphorylation in human endothelial cells. *J. Biol. Chem.* **2006**, *281*, 27335–27345. [[CrossRef](#)]
131. Thomas, P.; Dong, J. Binding and activation of the seven-transmembrane estrogen receptor GPR30 by environmental estrogens: A potent novel mechanism of endocrine disruption. *J. Steroid Biochem. Mol. Biol.* **2006**, *102*, 175–179. [[CrossRef](#)]
132. Yu, X.; Filardo, E.J.; Shaikh, Z.A. The membrane estrogen receptor GPR30 mediates cadmium-induced proliferation of breast cancer cells. *Toxicol. Appl. Pharmacol.* **2010**, *245*, 83–90. [[CrossRef](#)]
133. Huff, M.O.; Todd, S.L.; Smith, A.L.; Elpers, J.T.; Smith, A.P.; Murphy, R.D.; Bleser-Shartzner, A.S.; Hoerter, J.E.; Radde, B.N.; Klinge, C.M. Arsenite and cadmium activate MAPK/ERT via membrane estrogen receptors and G-protein coupled estrogen receptor signaling in human lung adenocarcinoma cells. *Toxicol. Sci.* **2016**, *152*, 62–71. [[CrossRef](#)]
134. Zhu, P.; Liao, L.Y.; Zhao, T.T.; Mo, X.M.; Chen, G.G.; Liu, Z.M. GPER/ERK&AKT/NF-κB pathway is involved in cadmium-induced proliferation, invasion and migration of GPER-positive thyroid cancer cells. *Mol. Cell. Endocrinol.* **2017**, *442*, 68–80. [[PubMed](#)]
135. Prins, G.S.; Hu, W.Y.; Xie, L.; Shi, G.B.; Hu, D.P.; Birch, L.; Bosland, M.C. Evaluation of bisphenol A (BPA) exposures on prostate stem cell homeostasis and prostate cancer risk in the NCTR-Sprague-Dawley rat: An NIEHS/FDA CLARITY-BPA consortium study. *Environ. Health Perspect.* **2018**, *126*, 117001. [[CrossRef](#)]
136. Shafei, A.; Ramzy, M.M.; Hegazy, A.I.; Husseny, A.K.; El-Hadary, U.G.; Taha, M.M.; Mosa, A.A. The molecular mechanisms of action of the endocrine disrupting chemical bisphenol A in the development of cancer. *Gene* **2018**, *647*, 235–243. [[CrossRef](#)]

