Fatty acid synthase inhibitor cerulenin suppresses liver metastasis of colon cancer in mice.

<table>
<thead>
<tr>
<th>基者</th>
<th>墨田守, 山西和宏, 福永幸博, 小林達也, 小澤良子, 東野信宏</th>
</tr>
</thead>
<tbody>
<tr>
<td>論文誌名</td>
<td>Cancer science</td>
</tr>
<tr>
<td>卷</td>
<td>101</td>
</tr>
<tr>
<td>号</td>
<td>8</td>
</tr>
<tr>
<td>頁</td>
<td>1861-1865</td>
</tr>
<tr>
<td>年</td>
<td>2010-08</td>
</tr>
<tr>
<td>版權</td>
<td>© 2010 Japanese Cancer Association</td>
</tr>
<tr>
<td>URL</td>
<td><a href="http://hdl.handle.net/2241/113708">http://hdl.handle.net/2241/113708</a></td>
</tr>
<tr>
<td>doi</td>
<td>10.1111/j.1349-7006.2010.01596.x</td>
</tr>
</tbody>
</table>
Fatty acid synthase inhibitor cerulenin suppresses liver metastasis of colon cancer in mice

Soichiro Murata, MD, PhD, Kazuhiko Yanagisawa, MD, PhD, Kiyoshi Fukunaga, MD, PhD, Tatsuya Oda, MD, PhD, Akihiko Kobayashi, MD, PhD, Ryoko Sasaki, MD, PhD, and Nobuhiro Ohkohchi, MD, PhD

Department of Surgery, Graduate School of Comprehensive Human Sciences, University of Tsukuba

Corresponding author
Nobuhiro Ohkohchi, Department of Surgery, Graduate School of Comprehensive Human Sciences, University of Tsukuba, 1-1-1 Tennodai, Tsukuba, Ibaraki, 305-8575 Japan
Tel.: +81-29853-3221
Fax.: +81-29853-3222
E-mail: nokochi3@md.tsukuba.ac.jp

Word count 2711
Number of figures 5
ABSTRACT

Fatty acid synthase (FAS) is highly expressed in many kinds of human cancers, including colorectal cancer (CRC), and we have investigated the potential use of FAS inhibitors for chemoprevention of liver metastasis of CRC in mice. Expression of FAS was evaluated in murine CRC cell line Colon 26 and CMT 93. Cerulenin, a natural inhibitor of FAS induced apoptosis in these cell lines. The ability of cerulenin to prevent development of liver metastatic lesion of Colon 26 was evaluated. The numbers and sizes of liver metastatic CRC tumors are significantly reduced by treating mice with cerulenin. Cerulenin treatment is associated with reduced levels of phosphorylated Akt in Colon 26 cells, suggesting that inhibition of this signal transduction pathway might be involved in the chemo preventive activity of this compound. Based on studies in mouse models, inhibiting FAS would be an effective strategy in preventing and retarding growth of liver metastatic tumors of CRC that have high expression of this enzyme.
Introduction

Colorectal cancer (CRC) is one of the most common cancers in the world with more than million new cases \(^{(1)}\). The liver is the commonest site of distant metastasis in CRC, and approximately 50% of patients ultimately develop liver metastasis in the course of CRC. \(^{(1,2)}\) Despite recent advances, systemic chemotherapy for metastatic disease is considered palliative, and we rarely see long-term survivors treated only by chemotherapy. \(^{(1)}\) Hepatic resection, the only curative treatment for liver metastasis of CRC, has become the standard treatment, but most cases of liver metastases are inoperable and approximately 50% of the patients treated with hepatectomy have a tumor recurrence in the liver. \(^{(3)}\)

Fatty acid synthase (FAS) is highly expressed in many human cancers including CRC, \(^{(4-7)}\) and previous studies have shown that cancer cell growth can be suppressed by inhibiting the activity of this enzyme with a natural antibiotic cerulenin, \(^{(8)}\) orlistat which is a pancreatic lipase inhibitor developed for obesity treatment, \(^{(9)}\) and C75, which is a stable synthetic small-molecule developed specifically for inhibiting FAS. \(^{(10)}\) But there are no previous studies in which FAS inhibitors suppress liver metastasis of CRC.

The aim of this study extends the investigation of the potential use of a FAS inhibitor for chemoprevention of liver metastasis of CRC in mice. In this study, we examined the effect of cerulenin on the murine CRC cell line, Colon 26 and CMT 93 cell proliferation and apoptosis. Then, the effect of cerulenin on the prevention of growth of liver metastasis lesion of Colon 26 was investigated.
Materials and Methods

Cerulenin

Cerulenin was obtained from Sigma (St. Louis, MO). For cell culture and i.p. injections, cerulenin was dissolved in acetone at a concentration of 20 mg/ml and stored at -20°C. In vitro experiment, 50 to 200 µM of cerulenin was added to the medium and cell viability assay and Western blot experiments were performed 24 hours later. In vivo experiments, treatment with cerulenin at 30 mg/kg was given i.p. at day 1, 4, and 7 after tumor inoculation in cerulenin group.

Cell culture

Two murine CRC cell lines, Colon 26 and CMT 93, were used and tested for mycoplasma-free cell lines. All cancer cell lines were subdivided in multiple tubes for stock in liquid nitrogen immediately after possession. All cell lines were subjected to present experiment within 6 months of resuscitation. Stock cultures were grown in high glucose DMEM containing 10% FBS and 1% antibiotics. The cells were grown in growth medium at 37°C in a 95% air, 5% CO₂-humidified incubator.

Cell viability assay

To measure the cytotoxicity of cerulenin against these cancer cells, 3×10³ cells were plated per well onto 96-well plates. Following overnight culture, cerulenin were added at specified concentrations. After 24 hours of incubation, cell viability was measured by the mitochondrial activity in reducing 2-(2-Methoxy-4-nitrophenyl)-3-(4-nitrophenyl)-5-(2,4-disulfophenyl)-2H-tetrazolium monosodium salt (WST-8) to formazan using Cell Counting kit-8 (Dojindo Laboratories, Kumamoto, Japan). Cells were incubated with a reagent as per manufacturer’s instructions. Plates were read at A450 on a spectrometer.
Cell Proliferation Assay

To measure the cell proliferation activity of cerulenin against these cancer cells, 3×10^3 cells were plated per well onto 96-well plates. Following overnight culture, cerulenin were added at specified concentrations. After 24 hours of incubation, cell proliferation was measured by BrdU assay kit (Roche Diagnostics GmbH, Penzberg, Germany). Cells were incubated with a reagent as per manufacturer’s instructions. Plates were read at A_{450} on a spectrometer.

Apoptosis assay

The In situ Cell Death Detection kit (Roche diagnostics Corp., Basel, Switzerland) was used for the demonstration of apoptotic cell death of cell culture and liver tissue. 3×10^4 /ml cells were plated per well onto Lab-Tek II Chamber Slide (Nalge Nunc International K. K., Tokyo, Japan) and paraffin-embedded liver samples were incubated with the terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling (TUNEL) reaction mixture according to manufacture’s recommendations.

Western blot analysis

For Western blot analysis, total protein extracts of Colon 26 and CMT 93 were obtained 24 hours after cerulenin treatment, and separated by 10% SDS-PAGE and transferred to nitrocellulose membrane (Millipore, Bedford, MA). The following antibodies were used as primary antibodies; total Akt (9272), phosphoserine 473 Akt (9271), p mTOR, cleaved caspase 3, and GAPDH (2118) (Cell Signaling, Beverly, MA). Purified mouse anti fatty acid synthase antibody (610962) was purchased from BD Biosciences (San Jose, CA). Secondary goat anti rabbit antibody conjugated with horseradish peroxidase was purchased from Zymed Laboratories Inc. (San Francisco, CA). Immunoblots were analyzed by enhanced chemiluminescence.
Animals

Eight-week-old male BALB/c mice (Clea, Japan), weighing 24–28g were utilized.
The mice were kept in a temperature-controlled room on a 12-hour light-dark cycle.
They had free access to water and standard chow throughout the experiment. After an
acclimation period of at least 7 days, the mice were separated into two groups as
follows: control group, mice without any treatment (n=8); and cerulenin group, mice
with cerulenin treatment (n=6). All animal experiments were carried out in a humane
manner after receiving approval from the Institutional Animal Experiment Committee
of the University of Tsukuba and in accordance with the Regulation for Animal
Experiments in our university and Fundamental Guideline for Proper Conduct of
Animal Experiment and Related Activities in Academic Research Institutions under the
jurisdiction of the Ministry of Education, Culture, Sports, Science and Technology.

Liver metastasis

Under ether anesthesia, laparotomy was performed with mid line incision, and
hepatic ischemia was induced by clamping the portal triad (the hepatic artery, portal
vein, and bile duct) with a microclip (Aesculap, Tuttlingen, Germany) for 1 min. After
1 min reperfusion of the liver, $1 \times 10^5$ cells of colon 26 cells were injected into the lower
splenic pole with a 27-gauge needle. Nine days after inoculation, mice were sacrificed
and livers were removed for examination.

Histochemical examination

To assess the liver metastatic area, left and middle lobes of liver were divided in
5mm slice, fixed with 10% formaldehyde, and embedded in paraffin. Thin sections
(4µm) were stained with hematoxylin-eosin, and the percentage of liver metastatic area
of Colon 26 cells was calculated using image-processing software WinROOF (Mitani
Co., Fukui, Japan).

**Serum parameter**

Blood was collected from the plexus of the retro-orbital vein of mice. Blood was centrifuged for 10 min at 4°C at 3500 rpm. Supernatants were collected and stored at -80°C until tested by a serum multiple biochemical analyzer (Fuji Drichem; Fuji Film Inc., Tokyo, Japan) for measuring triglyceride, cholesterol, and ALT levels.

**Statistical analysis**

All data are expressed as the mean ± standard deviation of samples. Unpaired t-test was used for the comparison between the two groups. Comparisons between various points were using one-way ANOVA. Significant data were examined by Bonferroni-Dunn multiple comparisons post hoc test. In all cases, a P value <0.05 was considered significant.

**Results**

**Dose-dependent inhibition of proliferation of CRC cell lines by cerulenin**

We initially determined whether cerulenin treatment led to inhibit CRC cell proliferation. CRC cells were treated with various doses of cerulenin for 24 h, and cell viability was assayed using WST-8 assay (Figure 1A) and BrdU assay (Figure 1B).

Figure 1 A and B showed that as the dose of cerulenin increased from 100-200 µM, cell growth inhibition increased in a dose-dependent fashion in CRC cell lines. Cerulenin-induced growth inhibition was found to be statistically significant (p<0.05) (one way ANOVA) in 100, 150, and 200 µM of cerulenin compared to 0 µM.

**Induction of apoptosis via activation of caspase-dependent pathway by cerulenin**

In subsequent experiments, we determined the mechanism of the observed
suppressive effect of cerulenin by WST-8 assays. The overexpression of FAS has been observed to cooperate with survival pathways, including the PI3K/Akt pathway. These two cell lines expressed FAS and p-AKT constitutively, and treatment of CRC cells with cerulenin suppressed FAS expression, dephosphorylated constitutive activated Akt, and increased cleaved caspase-3 in both Colon 26 and CMT 93 cells (Fig. 2A). TUNEL staining of CRC cells revealed that cerulenin induced apoptosis both in Colon 26 and CMT 93 cells in 50 to 100 µM (Fig. 2B).

**Inhibition of general lipogenesis by cerulenin**

To investigate the physiological consequences of in vivo inhibition of FAS, we administered cerulenin (30mg/kg body weight every 3 days) to mice by i.p. injection. We observed slight weight loss following treatment with no significant difference in comparison to the control group (Fig. 3). Serum triglyceride was significantly decreased in the cerulenin group compared to the control group. This suggests general lipogenesis in the liver was reduced by cerulenin (Fig. 4).

**Inhibition of tumor growth of liver metastasis of CRC by cerulenin**

We evaluated the potential effectiveness of cerulenin for metastatic liver tumors of the CRC cell line. Figure 5A showed histological cross sections of livers removed from representative control group and cerulenin groups. Growth reduction of metastatic liver tumors was recognized in the cerulenin group. Figure 5B indicates tumor areas of the liver in both groups. Tumor growth was significantly reduced by cerulenin administration. Figure 5C showed TUNEL staining of liver sections of control group and cerulenin group. In cerulenin group, apoptotic tumor cells were observed in the metastatic liver tumor.
Discussion

The present study extends the investigation of the potential use of a FAS inhibitor for chemoprevention of liver metastasis of CRC in mice. In this study, we revealed the effect of cerulenin on the murine CRC cell line, Colon 26 and CMT 93, on growth inhibition by inducing apoptosis. Also, the effect of cerulenin on the prevention of growth of liver metastasis lesions of Colon 26 was revealed. This is the first study on inhibiting liver metastasis of CRC by administrating FAS inhibitor, cerulenin. Many kinds of human cancer cells have high activities of FAS and numerous studies have described the cytotoxic effects of FAS both in vitro and in vivo. (4) The role of increased FAS in cancer cells and the mechanisms of cell killing by inhibitors of FAS are still not fully understood. (11)

The effect of an intermediate metabolite of fatty acid synthesis on cancer cells is likely mediated through cell signaling pathways. (12) For example, inhibiting FAS decreases phosphorylation of Akt in ovarian cancer cells and suppresses HER2 over expression in breast cancer cells. (12, 13) The mechanism by which FAS inhibitor decreases Akt activation is still unclear, but one mechanism that is advocated is that fatty acids synthesized by FAS are incorporated into membrane phospholipids, which are known modulators of Akt activation. (14, 15) FAS has a major role in the synthesis of phospholipids. (16) When FAS expression is decreased by the treatment of cerulenin, less fatty acid will be synthesized and less phospholipid will be available. One of the important phospholipid is PIP3. PIP3 binds to Akt and its activating kinase PDK-1 with high affinity, and the phosphorylation of Akt is dependent on PIP3. (17) In this study, a decrease of phospholipids may result in inhibition of Akt activity in Colon 26 and CMT 93 cell lines after cerulenin administration. Recently, Liu X et al. reported that inhibition
of PI3K/Akt by cerulenin induces apoptosis in breast cancer cell lines via release of
cytochrome and activation of caspase. (18) In our study it was clarified that inhibition of
PI3K/Akt by cerulenin induces apoptosis in CRC cell lines via activation of caspase.
Colon 26 is chemically derived colon cancer from BALB/c mice. (19) It is highly
metastatic and many researchers use this cell line for study of liver metastasis of CRC in
mice. (20) From these reports we used colon 26 for this study. P53 expression of colon 26
is not fully understood, but reacts to IFN alpha to increase p53, and induce apoptosis.
(21) Functional p53 reacts to interferon alpha or beta. (22) That means p53 in colon 26
would be functional. Further investigation of p53 in colon 26 should be required.
Many efforts to treat xenograft cancers with cerulenin or its derivative of C75 were
reported in ovary, (23) prostate, (24) mesothelioma, (25) breast cancer, (26) and CRC. (27, 28)
But there is no study of treatment of liver metastasis of CRC using cerulenin. In this
study we report that cerulenin inhibits liver metastasis of CRC in a mouse model. In the
prior studies, the use of cerulenin or C75 has been hampered by transient but severe
anorexia and weight loss. Therefore, this compound could also limit the use in the
clinical setting. (29, 30) Loftus et al (29) reported that cerulenin treatment 60mg/kg daily for
7 days caused severe weight loss. Pizer ES et al (23) reported that cerulenin treatment
80mg/kg/daily for 7 days caused severe weight loss. Uddin S et al (15) reported that C-75
treatment 10mg/kg or 20mg/kg twice weekly did not cause severe weight loss. In our
experiments we obtained adequate liver metastasis 9 days after tumor injection. We
treated cerulenin 10mg/kg or 30mg/kg twice weekly (every 3 days). In our preliminary
study cerulenin treatment of 10mg/kg had no effect (data not shown). By treating
30mg/kg of cerulenin every 3 days, inhibition of liver metastasis was observed. At this
amount, serum triglyceride was significantly decreased in cerulenin group. That means
FAS was really inhibited in our animal model. But we could not observe significant weight loss in this group compared to control group. This result indicated the amount of cerulenin in our experiment was appropriate for prevention of side effects of FAS inhibitor. We could not promote complete remission of metastatic tumors by cerulenin at this dose and the appropriate dose of cerulenin or other FAS inhibitors needs to be studied. Also, the effect of FAS inhibitors to the chemically induced colon cancer should be evaluated.

One of the reasons for anorexia brought on by cerulenin or C75 is fatty acid oxidation by activating carnitine O palmitoyltransferase 1. In recent years, it was reported that C93, which inhibits FAS but has no significant effect on fatty acid oxidation, is effective for treatment of human lung cancer xenografts. As a result, C93 does not cause anorexia or weight loss. In near future analogues of FAS inhibitors such as C93 would represent promising new treatments for cancer, including liver metastasis of CRC.

In conclusion, an FAS inhibitor of cerulenin inhibits CRC cell line survival in vitro and in vivo models with liver metastasis. FAS would be a new drug target for liver metastasis of CRC.

Acknowledgements

The authors thank Yuko Jinzenji and Ako Takahashi for technical assistance.

Reference

1. Leonard GD, Brenner B, Kemeny NE. Neoadjuvant chemotherapy before liver resection for patients with unresectable liver metastases from colorectal carcinoma.


10. Wang HQ, Altomare DA, Skele KL et al. Positive feedback regulation between AKT...


tumor growth in athymic mice bearing colon 26 adenocarcinoma cells. Journal of
Veterinary Science 2008; 9: 45-50.
22. Takaoka A, Hayakawa S, Yanai H et al. Integration of interferon-α/β signaling to
p53 responses in tumour suppression and antiviral defense. Nature 2003; 424:
516-23.
23. Pizer ES, Wood FD, Heine HS et al. Inhibition of Fatty Acid Synthesis Delays
Disease Progression in a Xenograft Model of Ovarian Cancer. Cancer Research
1996; 56: 1189-93.
24. Pizer ES, Pflug BR, Bova GS et al. Increased fatty acid synthase as a therapeutic
target in androgen-independent prostate cancer progression. Prostate 2001; 47:
102-10.
25. Gabrielson EW, Pinn ML, Testa JR, Kuhajda FP. Increased fatty acid synthase is a
26. Pizer ES, Thupari J, Han WF et al. Malonyl-coenzyme A is a potential mediator of
cytotoxicity induced by fatty-acid synthase inhibition in human breast cancer cells
28. Huang P, Zhu S, Lu S et al. Inhibitor of fatty acid synthase induced apoptosis in
29. Loftus TM, Jaworsky DE, Frehywot GL, et al. Reduced food intake and body
weight in mice treated with fatty acid synthase inhibitors. Science 2000; 288:
2379-81.
Figure Legends

Figure 1
A. Effect of cerulenin on the WST-8 assay of murine CRC cell lines. CRC cell lines were treated with 0 to 200 µM cerulenin for 24h. Left, Colon 26; right, CMT93. * p<0.01 compared to 0 µM cerulenin. The values indicate ratio compared to 0 µM cerulenin as 100%.

B. Effect of cerulenin on the BrdU assay of murine CRC cell lines. CRC cell lines were treated with 0 to 200 µM cerulenin for 24h. Left, Colon 26; right, CMT93. * p<0.01 compared to 0 µM cerulenin. The values indicate ratio compared to 0 µM cerulenin as 100%.

Figure 2
A. Cerulenin treatment causes down regulation of FAS and dephosphorylation of constitutive phosphorylation of Akt in CRC cell lines. Colon 26 and CMT 93 cell lines were treated with 0, 50, and 100 µM cerulenin for 24h. After cell lysis, equal amounts of proteins were separated by SDS-PAGE, transferred to Immobilon membrane, and immunoblotted with antibodies against FAS, p-Akt, cleaved caspase-3, and GAPDH as indicated.

B. Cerulenin treatment causes apoptosis in CRC cell lines. TUNEL staining of Colon 26 and CMT 93 cell lines which were treated with 0, 50, and 100 µM cerulenin for 24h.

Figure 3
Weight of animals for cerulenin and control groups in the experiment. Columns, mean;
bars, SD. White bar, control group; black bar, cerulenin group.

Figure 4
Serum triglyceride 9 days after tumor inoculation. Columns, mean; bars, SD. White bar, control group; black bar, cerulenin group. *p<0.05 vs control group, t-test.

Figure 5
Cerulenin inhibits tumor growth of liver metastasis of CRC cell line in mice.
A. Histologic cross sections of livers of control group (left) and cerulenin group (right). Arrows, metastatic tumors. Original magnification ×100.
B. Tumor area in the liver of all animals in the experiment. Columns, mean; bars, SD. White bar, control group; black bar, cerulenin group. *p<0.05 vs control group, t-test.
Figure 2

A

<table>
<thead>
<tr>
<th>ceruleniin µM</th>
<th>0</th>
<th>50</th>
<th>100</th>
</tr>
</thead>
<tbody>
<tr>
<td>Colon 26</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>FAS</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>p Akt</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>t Akt</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>cleaved Caspase 3</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GAPDH</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CMT 93</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

B

<table>
<thead>
<tr>
<th>ceruleniin µM</th>
<th>0</th>
<th>50</th>
<th>100</th>
</tr>
</thead>
<tbody>
<tr>
<td>Colon 26</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CMT 93</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Figure 3

The figure shows the body weight (g) of animals over the days after implantation. The data is represented by bars, with each bar indicating the mean body weight with error bars showing the standard deviation. The black bars represent the treatment group (cerulenin), and the white bars represent the control group. The x-axis represents the days after implantation, ranging from 0 to 9 days. The y-axis represents the body weight, ranging from 22 to 30 g. The graph indicates a significant difference in body weight between the control and treatment groups, with the treatment group showing a consistent increase in body weight over the days compared to the control group.