

# Inflammatory and degranulation effect of yellow sand on RBL-2H3 cells in relation to chemical and biological constituents

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1 **Inflammatory and degranulation effects of yellow sand on RBL-2H3 cells in**

2 **relation to chemical and biological constituents**

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28

29

30 **ABSTRACT**

31

32 Recent studie pointed out that allergic diseases have increased during the Asian dust storm  
33 event (ADSE) in Japan. Daily observations and the atmospheric concentrations of yellow sand  
34 (YS) aerosol have been increasing. In this study, YS samples collected from three sites of  
35 Japan during ADSE in 2009-2010 were used. The particles were analyzed by X-ray  
36 photoelectron spectroscopy (XPS) and X-ray fluorescence - energy dispersive spectrometer  
37 (XRF-EDS). We investigate ability of YS extract on enhancing the chemical mediator release  
38 and cytokine production from rat basophilic leukemia (RBL-2H3) cells. The dust particles at  
39 Fukuoka and Tsukuba were abundant in aluminum (Al), iron (Fe), potassium (K) and titan (Ti)  
40 than those at Naha. Concentration of the trace endotoxin and *Cryptomeria japonica* pollen  
41 allergen (Cry j 1) were measured in YS extract. After exposure of RBL-2H3 cells to YS  
42 extract, the  $\beta$ -hexosaminidase ( $\beta$ -hex) release, tumor necrosis factor-alpha (TNF- $\alpha$ ) production  
43 were enhanced in RBL-2H3 cells. This process depends on endotoxin, Cry j 1 and other  
44 allergen present in the YS extract. YS water extract also show a strong cytotoxic effect on the  
45 cells. This data suggest that low levels of endotoxin and Cry j 1 in YS may cause allergy  
46 during the ADSE.

47

48 ***Keywords***

49 Allergen, Asian dust storm,  $\beta$ -hexosaminidase, RBL-2H3, Yellow sand

50

51 ***Abbreviations***

52 ADSE, Asian dust storm event;  $\beta$ -hex,  $\beta$ -hexosaminidase; Cry j 1, *Cryptomeria japonica* pollen

53 allergen; Fuku., Fukuoka; LPS, Lipopolysaccharide; MTT, 3-(4, 5-dimethylthiazolyl)-2,  
54 5-diphenyltetrazolium bromide; PM, Particulate matter; RBL-2H3, Rat basophilic leukemia  
55 cells; TNF- $\alpha$ , Tumor necroses factor- $\alpha$ ; Tsu., Tsukuba; YS, Yellow sand.

56

57 **1. Introduction**

58

59 The prevalence and morbidity of asthma and other allergic diseases have increased  
60 dramatically during the last 30 years, particularly in industrial countries (Narita et al., 2007). A  
61 lot of studies have demonstrated that allergens commonly associated with dust storms include  
62 fungal spores, plant and grass pollens, anthropogenic emissions, and organic detritus (Griffin  
63 2007). Recently, it was clarified that dust event enables atmospheric long distance transport of  
64 bacteria, fungi, viruses, pollen etc. (Kellogg and Griffin 2006; Griffin 2010). Risk of  
65 hospitalization was increased during the Sahara dust event in European cities (Middleton et al.,  
66 2008). There was an association between the increase in paediatric asthma admissions and the  
67 increased Saharan dust event in Caribbean island (Gyan et al., 2005). Recently, daily  
68 observations and the atmospheric concentrations of YS aerosol have been increasing steadily in  
69 the eastern Asia region, including Japan (Goto et al., 2010). YS increase the risk of mortality  
70 and hospitalization in cardiovascular and respiratory patients during the ADSE in China, Korea,  
71 and Japan (Kwon et al., 2002; Meng and Lu 2007). Total fungi and particulate matter (PM)  
72 had significantly higher concentrations during the period affected by ADSE in Taiwan (Ho et al.,  
73 2005). In Japan, it was pointed out that fine YS particles make people's allergy more severe  
74 from February to May. Hua et al. (2007) have detected some bacteria, such as *Bacillus subtilis*,  
75 *B. licheniformis* in YS collected in Hiroshima, Japan during ADSE, and provided evidence that  
76 ADSE can transport these microorganisms in Northeast Asia.

77 Endotoxin, the lipopolysaccharide (LPS) components of the outer membrane of  
78 Gram-negative bacteria, has been associated with an increase in asthma symptoms, asthma

79 medications, and reductions in lung function in patients with atopic or asthma. Airborne  
80 endotoxin appears to be a risk factor for clinically symptomatic respiratory illnesses (Dales et al.,  
81 2006) and allergic inflammation (Imrich et al., 2000; Gon et al., 2005), and induces or  
82 aggravates a variety of respiratory diseases (Tulic et al., 2000). On the other hand, in recent  
83 decades in Japan, the number of Japanese cedar (*Cryptomeria Japonica*, JC) hay fever patients  
84 has increased and an increase in JC pollen is a probable principal cause (Sakurai et al., 2002).  
85 The pollen grains of JC pollen (*Cry j 1*) usually exist as coarse particles about 30  $\mu\text{m}$ .  
86 However, the conglomeration of dust-storm particles with sea salt forms larger hygroscopic  
87 particles during the long distance transport (Ma et al., 2004). Suspended PM (SPM) is present  
88 in the aqueous phase and induces the release of allergen particles (Wang et al., 2012). It was  
89 supposed that the major allergen *Cry j 1* could be release to the atmosphere as respirable-sized  
90 particles and modified by some air pollutants during airborne transportation (Wang et al.,  
91 2009a).

92 An increase in ambient antigen is one of the most probable reasons for the increase in  
93 allergic disorders such as allergic rhinitis, bronchial asthma, and atopic dermatitis. Allergic  
94 diseases are immunologic disorders, traditionally referred to as immediate or type I  
95 hypersensitivity reactions with IgE playing an important role. Crosslinking of the Fc $\epsilon$ RI  
96 (high-affinity IgE receptor) induced by complex formation of IgE with an antigenic protein is an  
97 essential event in the IgE-mediated allergic reaction (Beaven and Metzger 1993), The  
98 interaction of IgE with allergen on mast cells or basophils leads to allergic reactions causing the  
99 release of an array of inflammatory mediators resulting in the inflammation of airway mucus  
100 membrane leading to clinical symptoms in the target organ (Novak et al. 2001). Mast cells and

101 basophils play essential roles in the pathogenesis of allergic reactions and in protection against  
102 Gram-negative bacteria. RBL-2H3 cells, a tumor analog of mast cells, display characteristics  
103 of mucosal-type mast cells and express several hundred thousand FcεRI on the membrane  
104 surface. After sensitization with IgE, the cells respond to the antigen and release histamine.  
105 β-Hex, which is stored in the secretory granules of mast cells is released concomitantly with  
106 histamine when mast cells are immunologically activated (Ortega et al. 1988; Schroeder et al.  
107 1995). Thus, β-hex activity in the medium is used as a marker of mast cell degranulation  
108 (Yamada et al., 2010). Therefore, RBL-2H3 cells are considered as a good tool for studying  
109 the effect of environmental pollutant on chemical mediator release activity.

110       Recent studies regarding the impact of microbiological factors contained in YS on the air  
111 passage by Yanagisawa et al. (2007) and Ichinose et al. (2008) have indicated that YS has the  
112 potential of exacerbating symptoms in the human respiratory apparatus, eyes, and nose. Even  
113 though there are few reports concerning the biological activity of particular dust on macrophage  
114 and rat lung (Becker et al., 1996; Kim et al., 2003; Meng and Zhang 2007), and pulmonary  
115 toxicity and inflammatory allergy induced by administered of YS in mice and guinea pigs  
116 (Ichinose et al., 2005; 2009), there is no experimental evidence available regarding YS  
117 extract-mediated chemical mediator release by immediate type allergic model cells collected  
118 during ADSE in Japan. In this study, YS was collected from Naha, Fukuoka, and Tsukuba  
119 during ADSE in 2009 and 2010 of Japan, the chemical composition of YS was analyzed, and  
120 the endotoxin and Cry j 1 concentrations in the YS extracts were investigated. Furthermore,  
121 degranulation, cytokine production activity, and the cytotoxicity of YS extract on RBL-2H3  
122 cells were examined.

123

## 124 **2. Materials and methods**

125

### 126 *2.1. Chemicals and cells*

127

128 LPS was obtained from Alexis Biochemical (Enzo Life Sciences, Inc., San Diego, CA).

129 Fetal bovine serum (FBS) was obtained from Gibco BRL, Paisley, Scotland. The

130 enzyme-linked immunosorbent assays (ELISA) for analysis of rat TNF- $\alpha$  and IL-13 were

131 obtained from Invitrogen Co., CA. USA. 3-(4, 5-dimethylthiazol-2-yl)-2,

132 5-diphenyl-2H-tetrazolium bromide (MTT) was obtained from Dojindo, Japan. RBL-2H3

133 cells were purchased from JCRB Cell Bank, Japan. The cells were maintained in MEM

134 supplemented with 10%FBS and 2 mM L-glutamine, and incubated at 37°C in a 5%CO<sub>2</sub> .

135

### 136 *2.2. Sampling and sample location*

137

138 YS samples were obtained using a high volume sampler (HV-1000F, Shibata Science,

139 Tokyo, Japan) set on the open top floor of building (15-20 m) during March to June, 2009-2010

140 based on the information on the YS storm movement reported by the Japan Meteorological

141 Agency. Wind with aerosols including sands was sucked into sampler at a rate of 1000 L/min

142 and the aerosols that adhered onto the filter (Quartz Fiber Filters, Adventec QR-100, Toyo Roshi

143 Kaisha, Ltd.), which was set in the sampler were obtained. Three samples collected in 2009

144 and three samples collected in 2010, from Naha city (127°40'44"E, 26°12'44"N), Okinawa



145 prefecture (named Naha09 and Naha10, collected during April 27<sup>th</sup> to June 10<sup>th</sup> in 2009 and  
146 March 10<sup>th</sup> to April 14<sup>th</sup> in 2010), Fukuoka city (130°24'06"E, 33°35'24"N), Fukuoka prefecture  
147 (samples Fuku.09 and Fuku.10, collected during March 16<sup>th</sup> to 19<sup>th</sup> in 2009 and March 20<sup>th</sup> to  
148 24<sup>th</sup> in 2010 ) and Tsukuba city (140°10'37"E, 36°14'00"N), Ibaraki prefecture (samples Tsu.09  
149 and Tsu.10, collected during March 16<sup>th</sup> to 20<sup>th</sup> in 2009 and March 19<sup>th</sup> to 23<sup>th</sup> in 2010), Japan  
150 (Fig. 1), during the ADSE (MEGJ, 2010a) were used in this study. The sampling period,  
151 particle concentration, and particle density are shown in Table 1. The first sampling site in the  
152 city of Tsukuba (approximately 0.2 million inhabitants a population density of 284.07  
153 inhabitants/km<sup>2</sup>) is located in an industrialized area near Tokyo, and the samples from Fukuoka  
154 City, Fukuoka prefecture (approximately 1.45 million inhabitants, 341.32 inhabitants/km<sup>2</sup>)  
155 situated in an industrialized area near the Japanese sea, whereas the samples from Naha City,  
156 Okinawa prefecture (approximately 0.3 million inhabitants, 39.24 inhabitants/km<sup>2</sup>) represented a  
157 rural site. The geographical distances between Naha and Fukuoka sampling sites is about 860  
158 km, while Fukuoka and Tsukuba sampling site about 940 km.

159

### 160 *2.3. Analysis of sand components*

161

162 The collected samples were separated carefully from the filters and subjected to component  
163 analysis. First, the chemical components on the upper most 5 nm surface of the sands were  
164 analyzed by XPS with ESCA-300 (VG Scienta AB, Uppsala, Sweden). Then this surface  
165 layer was separated from bulk sands. The components of bulk sands were analyzed by  
166 XRF-EDS with JSX-3220 (JEOL Ltd., Tokyo, Japan).

167

168 *2.4. Extraction of LPS from YS sample*

169

170 The filters used to collect the dust samples were cut out and transferred into 15-ml Costar  
171 plastic tubes with 2 mL distilled MilliQ water. The tubes were then placed in an ultrasonic bath  
172 for 30 min. Then centrifugation was carried out at  $1000 \times g$  for 20 min at room temperature,  
173 and the supernatant (called water extract) were used for the determination of the endotoxin  
174 concentration,  $\beta$ -hex release, cytokine assay, and MTT assay.

175

176 *2.5. Extraction of Cry j 1 from YS sample*

177

178 The filters used to collect dust samples were cut out and transferred into 15-mL Costar  
179 plastic tubes with 2 mL Cry j1 extract buffer of TAC® Cry j 1 ELISA Kit (AKCJ1-010,  
180 Shibayagi, Gunma, Japan). After standing the solution in 4 °C for 2 h, shaking was carried out  
181 at room temperature for 30 min. Centrifugation was then carried out at  $1200 \times g$  for 30 min at  
182 room temperature, and the supernatant (called Cry j 1 extract) was used as the samples for the  
183 pollen allergen Cry j 1 determination and the  $\beta$ -hex release assay after filter-sterilization (0.45  
184  $\mu\text{m}$  Millipore filter).

185

186 *2.6. LPS determination*

187

188 Endotoxin in the water extract was measured by a quantitative kinetic chromogenic

189 Limulus Amoebocyte Lysate (LAL) method using Endospecy ES-50M (Seikagaku Co., Tokyo,  
190 Japan) at 37°C. Analyses were performed with an automated microtitre plate reader (Power  
191 Scan HT; Dainippon Pharmaceutical Co., Ltd.). Standard Endotoxin CSE-Kit (Seikagaku Co.,  
192 Tokyo, Japan) was used to determine the amount of endotoxin present in the sample. The  
193 detection limit was 0.002 EU/ml. The endotoxin concentration in each sample was calculated  
194 using a standard curve, and expressed as units per milliliters of sample (EU/mL). Endotoxin  
195 concentrations were reported as endotoxin units (EU) per mL of extract where 10 EU is  
196 equivalent to 1.25 ng of the reference standard endotoxin.

197

#### 198 *2.7. Cry j 1 determination*

199

200 Cry j 1 concentration in the sample was quantified by an ELISA method using TAC® Cry j  
201 1 ELISA Kit according to the manufacturer's instructions. Finally, the absorbance of each  
202 solution obtained by ELISA was converted into Cry j 1 concentration (pg/mL) of the Cry j 1  
203 extract.

204

#### 205 *2.8. Degranulation assay*

206

207 The  $\beta$ -hex release assay using RBL-2H3 cells was performed as previously described  
208 (Yamada et al., 2010). RBL-2H3 cells were seeded onto 96-well plates at  $5.0 \times 10^4$  cells/well in  
209 100  $\mu$ L of medium. Cells were incubated for 24 h at 37°C, then washed twice with PBS (-) to  
210 eliminate medium, and then the cells were exposed to various concentrations of water extract (0,

211 1/125, 1/100, 1/75, 1/50 dilution in medium) for 16 h at 37°C, 5%CO<sub>2</sub> incubator. As positive  
212 and negative controls, 5.0 ng/mL LPS and PBS (-) were used respectively. For Cry j 1  
213 extract-treated cells, cells were sensitized with 0.3 µg/mL anti-DNP-IgE antibody before  
214 incubation for 24 h at 37°C, then washed twice with PBS (-) to eliminate the free IgE, and then  
215 the cells were exposed to various concentration of Cry j 1 extract (0, 1/100, 1/50, 1/25, 1/10, 1.0  
216 dilution in medium) for 1 h. As positive and negative controls, 0.3 µg/mL DNP-BSA and PBS  
217 (-) were used respectively. The β-hex in the supernatant was measured as described in the  
218 previous study (Yamada et al., 2010).

219

## 220 2.9. Cytokine assay

221

222 To test the amount of TNF-α and IL-13, RBL-2H3 cells were seeded onto 24-well plates at  
223  $2.5 \times 10^5$  cells/well in 500 µL of medium. Cells were incubated for 16 h in the presence of a  
224 Naha 09 and Tsu.09 water extract at a final dilution of 1/100 in growth medium. For positive  
225 and negative controls, 5.0 ng/mL LPS and PBS (-) were used, respectively. Levels of TNF-α  
226 and IL-13 were measured in RBL-2H3 cells' supernatant using ELISA kits (Inivitrogen Co.,  
227 UAS, CA) according to the manufacturer's instructions. The study was performed in triplicate  
228 using cells from three different passages.

229

## 230 2.10. MTT assay

231

232 The MTT reduction assay is the most frequently used method for quantitative cell viability

233 (Yamada et al., 2007). RBL-2H3 cells were seeded on to 96-well plates at  $5.0 \times 10^4$  cells/well  
234 in 100  $\mu$ L of medium. After an overnight incubation, the growth medium was removed  
235 followed by addition 100  $\mu$ L of the water extract (dilution 0, 1/1000, 1/100, 1/10, 1/2, 1.0 with  
236 medium). For the highest concentrations, additional controls (1, 5 and 10% PBS (-), data not  
237 shown) were considered as references in order to consider the sample effect exclusively on the  
238 survival and proliferation of RBL-2H3 cells. The cells were incubated for 24 h, and cell  
239 viability was measured as described in the previous study (Yamada et al., 2010).

240

#### 241 *2.11. Statistical analysis*

242

243 Results are expressed as the mean  $\pm$  standard deviation of at least three independent  
244 experiments. Experiments were realized in triplicates. Comparisons with the control were  
245 performed by analysis of variance (ANOVA), followed by the Newman-Keuls post hoc test. *p*  
246 values of less than 0.05-0.01 were considered significant.

247

### 248 **3. Results**

249

#### 250 *3.1. Chemical compositions of YS*

251

252 To confirm chemical composition of collected sample during ADSE, the results of  
253 component analysis of YS by XPS and XRF-EDS collected in 2009 were shown in Table 2.  
254 The upper most surface mainly consists of carbon (C) and oxygen (O). In all 3 samples, there

255 are much calcium (Ca) and sulfur (S) contents. The dust particles frequently contained Al and  
256 Fe besides silicon (Si) as indicators of dust particles (Nishikawa et al., 2000), and the percentage  
257 were higher in Fuku.09 and Tsu.09 compared with Naha09. Our observation was correlated  
258 with ASDE information of Japan Meteorological Agency (MEGJ, 2010a). Other minerals  
259 components such as K, and magnesium (Mg) were also detected. A few amount nickel (Ni)  
260 and zinc (Zn) were detected in Naha and Fuku.09. The percentage of Ti was higher in Tsu.09  
261 compared with Naha09 and Fuku.09.

262

### 263 *3.2. Concentration of LPS in water extract of YS*

264

265 To examine the endotoxin concentration in the collected sample during ADSE, we  
266 measured the endotoxin concentrations of each water extract (Table 3). The results showed  
267 that all water extracts contained LPS, Tsu.10 ( $9.49 \pm 1.82$  EU/mL), Fuku.10 ( $16.25 \pm 2.34$   
268 EU/mL), and Naha10 ( $22.62 \pm 1.96$  EU/mL) contained the highest level of LPS, followed by  
269 Tsu.09 ( $8.39 \pm 1.81$  EU/mL), Fuku.09 ( $7.20 \pm 0.36$  EU/mL), and Naha09 ( $3.4 \pm 0.26$  EU/mL).  
270 The LPS level of the sample collected in 2010 was higher than the sample collected in 2009  
271 (Table 3).

272

### 273 *3.3. Concentrations of Cry j 1 in Cry j 1 extract of YS*

274

275 To examine the Cry j 1 concentration in the collected sample during ADSE, the Cry j 1  
276 concentration of each Cry j 1 extract was measured (Table 3). The results showed that all Cry j

277 1 extract contained Cry j 1. Fuku.10 ( $19.7 \pm 2.1 \text{ pg/m}^3$ ) contained the highest level of Cry j 1,  
278 followed by Tsu.10 ( $13.4 \pm 2.3 \text{ pg/m}^3$ ) and Naha10 ( $1.7 \pm 0.3 \text{ pg/m}^3$ ). Tsu.09 also contained  
279 the a high level of Cry j 1 ( $17.2 \pm 2.3 \text{ pg/m}^3$ ) (Table 3).

280

#### 281 *3.4. YS extract induces $\beta$ -hex release from RBL-2H3 cells*

282

283 We performed a series of experiments to screen for the effects of water extract on  
284 RBL-2H3 cells degranulation over a range of noncytotoxic concentrations (dilution rate  
285 1/125-1/50). The  $\beta$ -hex release from RBL-2H3 cells as caused by treatment with the water  
286 extract is shown in Fig. 2. The water extract collected in 2009 and 2010 from each sampling  
287 site induced the  $\beta$ -hex release from RBL-2H3 cells after 16 h of exposure, the effect being  
288 significant at 1/100 treatment (except Fuku.10 sample, Fig. 2B) ( $p < 0.05$  -  $p < 0.01$  vs. the  
289 negative control of PBS (-)), and only Fuku.10 sample showed significant effect at 1/125  
290 treatment (Fig. 2B). For comparison, a LPS (*Escherichia coli* 0111:B4) induced approximately  
291 150% release of  $\beta$ -hex at 5 ng/mL concentration (Fig. 2D).

292 To confirm the effect of Cry j 1 extract on  $\beta$ -hex release, we performed a series of  
293 experiments to screen for the effects of various concentrations (dilution rate 1/100-1) of Cry j 1  
294 extract on RBL-2H3 cells degranulation, and the result is shown in Fig. 3. For comparison,  
295 BSA induced approximately 193%  $\beta$ -hex release at 0.3  $\mu\text{g/mL}$  treatment. The Cry j 1 extract  
296 collected in 2009 and 2010 from each sampling site induced the  $\beta$ -hex release from RBL-2H3  
297 cells, the effect being significant at all treatment ranges with Fuku.10 sample (Fig. 3B), 1/10-1

298 treatment with Naha10 sample (Fig. 3A), and treated with Tsu.09 sample (Fig. 3C) ( $p < 0.05$  -  
299  $p < 0.01$  vs. the negative control of PBS (-)).

300

### 301 *3.5. Water extract induces cytokine production in RBL-2H3 cells*

302

303 The production of TNF- $\alpha$  and IL-13 following 16 h incubation with water extracts by both  
304 Naha09 and Tsu.09 analyzed using ELISA assay are shown in Fig. 4. TNF- $\alpha$  production was  
305 increased RBL-2H3 cells treated with noncytotoxic concentrations of water extract (1/100  
306 dilution rate). The TNF- $\alpha$  production was  $7.76 \pm 1.24$  pg/mL after stimulation with Tsu.09 for  
307 16 h, and Naha09 was  $7.66 \pm 0.62$  pg/mL, the effect being significantly ( $p < 0.05$ - $p < 0.01$ )  
308 different from the control ( $3.06 \pm 0.53$  pg/mL). Furthermore, the TNF- $\alpha$  production was  $9.28$   
309  $\pm 0.15$  pg/mL after stimulation with 5 ng/mL LPS, and the effect being significant ( $p < 0.01$ ),  
310 while there was no significant different in the IL-13 production after stimulation with water  
311 extract (data not shown).

312

### 313 *3.6. Water extract exhibits cytotoxic effects on basophil cells*

314

315 To characterize YS-induced cytotoxicity, cell viability was determined in water extract- and  
316 Cry j 1 extract-stimulated RBL-2H3 cells using the MTT assay. The results showed that  
317 exposure to water extract at different concentration rate for 24 h reduced the cell viability in a  
318 concentration-dependent manner. The dilution rate of 1/10-1 can decrease the cell viability to  
319 Naha09 was  $69.2\% \pm 0.9\%$  (Fig. 5A), Fuku.09 was  $82.0\% \pm 1.7\%$  (Fig. 5B), Tsu.09 was  $79.8\%$



320  $\pm 1.1\%$  (Fig. 5C), Naha10 was  $64.2\% \pm 1.0\%$  (Fig. 5A), Fuku.10 was  $73.6\% \pm 1.2\%$  (Fig. 5B),  
321 and Tsu.10 was  $67.6\% \pm 1.4\%$  (Fig. 5C), the effect being significant compared with the control  
322 ( $p < 0.05 - p < 0.01$  vs. the negative control of PBS (-)). Among the samples, those collected in  
323 2010 showed the strongest toxic effect on RBL-2H3 cells compared with the 2009 samples (Fig.  
324 5). However, Cry j 1 extract did not cause significant reduction in cell viability (data not  
325 shown).

326

## 327 **4. Discussion**

328

### 329 *4.1. Allergenic*

330

331 The presence of water-soluble material around dust-storm particles over Japan has been  
332 reported by previous research reports (Iwasaka et al., 2004; Iwasaka et al., 2008). Most of the  
333 dust particles are coated with water-soluble materials after their long-range transport from China  
334 to Japan, and that the dust storm particles were modified by sea-salt and/or anthropogenic  
335 pollutants and cloud process (Zhou et al., 1996). Our results showed that, YS extracts caused  
336 degranulation of RBL-2H3 cells. Biological species, such as pollen allergens and endotoxin,  
337 as well as transition metals in acidic environments are water soluble (Monn and Becker 1999).  
338 In concordance with several other studies (Becker et al., 1996; Dong et al., 1996; Long et al.  
339 2001), our data suggest the possible involvement of endotoxin in promoting allergy after long  
340 exposure to low level of LPS (Fig. 2D). The endotoxin levels of water extract of samples  
341 collected from 2010 was higher than those of the 2009 samples from all sampling points (Table

342 3), and the differences in LPS content were weakly associated with the increase in filter density  
343 ( $R^2 = 0.5888$ , Fig. 6A), and only the Fukuoka and the Tsukuba samples were strongly correlated  
344 ( $R^2 = 0.9592$ , Fig. 6B). However, LPS content were weakly associated with the  $\beta$ -hex release  
345 rate ( $R^2 = 0.461$ , Fig. 6C). As shown in Fig. 2D, the effect of low level LPS on  $\beta$ -hex release  
346 was not dose-dependent, there is an optimal concentration. In addition, long-range transported  
347 pollutants such as nitrogen oxides (NO<sub>x</sub>) and sulfur oxides (SO<sub>x</sub>) were likely to interact with  
348 aeroallergens (Ichinose et al. 2005) that induce degranulation. Furthermore, metal elements  
349 have inhibitory effect (Tanaka et al., 1991) or induce and enhance (Walczak-Drzewiecka et al.,  
350 2003) action on chemical mediator release by mast cells. Therefore, YS water extract may  
351 have several different ways of inducing chemical mediator release from RBL-2H3 cells,  
352 suggesting the complexity of its action, although most of the mechanisms remain to be studied.

353 It is well known that JC pollinosis patients who start their medication several weeks before  
354 the first day of the pollen season can spend the pollen season without severe symptoms.  
355 Takahashi et al. (2007) found that the total pollen counts and the total Cry j 1 amounts during the  
356 pollen season (Jan. – May. in 2003 – 2006) collected from Yamagata prefecture of Japan did not  
357 correlate with pollen precisely, and the Cry j 1 levels reached 1 pg/m<sup>3</sup> 2-3 weeks before the first  
358 day of the pollen season (5-25 ng/m<sup>3</sup>). We hypothesize that the higher concentration of Cry j 1  
359 collected before pollen season was probably transported from other sites to the sampling sites by  
360 ADSE. This study has shown that there was no correlation between the Cry j 1 contents (Table  
361 3) and the increase in filter particle density (Table 1) ( $R^2=0.0236$ , Fig. 6D). Comparing  
362 samples from Naha and Fukuoka, the Cry j 1 concentration were increased when the filter  
363 particle density increased, but this result do not correlate with the pollen count information

364 (MEGJ, 2010b). On the other hand, the results of this study have shown that Cry j 1 extract  
365 exhibited an effect on the  $\beta$ -hex release in both 2009 and 2010 (Fig. 3), and the differences in  
366  $\beta$ -hex release rate were weakly associated with the increase in the Cry j 1 content ( $R^2 = 0.5168$ ,  
367 Fig. 6E), but higher correlation for the Naha and Fukuoka samples were observed ( $R^2 = 0.9455$ ,  
368 Fig. 6F). This result was consistent with Wang et al. (2009a) observation. This suggests that  
369 ADSE can transport, absorb or concentrate Cry j 1 allergen, and these have the possibility of  
370 contributing to the cause of the allergic reaction (Mori et al., 2003; Takahashi et al., 2007).

371 Sandstorm dust is a prolific source of potential triggers of allergic and non-allergic  
372 respiratory ailments (Kwaasi et al., 1998). Many of the cellular signals activated by  
373 Gram-negative bacteria are attributed to TLR4-mediated recognition of LPS (Gon et al., 2005).  
374 LPS is an agonist of TLR4, although TLR4 receptors are expressed on the RBL-2H3 cell  
375 surface (Passante et al., 2009). In this study, the role of endotoxin as a stimulant of cytokine  
376 production was confirmed (Becker et al., 1996; Dong et al., 1996; Long et al., 2001). The  
377 results have shown the possibility that low level LPS can induce TNF- $\alpha$  production in  
378 RBL-2H3 cells. Cytokine release showed a similar pattern only for TNF- $\alpha$  after stimulation  
379 with Tsu.09 and Naha09 samples (Fig. 4), while there was no effect on IL-13 expression.  
380 Mitogen activated protein kinase (MAPK) JNK was involved in LPS-induced IL-13, but not in  
381 TNF- $\alpha$  synthesis (Gon et al., 2005). This difference may be contributed by the  
382 proinflammatory activity of water extract. However, the endotoxin levels of water extract  
383 from Tsu.09 was 2-fold higher than Naha09 (Table 3). These results indicate that the  
384 cytokine-inducing capacity of Naha09 and Tsu.09 are not only due to the low amount of LPS  
385 present in these water extract. It was hypothesized that there are other proinflammatory

386 components, ambient particles (Imrich et al., 2000) or other mechanism (Shoenfelt et al., 2009),  
387 and there may be greater synergism between endotoxin and components of ambient particles  
388 (Long et al., 2001). Dong et al. (1996) demonstrated that there was no cytokine induction by  
389 diesel particles. This suggests that there are other proinflammatory components in the water  
390 extract that may be responsible for the effects observed in cells treated with YS.

391

#### 392 4.2. Cytotoxicity

393

394 Our results suggest that YS water extract exhibited cytotoxicity, with the cell treated with  
395 2010 samples exhibiting higher cell viability than the 2009 samples (Fig. 5A, B, C). The  
396 differences in the cell viability were weakly associated with the increase in filter particle density  
397 ( $R^2 = 0.6711$ , Fig. 5D), but higher correlation for the Naha and Fukuoka samples was observed  
398 ( $R^2 = 0.9253$ , Fig. 5E). These results agree with the results of the study of Meng and Zhang  
399 (2007) which showed that the effect of PM on health is greater during dust storms because  
400 airborne PM's mass is high. Ichinose et al. (2005) found that inflammatory lung injury was  
401 induced by microbiological materials, such as  $\beta$ -glucan, and by chemical materials such as  $\text{SO}_4^{2-}$ .  
402 Therefore, these materials adsorbed onto dust particles, are implicated in the pathogenesis of  
403 human respiratory disorders during a dust event. YS exhibits a cytotoxic effect on  
404 pneumocytes *in vitro*, and reactive oxygen species (ROS), fenton activity, reactive nitrogen  
405 species (RNS), and titania ( $\text{TiO}_2$ ) are involved in this toxicity (Kim et al., 2003). Becker et al.  
406 (1996) also found that  $\text{TiO}_2$  is toxic to both human and rat alveolar macrophage (AM). Monn  
407 and Becker (1999) reported that LPS is cytotoxic on human monocytes and rat AM. Toxicity

408 in macrophages has also been associated with metals, and higher metal concentrations are  
409 usually found in the PM (Monn and Becker 1999; Schins et al., 2004). Some studies also  
410 suggest that metals, either water-soluble or PM can catalyze reactions involved in oxidative  
411 stress and DNA damage (Meng and Zhang 2007). Nickel (Ni) has slight cytotoxic effect on  
412 CHO cells (Fredj et al., 2010), and significant positive dose response for LDH, a marker of cell  
413 death (Cho et al., 2010). In this study, composition analysis results showed that YS surface  
414 contained Al, C, Fe, nitrogen (N), Ni, S, Ti, and Zn (Table 2). Previous studies showed that  
415 transition metals such as Fe and Zn, components that might be bound to the fine particles,  
416 endotoxins, other organic compounds, and polycyclic aromatic hydrocarbon content of particles  
417 participated in the production of ROS and was correlated with oxidative stress in macrophages  
418 (Ortiz et al., 2006). Effect of diesel particulate matters on ROS production in human alveolar  
419 epithelial cells was observed (Patel et al., 2011). YS contain ultrafine particles from  
420 combustion sources, such as diesel and gasoline engines, and coal fired power plants, and  
421 concentrated during the long distance transport (Zhou et al., 1996; Mori et al., 2003; Ichinose et  
422 al., 2005). This was evident in the cytotoxic effect of the water extract which could have been  
423 induced by their constituents and enhanced by ADSE.

424

## 425 **5. Conclusion**

426

427 In conclusion, YS can rapidly enhance preformed chemical mediator release in basophilic  
428 cells. This process may depend on endotoxin, Cry j 1, and other airborne allergen content of  
429 YS. These findings may help explain the increase in the incidence of allergic diseases during

430 ADSE in East Asia. The detailed mechanism behind the enhancement of allergic affect of YS  
431 will be the subject of our future study. It is recommended that patients suffering from  
432 pollinosis, and physicians be made aware of the pollen dispersal that may be cause by YS during  
433 the ADSE.

434

#### 435 **Acknowledgements**

436

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439

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601

602 **Figure captions**

603

604 **Fig. 1.** A meteorological map showing areas in Japan, Korea and China impacted by YS. The  
605 sampling sites, Tsukuba, Fukuoka and Naha are indicated.

606

607 **Fig. 2.** Effect of water extract on  $\beta$ -hex release from RBL-2H3 cells. The cells ( $5.0 \times 10^4$   
608 cells/well in 100  $\mu$ g/mL) were preincubated with each extract at 37°C for 16 h. Results  
609 represent one trial (n = 6). Tree additional trials show similar results. \*Significantly different  
610 from the negative control (PBS (-)) (\*:  $p < 0.05$ , \*\*:  $p < 0.01$ ).

611

612 **Fig. 3.** Effect of Cry j 1 extract on  $\beta$ -hex release from RBL-2H3 cells. The cells ( $5.0 \times 10^4$   
613 cell/well in 100  $\mu$ g/mL) were incubated with each extract at 37°C for 1 h after IgE sensitization.  
614 Results represent one trial (n = 6). Tree additional trials show similar results. \*Significantly  
615 different from the negative control (PBS (-)) (\*:  $p < 0.05$ , \*\*:  $p < 0.01$ ).

616

617 **Fig. 4.** Effect of water extract on TNF- $\alpha$  production in RBL-2H3 cells. After exposed with  
618 extract (1/100 dilution) and LPS (5 ng/mL) for 16 h, conditioned media were collected for  
619 TNF- $\alpha$  ELISA Kit. Results represent one trial (n = 3). Tree additional trials show similar results.  
620 Tsu.09: collected from Tsukuba city during ADSE in 2009; Naha09: collected from Naha city  
621 during ADSE in 2009. \*Significantly different from the negative control (PBS (-)) (\*:  $p < 0.05$ , \*\*:  $p < 0.01$ ).

622

623  
624 **Fig. 5.** Effect of water extract at different concentration on the cell viability of RBL-2H3 cells.  
625 After 24 h incubation, cell viability was determined using the MTT assay. The percent cell  
626 viability was calculated relative to the untreated control. The cells ( $5.0 \times 10^4$  cells/well) were  
627 incubated with yellow sand extract at 37°C for 24 h in 5%CO<sub>2</sub>. Results represent one trial (n = 6).  
628 Tree additional trials show similar results. \*Significantly different from the negative control  
629 (PBS (-)) (\*:  $p < 0.05$ ). A, B, and C: Cell viability of water extract; D and E: Correlation  
630 between cell viability and filter particle density.

631

632 **Fig. 6.** A and B: Correlation between LPS concentration of water extract and filter particle  
633 density; C: Correlation between  $\beta$ -hex release at 1/100 dilution and LPS concentration of water  
634 extract; D: Correlation between Cry j1 concentration of Cry j1 extract and filter particle density;  
635 E and F: Correlation between  $\beta$ -hex release at 1/100 dilution and Cry j 1 concentration of Cry j 1  
636 extract.

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639 **Fig. 1.**

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656 **Fig. 2.**

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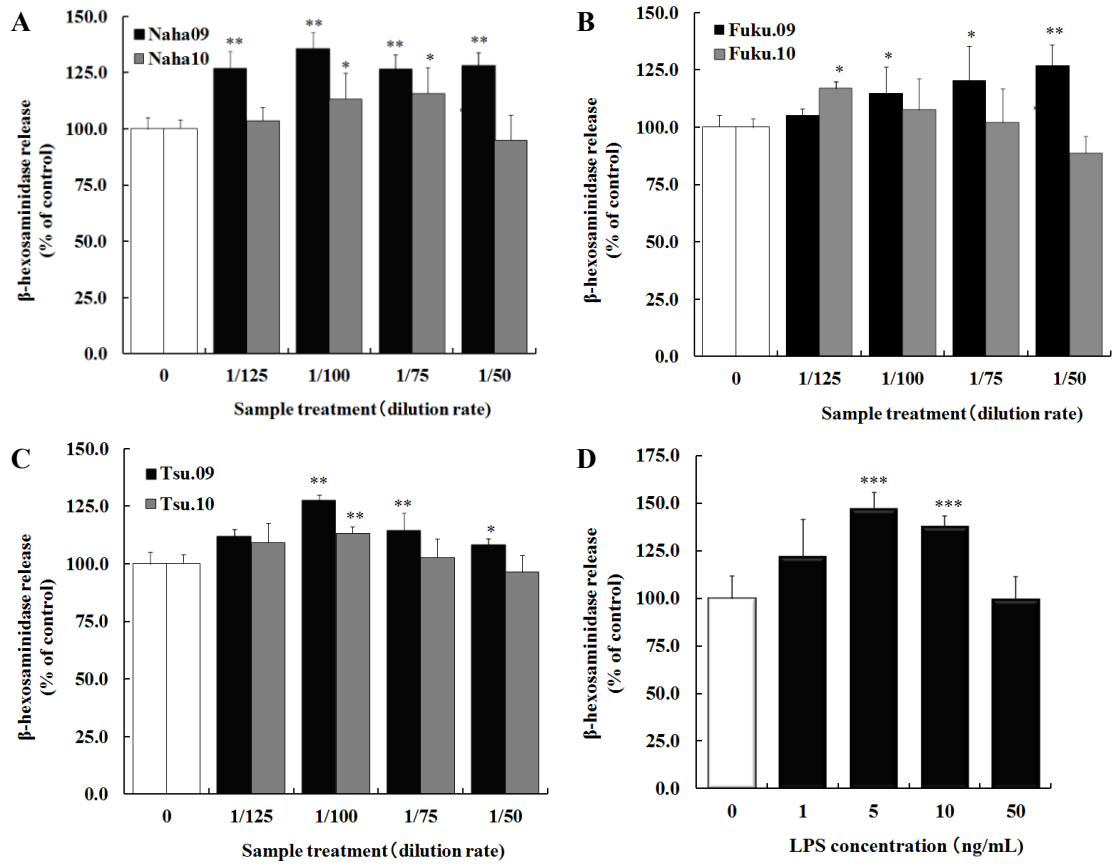
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678 **Fig. 3.**

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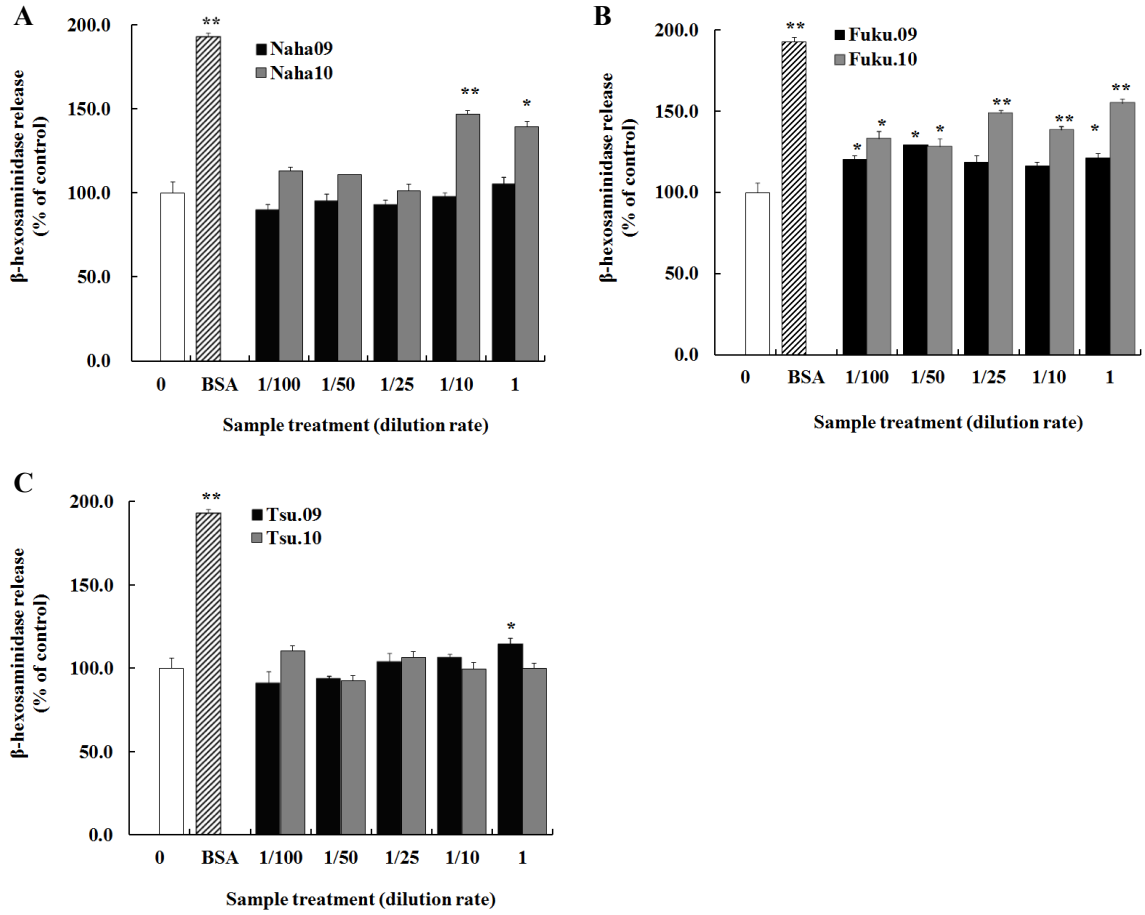
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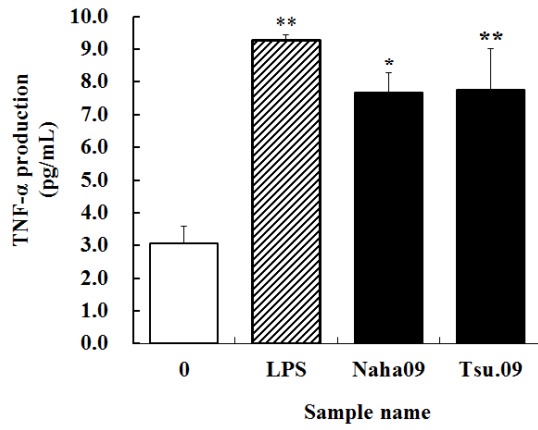


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703 **Fig. 4.**

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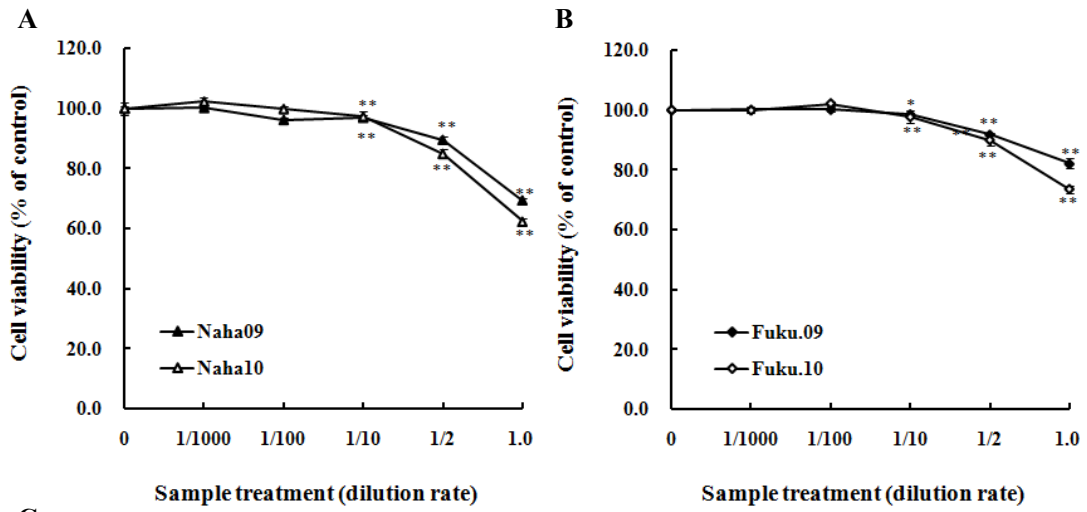
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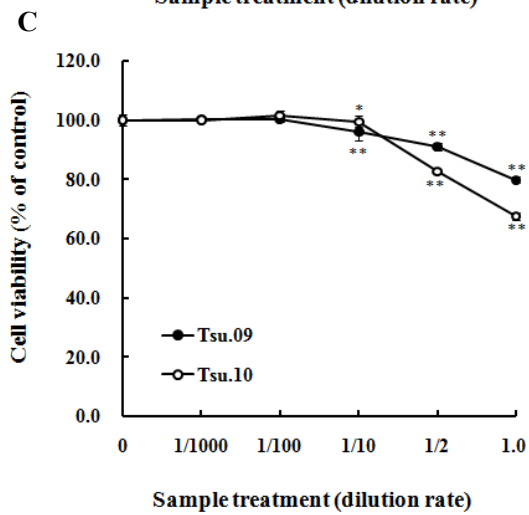
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726 Fig. 5.

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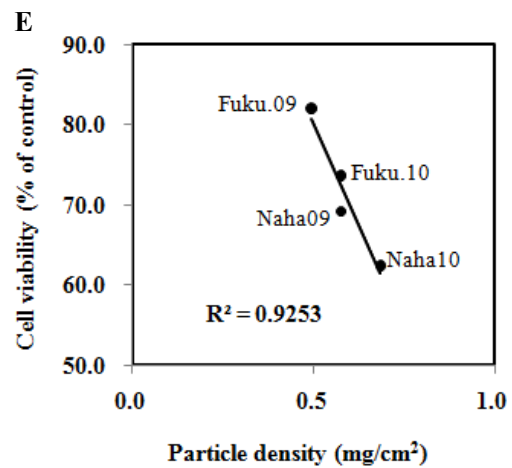
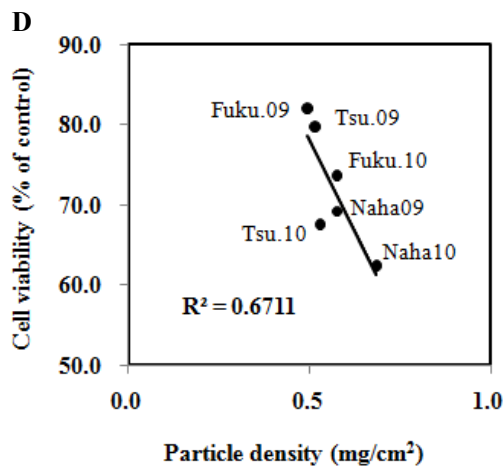
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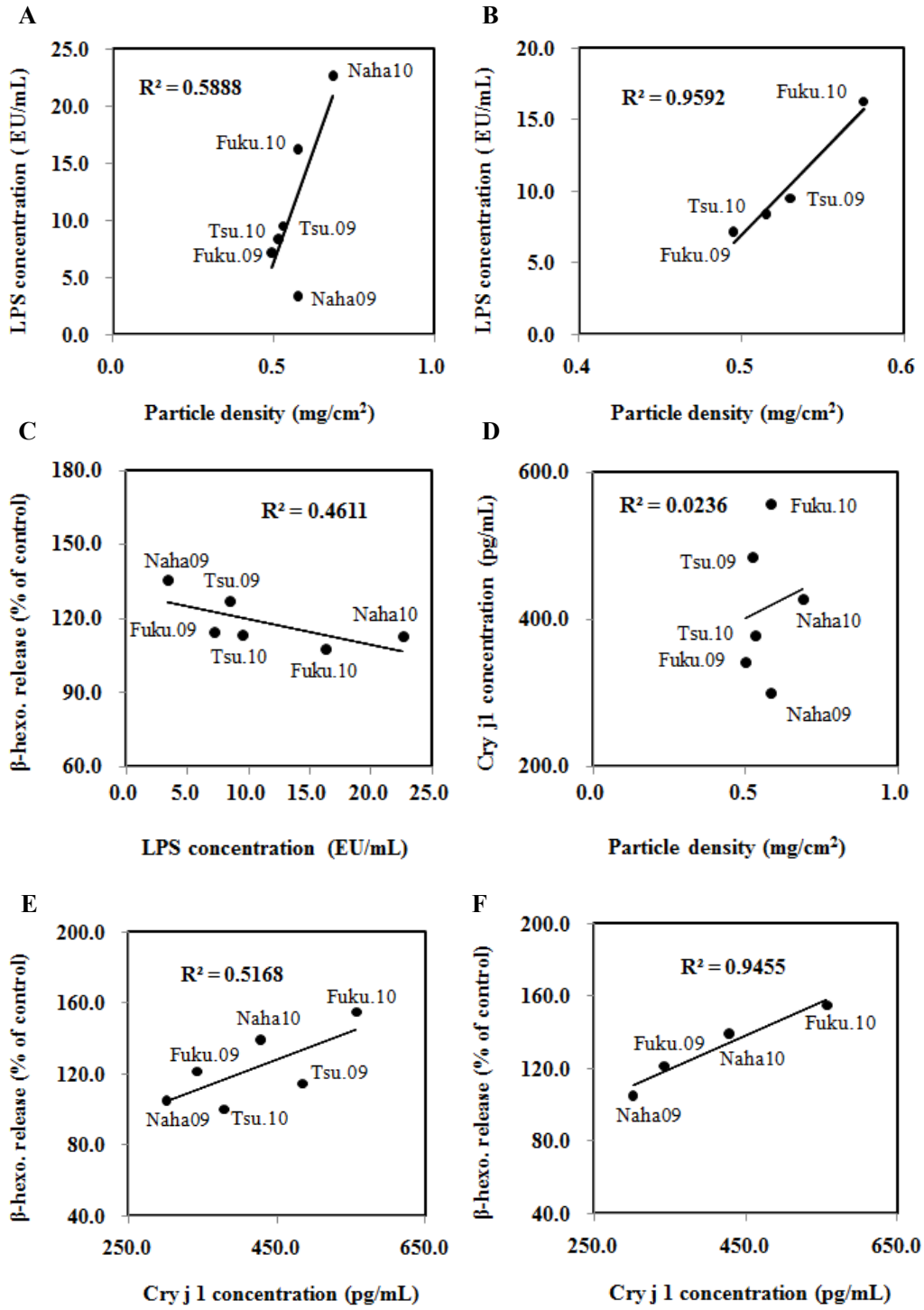
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741 **Fig. 6.**

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756 **Table 1**

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**Table 1**

Sampling site and collection days during the ADSE.

Year	Sample Name	Site	Days	Periods (days)	Particle conc. ( $\mu\text{g}/\text{m}^3$ )	Particle density ( $\text{mg}/\text{cm}^2$ )
2009	Naha09	Naha	4/27 - 6/10	45	$4.6 \pm 0.1$	$0.58 \pm 0.01$
	Fuku.09	Fukuoka	3/16 - 3/19	3	$59.7 \pm 3.6$	$0.50 \pm 0.03$
	Tsu.09	Tsukuba	3/16 - 3/20	4	$46.6 \pm 1.8$	$0.52 \pm 0.02$
2010	Naha10	Naha	3/10 - 4/14	35	$7.1 \pm 0.4$	$0.69 \pm 0.04$
	Fuku.10	Fukuoka	3/20 - 3/24	4	$52.0 \pm 1.8$	$0.58 \pm 0.02$
	Tsu.10	Tsukuba	3/19 - 3/23	4	$47.5 \pm 0.9$	$0.53 \pm 0.01$

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779 **Table 2**

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**Table 2**

Components analysis of YS by XPS and XRF-EDS collected in 2009.

	XRF-EDS analysis			XPS analysis				
	Naha	Fuku.	Tsu.			Naha	Fuku.	Tsu.
Na	13.91	7.58	7.75	C	1s1/2	40.57	38.31	45.75
Mg	2.83	5.66	5.45	N	1s1/2	3.65	3.29	2.34
Al	9.45	13.42	17.87	O	1s1/2	38.84	34.81	33.58
P	1.12	1.96	2.05	Na	1s1/2	4.32	1.42	1.38
S	35.78	24.90	30.80	Mg	2s1/2	0.76	3.68	3.06
Cl	11.60	13.89	— *	Al	2s1/2	2.62	2.20	2.92
K	6.76	7.89	9.23	Si	2s1/2	4.36	7.77	6.55
Ca	13.90	19.24	18.73	S	2s1/2	2.76	2.41	1.58
Ti	0.65	0.81	1.14	Cl	2s1/2	0.39	2.85	0.57
Cr	0.06	0.08	0.09	K	2p3/2	0.08	0.77	0.62
Mn	0.09	0.14	0.18	Ca	2p3/2	1.05	1.84	0.88
Fe	3.59	4.37	6.58	Fe	3s1/2	0.59	0.65	0.77
Ni	0.02	0.01	—					
Zn	0.25	0.07	0.13					
Total	100.00	100.00	100.00	Total		100.00	100.00	100.00
			(atm%)				(atm%)	(atm%)

\* — : No detectable.

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794 **Table 3**

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**Table 3**

LPS and Cry j 1 content of YS extract.

Sample Name	LPS (EU/mL)	Cry j 1 (pg/m <sup>3</sup> )	Cry j 1 (pg/mL)
Naha09	3.40 ± 0.26	1.0 ± 0.2	301.1 ± 68.1
Fuku.09	7.20 ± 0.36	16.2 ± 2.8	342.6 ± 58.9
Tsu.09	8.39 ± 1.81	17.2 ± 2.3	484.4 ± 65.3
Naha10	22.62 ± 1.96	1.7 ± 0.3	428.7 ± 74.5
Fuku.10	16.25 ± 2.34	19.7 ± 2.1	556.8 ± 60.0
Tsu.10	9.49 ± 1.82	13.4 ± 2.3	378.8 ± 64.9

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