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Unc-51/ATG1 Controls Axonal and Dendritic Development via Kinesin-Mediated Vesicle Transport in the Drosophila Brain

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Abstract

**Background:** Members of the evolutionary conserved Ser/Thr kinase Unc-51 family are key regulatory proteins that control neural development in both vertebrates and invertebrates. Previous studies have suggested diverse functions for the Unc-51 protein, including axonal elongation, growth cone guidance, and synaptic vesicle transport.

**Methodology/Principal Findings:** In this work, we have investigated the functional significance of Unc-51-mediated vesicle transport in the development of complex brain structures in Drosophila. We show that Unc-51 preferentially accumulates in newly elongating axons of the mushroom body, a center of olfactory learning in flies. Mutations in unc-51 cause disintegration of the core of the developing mushroom body, with mislocalization of Fasciclin II (Fas II), an IgG-family cell adhesion molecule important for axonal guidance and fasciculation. In unc-51 mutants, Fas II accumulates in the cell bodies, calyx, and the proximal peduncle. Furthermore, we show that mutations in unc-51 cause aberrant overshooting of dendrites in the mushroom body and the antennal lobe. Loss of unc-51 function leads to marked accumulation of Rab5 and Golgi components, whereas the localization of dendrite-specific proteins, such as Down syndrome cell adhesion molecule (DSCAM) and No distributive disjunction (Nod), remains unaltered. Genetic analyses of kinesin light chain (Klc) and unc-51 double heterozygotes suggest the importance of kinesin-mediated membrane transport for axonal and dendritic development. Moreover, our data demonstrate that loss of Klc activity causes similar axonal and dendritic defects in mushroom body neurons, recapitulating the salient feature of the developmental abnormalities caused by unc-51 mutations.

**Conclusions/Significance:** Unc-51 plays pivotal roles in the axonal and dendritic development of the Drosophila brain. Unc-51-mediated membrane vesicle transport is important in targeted localization of guidance molecules and organelles that regulate elongation and compartmentalization of developing neurons.

Introduction

Neurons are highly polarized and compartmentalized cells with an extended axon and highly branched dendrites. Apart from their role in propagating electrical signals, axons serve as a track for long-distance transport of synaptic components that are synthesized in the soma and delivered to the nerve terminals [1]. In mature neurons, membrane trafficking and cargo delivery are essential for their dynamic physiological functions, which depend on active transport of synaptic vesicles [1]. Membrane trafficking is also essential in developing neurons for delivering diverse organelles and molecules that are required for elongation and guidance of the growing axonal and dendritic termini [1,2,3,4,5,6]. Given the elaborate internal compartments, efficient and controlled membrane vesicle trafficking in neuronal cells is thought to be critical in brain development to establish the functional circuitry [7,8,9,10].

Members of the conserved Ser/Thr kinase Unc-51 family (Unc-51/Uncl.1/Uncl.2) are key regulatory proteins that control axonal elongation during nervous system development in C. elegans and mice [11,12,13,14,15,16]. In mouse, Unc51.1 and Unc51.2 are expressed in a number of neuronal populations during development, including cerebellar granule cells and spinal sensory neurons, in which these proteins are localized to vesicular structures in growth cones [13,14,17]. Molecular studies have shown that Unc-51 controls axon formation in the granule cells through the endocytic membrane trafficking pathway via directly binding the synaptic GTPase activation protein (SynGAP) and...
Syntenin, which is a PDZ domain-containing scaffolding protein that binds Rab5 GTPase and is involved in endocytic vesicular turnover [14]. In *C. elegans*, *unc-51* mutants exhibit premature axonal termination, abnormal trajectories and extra-axon branches with abnormal vesicles and the accumulation of cisternae, suggesting underlying defects in membrane vesicle trafficking [12,10]. *Unc-51* interacts with Vab-6, which contains a kinesin motor-like domain, to control axon extension through regulation of the Netrin receptor Unc-40/DCC and the Slit receptor SAX-3/ROBO [19,20,21]. Studies in *C. elegans* have also shown that *Unc-51* interacts with Unc-14, a RUN domain protein, which regulates kinesin 1-dependent vesicle transport by binding to Unc-16/JIP3/JSAP1, a cargo adaptor for the kinesin motor proteins [22,23]. In addition, recent studies in *Drosophila* have shown that *Unc-51* regulates the association of synaptic vesicles and motor proteins via interacting with Unc-76, a kinesin heavy chain (Khc) adaptor protein [16]. *Unc-51* phosphorylates Unc-76, which then interacts with Synaptotagmin 1 (Syt 1), a synaptic vesicle protein. Collectively, these lines of evidence suggest that *Unc-51* regulates the trafficking of early endosomes and their molecular cargos in diverse neuronal cells.

In this work, we investigated the functional significance of *Unc-51* in neural development, focusing on its regulatory functions in kinesin-dependent vesicle transport in the developing *Drosophila* brain. In the larval mushroom bodies (MBs), which are centers of olfactory associative learning [24,25,26,27,28], *Unc-51* is preferentially expressed in newly elongating axons. Mutations of *unc-51* cause disintegration of axonal bundles with aberrant accumulation and mislocalization of Fasciclin II (Fas II), an IgG type cell adhesion/guidance molecule. By genetic interaction experiments, we show that *unc-51* interacts with the kinesin light chain (*klc*) gene in MB development, and that *Klc* mutation causes dendritic and axonal defects that are reminiscent of *unc-51* mutants. These results demonstrate the pivotal role of *Unc-51* in axonal and dendritic development in the brain, and highlight the conserved functions of *Unc-51* in the trafficking of diverse membrane vesicles and cargo molecules that are essential to the growth and guidance of developing neurons.

**Results**

**Organization of the *Drosophila* larval brain**

To analyze the functions of *Unc-51* in neural development in the *Drosophila* brain, we first focused on the developing MBs at the larval stage (Fig. 1A). The cell bodies of the MBs (Kenyon cells) are localized to the dorsal cortex of the brain and extend dendrites into a structure called the calyx (Fig. 1B), which receives olfactory information from the projection neurons (PNs) of the antennal lobe (AL). The axons of the Kenyon cells elongate and fasciculate into a thick bundle to form the peduncle, a parallel axonal tract that extends ventrally and then splits into the dorsal and medial lobes in the larval brain (Fig. 1B). Internally, axons are organized in concentric layers in the developing MBs [28]. The temporal order of the generation of the Kenyon cells is reflected in the organization of the axons in the larval peduncle and lobes, in which the axons of newly born neurons first project into the center (the core) and then shift to the peripheral layers, as they mature [20].

*Unc-51* colocalizes with kinesin motor proteins in newly elongating MB axons

To analyze the functions of *Unc-51* in *Drosophila* brain development, we performed double/triple immunostaining of the larval brain with anti-*Unc-51* antibody [16] and various neuronal markers. Although ubiquitous basal expression was detected in most part of the larval brain, elevated *Unc-51* expression was detected in several regions, including the optic lobe (OL) and the MBs (Fig. 2A). In MBs, *Unc-51* was enriched in the cell bodies (Fig. 2B), the core of the lobes and the peduncle (Fig. 2D, E). Sparse *Unc-51* signals were also detected in the calyx and the outer layers of the peduncle. Double immunostaining with anti-Khc and anti-Fas II [28] antibodies show that, as with *Unc-51*, Khc was preferentially expressed in the core fibers (Fig. 2F). Similarly, double immunostaining with anti-Klc and anti-N-Cadherin antibodies [29] revealed that Klc also was preferentially expressed in the core fibers (Fig. 2G), suggesting active cytoplasmic transport by the kinesin motors in the newly elongating MB axons.

Consistent with its expression in the core fibers, confocal optical sections of the MB cell bodies revealed that *Unc-51* was enriched in the newly differentiated neurons that located at the interface of the ganglion mother cells and the postmitotic neurons that express Dachshund (*Dac*), a marker of differentiation (Fig. 2H).

Loss of *unc-51* causes axonal transport defects in MB neurons

In the segmental nerves of *Drosophila* larvae, *Unc-51* plays a pivotal role in the regulation of axonal transport by mediating...
motor-cargo assembly [16]. Moreover, the observation that Unc-51 colocalizes with the kinesin motor proteins in the MB core fibers suggested that Unc-51 might have a regulatory function in axonal transport in MB neurons. To determine the functional importance of Unc-51 in MB neurons, we analyzed the intracellular transport of synaptic proteins in developing larval MBs. In wild-type MBs, both Synaptobrevin (n-Syb) and Syt 1 were detected in cell bodies, the calyx and the lobes, but not in the proximal peduncle (Fig. 3B, D). In contrast, aberrant accumulation of these synaptic vesicle proteins was detected in the cell bodies and the core fibers of the lobe (arrow in D) but not in the calyx (demarcated with a dashed circle), DL, dorsal lobe; ML, medial lobe. (E) Peduncle section showing the localization of Unc-51 in the core fibers, co-stained with phalloidin. UAS-mCD8::GFP (green) was driven by OK107-GAL4, which is expressed in outer layers. (F, G) Peduncle sections showing the localization of kinesin heavy chain (Khc) and kinesin light chain (Klc) in the core fibers (arrows). Note that Fas II is expressed in the outer layers while N-Cadherin (N-Cad) is expressed in the core fibers. (H) Higher magnification image of MB cell bodies. Arrow indicates elevated Unc-51 expression in newly differentiated cells located at the interface of the ganglion mother cells (GMCs) and the Dachshund (DAC)-positive postmitotic neurons. Nb, neuroblast. Cells were labeled with UAS-mCD8::GFP (green) driven by elav-GAL4. Scale bars: (A–D) 50 μm; (E–H) 10 μm.

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Figure 2. Expression of Unc-51 in the larval brain. (A–E) Late third instar larval brains stained with an anti-Unc-51 antibody (magenta). MB neurons are labeled with UAS-mCD8::GFP (green) driven by OK107-GAL4. (A) Brain hemisphere of a late third instar larva. OL, optic lobe. Arrows indicate the MB core. (B–D) Localization of Unc-51 in the MB cell bodies (B), calyx (C) and the lobe (D). Note that Unc-51 expression is detected in the cell bodies and the core fibers of the lobe (arrow in D) but not in the calyx (demarcated with a dashed circle). DL, dorsal lobe; ML, medial lobe. (E) Peduncle section showing the localization of Unc-51 in the core fibers, co-stained with phalloidin. UAS-mCD8::GFP (green) was driven by OK107-GAL4, which is expressed in outer layers. (F, G) Peduncle sections showing the localization of kinesin heavy chain (Khc) and kinesin light chain (Klc) in the core fibers (arrows). Note that Fas II is expressed in the outer layers while N-Cadherin (N-Cad) is expressed in the core fibers. (H) Higher magnification image of MB cell bodies. Arrow indicates elevated Unc-51 expression in newly differentiated cells located at the interface of the ganglion mother cells (GMCs) and the Dachshund (DAC)-positive postmitotic neurons. Nb, neuroblast. Cells were labeled with UAS-mCD8::GFP (green) driven by elav-GAL4. Scale bars: (A–D) 50 μm; (E–H) 10 μm.

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Figure 3. Loss of unc-51 causes axonal transport defects in MB neurons. (A) Schematic representation of the larval MB. Dashed lines and box indicate the positions of the cross-sections presented in B–E. (B–E) Localization of n-Syb and Syt1 in the larval MB. Transport of n-Syb and Syt1 was monitored with green fluorescent protein (GFP) fusion constructs expressed in MB neurons using the 201Y-GAL4 driver. Note the accumulations of both markers in cell bodies (C1, E1; arrows), the calyx (C2, E2; arrows) and the lobes (C3, E3; arrows) in unc-51(25/25) mutant MBs. Dashed circles demarcate the calyx. Arrowheads in C2 and E2 indicate aberrant vesicle accumulation in the proximal peduncle. (F, G) Number of axonal clogs. Clog accumulations in the lobes were counted. (F) nSyb::GFP. (G) Syt::eGFP. (H–L) Unc-51 kinase activity is required for axonal transport in MB neurons. (H, I) mCD8::GFP distribution in wild type and unc-51(25/25) mutant MBs. Note the aberrant aggregates in the mutant lobes (arrows in I). (J, K) Genetic rescue of the axonal transport defect in unc-51 mutant by an unc-51 transgene. The axonal aggregation phenotype in unc-51(25/25) mutant was rescued by wild-type unc-51 (J) but not by the kinase-deficient (K38A) mutant, unc-51(K38A) (K). UAS-mCD8::GFP and UAS-unc-51 were driven by elav-GAL4. Rescue genotypes: (J) w; elav-GAL4, UAS-mCD8::GFP, UAS-unc-51WT; unc-51(25/25) and (K) w; elav-GAL4, UAS-mCD8::GFP, UAS-unc-51WT; unc-51(25/25). (L) The number of puncta observed in the lobe (mean ± SEM) was plotted for each genotype. **P<0.01 by Student’s t test. n.s., not significant (P=0.125). Scale bars, 10 μm. doi:10.1371/journal.pone.0019632.g003
Mutations in unc-51 cause dendritic targeting defects in MBs

In addition to the fasciculation defects, mutations in unc-51 also caused aberrant projections from the calyx. Approximately 83% of the unc-51 mutant MBs (n = 24) exhibited overextension of calyx fibers. Among them, more than half (54%) of the MBs exhibited overextensions longer than the calyx diameter. On the other hand, the wild-type MBs rarely exhibited calyx overextensions, with only short extensions (3/21; shorter than the calyx diameter) (Fig. 5A–C). Analyses with wild-type and mutant MARCM clones suggested that cell-cell autonomous activity of unc-51 was required for the suppression of dendritic overshooting (Fig. 5D, E); most of the unc-51 mutant clones exhibited either long (48%) or short (39%) overextensions, while wild-type clones exhibiting short extensions were very rare (6%) (Fig. 5F).

The kinase activity of Unc-51 was again critical to suppress dendritic overextension from the calyx (Fig. 5G–J). Expression of the unc-51WT transgene markedly suppressed the calyx overextension defects while expression of the kinase-deficient unc-51K38A transgene failed to rescue the phenotype (Fig. 5K–K). This implies an additional role for Unc-51 kinase activity in confining dendritic extensions of MB neurons within the normal dendritic compartment.

Mutations in unc-51 cause axonal and dendritic targeting defects in AL-PNs

To further investigate the importance of unc-51 in axonal and dendritic development, we analyzed the development of the AL-PNs in the adult brain, which convey olfactory information to higher brain centers by targeting stereotypic locations in the MBs and the lateral horn (LH) (Fig. 1C) [33]. Using a GH146-GAL4 driver, which is expressed in approximately 90 of the estimated 150 PNs [34], we generated PN-MARCM clones that were homozygous for unc-51 mutation.

The wild-type AL-PN axons were bundled in a single fascicle, extending from the AL to the LH (Fig. S2A, C). Although most of the unc-51−/− anterodorsal neuroblast clones (0/11) failed to show clear defects in axonal projections (Fig. S2B), 71% of the unc-51 lateral neuroblast clones (5/7) exhibited fasciculation defects (Fig. S2D). We also generated wild-type and mutant single-cell clones that innervated the DL1 glomerulus (Fig. S2E, F). Analyses with mutant DL1 single-cell clones showed that the number of branches in the calyx was slightly increased in the mutant clones (wild type = 6.4; unc-51 mutant = 7.8, p < 0.05) while the axonal projection patterns were mostly unaffected, with a normal number of terminal branches in the LH (Fig. S2G, H).

Reminiscent of the dendritic phenotype in the mutant MB calyces, many of the unc-51−/− mutant PNs (12/22) exhibited overshooting of their dendrite terminals beyond the AL compartment, while none of the wild-type clones (0/14) exhibited the corresponding phenotype (Fig. S3). Overshooting was observed for all three types of neuroblast clone (anterodorsal, lateral, and ventral neuroblasts) (Fig. S3B, D, F) with some of the mutant PN clones having multiple extensions (Fig. S3D).

To investigate the regulatory functions of unc-51 in dendritic development further, we analyzed the precise targeting phenotypes of the mutant PN clones to the individual glomeruli. We focused on 12 landmark glomeruli that are innervated by GH146-positive anterodorsal PNs and are clearly identifiable by their positions and shapes [35]. Eight of the landmark glomeruli (VA1d, VA1lm, VA3, VM2, DM6, D, DL1, VM7) were normally innervated by the wild-type anterodorsal PNs, whereas four (VA2, DA1, DM5, DM2) were never or rarely innervated (Fig.
Although *unc-51* anterodorsal neuroblast clones exhibited complete innervation of all the on-target glomeruli (Fig. S4B, D, F, M), they often showed mild to severe ectopic innervation of the off-target glomeruli DA1, DM5 and DM2 (Fig. S4B, F, N). These results suggest that *unc-51* is dispensable for correct targeting of on-target glomeruli but is required for restriction of the dendritic arbors within the normal target glomeruli, by suppressing ectopic dendritic innervation.

**unc-51** mutant MB neurons exhibit aberrant accumulation of endosomes and Golgi

To investigate the mechanisms underlying dendritic abnormalities, we analyzed the localization of endosomes and the Golgi. The small GTPase Rab5, a component of early endosomes, plays a pivotal role in dendritic branching of the larval peripheral sensory neurons [10]. Moreover, Unc51.1 regulates Rab5-mediated endocytic pathways in developing granule cells in mice [14]. In the wild-type larval brain, punctate signals of Rab5-containing endosomes were detected in the cell bodies and the calyx of the MBs (Fig. S5A). In contrast, increased numbers of Rab5-containing endosome puncta and irregular accumulations were observed in the cell bodies and the calyx of the mutant MBs (Fig. S5B, G). The Golgi secretory pathway plays a critical role in dendritic growth and branching [5,7,8]. We examined

**Figure 4.** *unc-51* is required for fasciculation of MB axons. (A, B) Peduncle sections of late third instar MBs stained with anti-N-Cadherin (green), Fas II (red) and phalloidin (blue). (A) Wild type. (B) *unc-51*<sup>3/3</sup> mutant. Note the duplicate cores of the mutant. Whereas Fas II is expressed only in mature fibers surrounding the core in the wild type, in the mutant it is aberrantly expressed in a subset of the core fibers (arrow). (C-G) Peduncle sections showing that *Unc-51* kinase activity is required for the fasciculation of core fibers. Core fibers were visualized with anti-N-Cadherin. (C) Wild type. (D) *unc-51*<sup>3/3</sup> mutant. The core defect was rescued by pan-neuronal expression of the *unc-51<sup>WT</sup>* transgene (E) but not by the kinase deficient (K38A) mutant (*unc-51<sup>K38A</sup>*) (F). Genotypes: (E) w; elav-GAL4, UAS-mCD8::GFP, UAS-*unc-51<sup>WT</sup>*; *unc-51*<sup>3/3</sup> and (F) w; elav-GAL4, UAS-mCD8::GFP, UAS-*unc-51<sup>K38A</sup>*; *unc-51*<sup>3/3</sup>. (G) Number of cores in the peduncle sections. Sixty-seven percent of the *unc-51*<sup>3/3</sup> mutant MBs had multiple core layers. The core defect was rescued by the *unc-51<sup>WT</sup>* but not the *unc-51<sup>K38A</sup>* transgene. Sample sizes: n = 16 for wild type, n = 18 for *unc-51*<sup>3/3</sup>, n = 6 for *unc-51<sup>WT</sup>* rescue and n = 8 for *unc-51<sup>K38A</sup>* rescue. (H, I) Cell autonomous activity of *unc-51* is required for the fasciculation of MB axons. Wild type (H) and *unc-51*<sup>3/3</sup> mutant (I) neuroblast clones. Mutant clones have wild type-like dorsal lobe (DL) and medial lobe (ML), but exhibit de-fasciculation of the peduncular axons (arrow). (J, K) Cross-sections of the wild-type (J) and mutant (K) peduncles. Clones were induced by a heat shock at the early first instar stage and labeled with mCD8::GFP (green) driven by elav<sup>C155</sup>. Mature axons were stained with anti-Fas II (magenta). (L) Quantification of the number of peduncular axons in MARCM clones. While only 13% of the wild-type clones (n = 8) exhibited separate bundles, 44% of the *unc-51*<sup>+/−</sup> clones (n = 16) demonstrated split fascicles. Scale bars, 10 μm.

S4A, C, E, K). Although *unc-51* anterodorsal neuroblast clones exhibited complete innervation of all the on-target glomeruli (Fig. S4B, D, F, M), they often showed mild to severe ectopic innervation of the off-target glomeruli DA1, DM5 and DM2 (Fig. S4B, F, N). These results suggest that *unc-51* is dispensable for correct targeting of on-target glomeruli but is required for restriction of the dendritic arbors within the normal target glomeruli, by suppressing ectopic dendritic innervation.
localization of the Golgi machinery using a fluorescently labeled Golgi marker, α-Man-E::eGFP [7]. In the wild-type MB neurons, punctate α-Man-E::eGFP signals were detected in the cell bodies but not in the calyx and the lobes (Fig. S5E). On the other hand, ectopic punctate signals and accumulations of the Golgi marker were detected in the calyx and the lobes of the unc-51 mutant MB neurons (Fig. S5F, I). Aberrant accumulations were also detected in the cell bodies. These results suggest that unc-51 has an essential function in intracellular trafficking and localization of endosomes and Golgi components in developing MB neurons.

In contrast to the significant transport defects of endosomes and Golgi components, accumulation of Mito::GFP, a fluorescently labeled mitochondrial marker [36], was rarely observed in the mutant MBs, although ectopic signals were detected in the proximal peduncle (Fig. S6A, B). We also examined the expression of hLC3::GFP, a fluorescently labeled marker of autophagosomes.

Figure 5. Loss of unc-51 causes dendritic defects in MBs. (A, B) MBs in the wild-type and unc-51 \(25/25\) mutant larvae at the late third instar stage. MBs were visualized with mCD8::GFP driven by 201Y. (C) Quantification of the dendrite overextension phenotype in wild-type and unc-51 mutant larvae. While 14% of the wild-type MBs (\(n = 21\)) exhibited short overextensions, 83% of the unc-51 \(2^-\) MBs (\(n = 24\)) exhibited short or long overextensions (shorter or longer than the diameter of the calyx, respectively). (D, E) Analyses of the dendritic phenotype using MARCM clones. Clones were induced by a heat shock at the early first instar stage. MB neurons were labeled with UAS-mCD8::GFP driven by elav\(^{G42}\). (D) Wild-type neuroblast clone. (E) unc-51\(^{66}\) mutant neuroblast clone. (F) Quantification of the dendrite overextension phenotype in MARCM clones. While only 6% of the wild-type clones (\(n = 17\)) showed overextensions, all of the unc-51\(^{66}\) clones (\(n = 23\)) exhibited short or long overextensions (shorter or longer than the diameter of the calyx, respectively). (G–J) Dendrite overextension phenotype in unc-51 mutant MBs was rescued by pan-neuronal expression of unc-51\(^{WT}\) but not by the kinase-deficient (K38A) unc-51\(^{K38A}\) transgen. (G) Wild-type. (H) unc-51\(^{25/25}\) mutant. Rescue genotypes: (I) w; elav-GAL4, UAS-mCD8::GFP, UAS-unc-51\(^{WT}\); unc-51\(^{25/25}\) and (J) w; elav-GAL4, UAS-mCD8::GFP, UAS-unc-51\(^{K38A}\); unc-51\(^{25/25}\). (K) Quantification of the dendrite overextension phenotype. While none of the wild-type MBs (elav-GAL4, UAS-mCD8::GFP) (\(n = 6\)) exhibited calyx overextension, 83% of the unc-51\(^{25/25}\) mutant MBs (elav-GAL4, UAS-mCD8::GFP; unc-51\(^{25/25}\)) exhibited short or long overextensions (\(n = 6\)). The dendritic overextension phenotype was significantly suppressed by the unc-51\(^{WT}\) transgene (\(w; elav-GAL4, UAS-mCD8::GFP, UAS-unc-51\(^{K38A}\); unc-51\(^{25/25}\)), with 31% of the MBs exhibiting short extensions (\(n = 16\)). Expression of unc-51\(^{K38A}\) failed to rescue the phenotype (\(w; elav-GAL4, UAS-mCD8::GFP, UAS-unc-51\(^{K38A}\); unc-51\(^{25/25}\)), with 73% of the MBs exhibiting short or long overextensions (\(n = 22\)). Note different genetic background from that of the experiments in (A–C). Arrows in (B, E, H and J) indicate overextensions from the calyx. Scale bar, 10 \(\mu\)m.

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[37,38,39], and observed no difference between the wild-type and mutant MBs (Fig. S6C, D).

**Loss of unc-51 causes mislocalization of Fas II in MB neurons**

Fas II plays a critical role in the development of the concentric layers in the MBs [28]. In the wild-type larval MBs, Fas II was localized to the lobes and the distal peduncles but absent from the calyx and the proximal peduncle (arrowheads in Fig. 6A). Although Fas II localization to the distal peduncle was not altered, mutations in unc-51 caused mislocalization of Fas II to the proximal peduncle (arrowheads in Fig. 6B). Moreover, Fas II was detected in the calyx of unc-51/− mutant MBs (Fig. 6B). To further investigate the localization of Fas II in the developing MBs, we examined Fas II distribution using a Fas II::yellow fluorescent protein (YFP) fusion construct driven by the MB-GAL4 driver, 201Y. In the wild-type MBs, Fas II::YFP was localized to the lobes as was the endogenous protein, but was absent from the cell bodies and the calyx (Fig. 6C–E). In contrast, Fas II::YFP aberrantly accumulated in the cell bodies and the calyx in the unc-51 mutant MBs (Fig. 6F, G). In addition, punctate accumulations of Fas II::YFP were detected in the mutant calyx and the lobes (Fig. 6H; quantification in I). Furthermore, clonal studies demonstrated that the unc-51/− mutant clones (13/13) exhibited aberrant localization...
Suppression of kinesin-mediated transport results in dendritic targeting defects in MB neurons

While the above results suggest that unc-51 regulates intracellular trafficking of various vesicles and molecular cargos in developing MB neurons, previous studies have suggested multiple roles for unc-51 in diverse cellular and neuronal processes [12,13,15,17,23,37,38,40,41,42,43]. To examine whether kinesin-mediated transport plays a critical role in developing MB neurons, we studied Klc and unc-51 double heterozygous flies, based on the fact that unc-51 genetically interacts with Klc in kinesin-mediated axonal transport in the segmental nerves [16].

While calyx abnormality was rarely observed in the MBs of either unc-51+/− (7%, n = 15) or Klc+/− (8%, n = 27) single heterozygotes (Fig. 7A, B), many of the MBs of Klc+/− unc-51+/− double heterozygotes exhibited overshooting (56%, n = 53) (Fig. 7C; quantification in E), reminiscent of the MB phenotypes in unc-51+/− homozygotes. On the other hand, axonal fasciculation in the MB core was barely affected in the Klc+/− unc-51+/− double heterozygotes (inset in Fig. 7C; quantification in F). As in the MBs of unc-51+/− homozygotes, Fas II localization was also altered in the MBs of Klc+/− unc-51+/− double heterozygotes.

Accumulation of Fas II was rarely detected in either the Klc+/− or unc-51+/− single heterozygotes (Fig. 8A, B), but the majority of Klc+/− unc-51+/− double heterozygotes exhibited aberrant accumulations and mislocalization of Fas II in the MBs (Fig. 8C), as did the unc-51−/− homozygotes (Fig. 8E; quantification in F-H).

To further investigate functional importance of the kinesin motors in MB development, we examined MBs of klc mutant larvae. Because of the homozygous lethality of the Klc c02312 null allele [44], we used trans-heterozygotes with Klc c02312, a hypomorphic P-element insertion allele [45]. Notably, Klc+/−/ Klc c02312 larvae exhibited weak but significant Fas II accumulations in the MBs (Fig. 8D and F), suggesting that kinesin-mediated intracellular transport plays an essential role in MB development. In addition, as with the unc-51+/−/− larvae, Klc+/−/ Klc c02312 larvae showed mislocalization of Fas II in the calyx and the proximal peduncle (Fig. 8D; quantification in G, H). Moreover, the internal core structure was partially disrupted in the Klc+/−/ Klc c02312 trans-heterozygotes (Fig. 8D3).

To further confirm the functional requirement of kinesin-mediated transport, we generated Klc+/−/ MARCM clones, and found that loss of the klc activity caused disintegration of the axonal bundles and aberrant dendritic overshooting (Fig. S7A-F). In contrast, Klc+/− clones also exhibited mislocalization of Fas II in the proximal part of the peduncle, recapitulating the phenotype of the unc-51+/−/− clones (Fig. S7G). These results thus clearly demonstrate that cell-autonomous activity of the kinesin motor protein is critical for the normal axodendritic development of MB neurons.

Figure 7. unc-51 genetically interacts with Klc in dendrite development. (A–D) Larval MBs of single or double heterozygous mutants. Late third instar stage. MBs were labeled with UAS-mCD8:GFP driven by 201Y-GAL4. Inset panel shows a peduncle cross-section stained with anti-N-Cadherin. Arrowheads indicate overextending dendrites. Arrows in the inset of (D) indicate multiple MB cores in a unc-51+/−/− larva. (E) Quantification of dendritic targeting defects. Sample sizes: unc-51+/−/− (n = 15), Klc+/−/− (n = 27), Klc+/−/− unc-51+/−/− (n = 53) and unc-51+/−/− (n = 21). (F) Quantification of MB core defects. Sample sizes: unc-51+/−/− (n = 13), Klc+/−/− (n = 8), unc-51+/−/− Klc+/−/− (n = 12) and unc-51+/−/− (n = 18). Scale bars, 10 μm. doi:10.1371/journal.pone.0019632.g007
Kurusu et al. [46] have shown that members of the receptor tyrosine phosphatase (RPTP) protein family play a critical role in axonal fasciculation and elongation of the core fibers in developing MBs. To examine possible interaction of \textit{unc-51} and the RPTP genes, we performed genetic interaction experiments between the \textit{unc-51} and the RPTP mutations but failed to detect any abnormalities in the MBs of double heterozygotes for \textit{unc-51} and either \textit{Ptp10D}, \textit{Ptp69D} or \textit{Lar}. Having a wild-type like lobes with a single distinctive core, none of the heterozygotes exhibited abnormality in the fasciculation of MB axons (Fig. S8). Neither aberrant Fas II accumulation nor dendritic abnormality was caused in these double heterozygote larvae.

\textit{unc-51} is dispensable for polarized transport of Dscam and Nod

Having demonstrated the importance of \textit{unc-51} in the kinesin-mediated intracellular transport in the development of MB neurons, we then asked whether \textit{unc-51} was required for the localization of proteins that are distinctively localized to dendrites by retrograde transport motors. Among the multiple isoforms of the Dscam proteins, isoforms carrying exon 17.1 (Dscam[TM1]), which encodes a transmembrane domain, are targeted to dendrites by dynein/dynactin-mediated transport [47,48,49]. In wild-type MBs, Dscam[TM1] was localized specifically to the calyx and absent from the lobes (Fig. S9A–C). In the \textit{unc-51} mutant MBs, Dscam[TM1] partly accumulated in cell bodies but nonetheless was normally targeted to the dendrites, with no detectable signals in the lobes in all the samples examined (\(n=12\)) (Fig. S9D–F; see clog quantification in M, N).

To examine retrograde dendritic transport further, we analyzed the localization of Nod::\(\beta\)-Gal, a chimera comprising \(\beta\)-galactosidase fused to the Nod motor domain, which binds to microtubule minus ends [50]. In wild-type MBs, Dscam[TM1] was localized specifically to the calyx and absent from the lobes (Fig. S9A–C). In the \textit{unc-51} mutant MBs, Dscam[TM1] partly accumulated in cell bodies but nonetheless was normally targeted to the dendrites, with no detectable signals in the lobes in all the samples examined (\(n=12\)) (Fig. S9D–F; see clog quantification in M, N).

To examine retrograde dendritic transport further, we analyzed the localization of Nod::\(\beta\)-Gal, a chimera comprising \(\beta\)-galactosidase fused to the Nod motor domain, which binds to microtubule minus ends [50]. In wild-type MBs, Nod::\(\beta\)-Gal was detected in cell bodies and specifically targeted to the calyx with virtually no signals in the lobes (Fig. S9G–I), confirming previous observations [9,51,52]. Although \textit{unc-51} mutant MBs exhibited accumulations in the cell bodies, Nod::\(\beta\)-Gal was specifically targeted to the calyx in all the samples examined (\(n=14\)) (Fig. S9J–L; see clog quantification in M, N).

Figure 8. Aberrant Fas II accumulation and mislocalization in \textit{unc-51} and \textit{Klc} mutants. (A–E) Fas II::YFP distribution patterns in the cell bodies (A1–E1), calyx (A2–E2) and lobes (A3–E3). Fas II::YFP was expressed in MBs using the 201Y-GAL4 driver. Yellow arrows indicate aberrant Fas II clogs. Arrowheads (C2, D2 and E2) indicate Fas II mislocalization in the proximal peduncle. Scale bars, 10 \(\mu\)m. (F) Number of clogs in the calyx. Error bars, mean ± SEM. **P<0.01, Student’s t test. Comparisons were also made between \textit{Klc}\textit{c02312}/8e\textit{94} and \textit{Klc}\textit{c02312}/8e\textit{94} (**P<0.01) as well as between \textit{unc-51}\textit{25}/25 and \textit{unc-51}\textit{c02312}/8e\textit{94} (***P<0.01). (G) Quantification of Fas II::YFP mislocalization in the calyx. (H) Quantification of Fas II mislocalization in the proximal peduncle. Sample sizes: \textit{unc-51}\textit{25}/25 (\(n=6\), \textit{Klc}\textit{c02312}/8e\textit{94} (\(n=14\), \textit{unc-51}\textit{25}/25 \textit{Klc}\textit{c02312}/8e\textit{94} (\(n=4\), \textit{Klc}\textit{c02312}/8e\textit{94} (\(n=8\) and \textit{unc-51}\textit{25}/25 (\(n=8\).

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quantification in O, P3. Of note, Nod::β-Gal was observed in the
tips of the overextensions of the unc-51 mutant calyx (arrowheads
in Fig. S9K), arguing for their minus end polarity and thus
dendritic identity. These results imply that the activity of unc-51 is
dispensable for polarized dynein/dynactin-mediated retrograde
transport in dendrites. They also argue that the microtubule
system in mutant MB neurons is functionally intact.

Discussion

Active transport and delivery of molecular and cellular
components from the soma to the distinct cytoplasmic compart-
ments is critical not only for synaptic function in mature neurons
but also for axonal elongation and guidance in developing neurons
[1,3,6]. The kinesin-mediated anterograde transport plays a major
role in the active traffic in the developing neurons, delivering a
wide range of cargos along the axon, including synaptic vesicles,
mitochondria, cytoskeletal elements, and mRNAs.

Unc-51 controls membrane vesicle transport in
developing brain via kinesin-mediated transport

In this work, we have demonstrated preferential expression of
Unc-51 and kinesin motor proteins in larval MBs, and shown that
loss of unc-51 activity causes severe defects in kinesin-mediated
transport in developing MB neurons, while dynein/dynactin-
mediated retrograde transport is unaffected in unc-51 mutant MBs.
In addition, loss of unc-51 activity results in disintegration of axonal
bundles and aberrant extensions of dendrites in both MB and AL
neurons. These results suggest that unc-51 is essential for the
development of brain neurocircuities, participating in molecular
and/or cellular mechanisms that regulate the formation of the
complex structures such as MBs. Indeed, our results demonstrate
that unc-51 is essential for the specific intracellular localization of
axonal fasciculation and guidance molecules such as Fas II.

Although previous studies have demonstrated that unc-51 has diverse
functions in developing neurons [12,13,15,17,23,37,38,40,41,42,43],
our analyses of the Khc+/− unc-51+/− double heterozygotes clearly
demonstrate that suppression of kinesin-mediated transport results
in dendritic overextensions. Concomitant suppression of unc-51
and Khc also causes axonal transport defects that are reminiscent
of unc-51 mutants. Furthermore, the double heterozygotes exhibit
mislocalization of Fas II in both the calyx and the proximal
peduncle. This recapitulates the salient phenotype of unc-51+/−
mutants, and argues that defective kinesin-mediated transport is
the major molecular process that underlies the developmental
defects in the unc-51 mutants. On the other hand, Khc+/− unc-51+/−
double heterozygotes failed to exhibit a full range of the unc-51
mutant phenotypes. Although we cannot exclude the possibility
that other molecular processes are involved in the unc-51 mutant
phenotypes, the weaker phenotypes of the double heterozygotes
might be accounted by partial suppression of the gene activity given
that other molecular processes are involved in the
mutant phenotypes. Although we cannot exclude the possibility
that other molecular processes are involved in the
defects in the
Khc+/− unc-51+/− double heterozygotes clearly
indicate that retrograde dynein/dynactin-mediated transport
remains intact in the mutant MB neurons. Moreover, another
dendritic molecule, Nod, is correctly targeted to the calyx, clearly
indicating that unc-51 is not required for polarized retrograde
transport mediated by dynein/dynactin in developing MB neurons.
These results are consistent with our previous observations that
unc-51 fails to interact with retrograde motor genes such as dynemin heavy
chain and Lissencephaly-1 in segmental nerves [16].

Previously, Toda et al. [16] demonstrated that unc-51 plays an
essential role in axonal transport by mediating the assembly of the
cargos and the kinesin motor proteins. In unc-51 mutants,
membrane vesicle transport is severely affected, resulting in
accumulation of synaptic vesicles in the larval segmental nerves.
Loss of unc-51 activity also causes aggregation of Rab5-positive
membranes in the segmental nerves [16]. Notably, as in the mutant
MBs, kinesin motors are accumulated in the mutant segmental
nerves while overall mitochondrial localization was unaffected [16].
Genetic studies have shown that the wild-type but not a kinase-
deficient form of the unc-51 transgene rescues the transport defect in
synaptic vesicles in the mutant segmental nerves [16]. Moreover, the
kinesin activity of Unc-51 is critical for phosphorylation of an
adapter molecule, Unc-76/FEZ, which mediates the assembly
between synaptic vesicle cargos and the kinesin motor complex.
Our finding that the dendritic and axonal defects in MB neurons are
rescued by the wild-type but not by the kinase-deficient unc-51 transgene
suggests that a similar phosphorylation-dependent
regulatory mechanism is involved in the intracellular transport in
developing MB neurons. In line with this, it is noteworthy that Unc-
76 is preferentially expressed in developing MB neurons, in which it
co-localizes with Unc-51 and kinesin motor proteins in the core
fibers (H. M. and K. F. T., unpublished observation). Intriguingly,
both Unc-51 and Unc-76 are downregulated in the mature neurons
that surround the core fibers as they mature and shift to the
peripheral layers. Concomitantly, both Khc and Klc are downreg-
ulated in the mature MB neurons, suggesting dynamic control of the
expression of the molecular components that mediate active vesicle transport in developing MB neurons.

Kinesin-mediated vesicle transport in dendritic
development

Recent genetic studies on the dendritic development of Drosophila
larval sensory neurons showed that the microtubule motor protein
dynein controls dendritic branching by directing polarized intracel-
lar vesicle trafficking [9,10]. Dynemin is also necessary for the
dendrite-specific localization of Golgi outposts [9], which secretory
pathway plays critical roles in dendritic growth and branching
[5,7,8]. These studies also showed that Rab5 was essential to normal
dendritic branching, via its role in controlling endosomal trafficking
[10]. Our results show that mutations of unc-51 leads to aberrant
accumulations of Rab5-containing endosomes and Golgi compo-
nents in developing larval MBs. On the other hand, polarized
dendritic transport of Dscam is not altered in the unc-51 mutant
MBs, implying that retrograde dynein/dynactin-mediated transport
remains intact in the mutant MB neurons. Moreover, another
dendritic molecule, Nod, is correctly targeted to the calyx, clearly
indicating that unc-51 is not required for polarized retrograde
transport mediated by dynemin/dynactin in developing MB neurons.
These results are consistent with our previous observations that
unc-51 fails to interact with retrograde motor genes such as dynemin heavy
chain and Lissencephaly-1 in segmental nerves [16].

On the other hand, Satoh et al. [10] found that khc mutants
showed dendritic branch abnormalities that were almost identical
to those of dynemin light intermediate chain (Dlic) mutants, with reduced
arbor and a marked shift in branching activity in the proximal
area within the arbors. In the khc mutants, the dynein complexes
become aggregated distally, suggesting that kinesin plays a role in
recycling dynein proximally after it has carried its organelle cargo
distally. These phenotypes seem to contrast with those observed in
unc-51 mutant MB neurons, which show dendritic overextensions
with normal dynemin/dynactin-mediated transport. The exact
mechanisms underlying these discrepancies are unknown, but this
might suggest different regulatory processes for kinesin-mediated
transport that operate in the distinct cellular contexts of the peripheral sensory neurons and MB neurons. Toda et al. [16] showed that unc-51 mutation resulted in varying degrees of axonal membrane defects that were dependent on the cargo. It is possible that Unc-51 differentially regulates the transport of specific cargo subsets by phosphorylation of distinct groups of adaptor proteins in different cell types.

**Unc-51 controls axonal growth and guidance by endosomal vesicle trafficking**

Studies in *Drosophila* identified unc-51 as a homolog of the yeast atg1, and suggested that Unc-51 kinase activity is required for the induction of autophagy [37,38,39]. Recently, Shen and Ganetzky [43] showed that autophagy positively regulates synapse development at the *Drosophila* neuromuscular junction. Mutations in autophagy genes including atg1/unc-51, caused significant reduction in terminal branching of the segmental motoneurons, with reduced numbers of boutons. In contrast, our single-cell analysis of AL-PNs (Fig. S2) shows that loss of unc-51 activity results in an increase in the number of the calyx branches. Our results also demonstrate that the distribution of an autophagy marker is not altered between the wild-type and the unc-51 mutant MBs (Fig. S6). These results argue against autophagy as an underlying mechanism of the axodendritic abnormalities in the *unc-51* mutant larval brain, and are consistent with a previous report that autophagy is not involved in Unc-51-mediated regulation of axonal transport [16].

In *C. elegans*, mutations in unc-51 cause diverse axonal defects, including premature termination, abnormal trajectories, and extra axon branches [12]. Developing neurites accumulate abnormal vesicles and cisternae, suggesting underlying defects in membrane vesicle trafficking [18]. Intriguingly, Unc-51 directly interacts with Unc-14, a RUN domain protein, to regulate axonal elongation and guidance, and mutations in unc-14 cause neurite growth and guidance defects that are very similar to those of unc-51 [51]. Unc-14 regulates vesicle transport and localization by binding to Unc-16/JIP3/JSAP1, which is a cargo adaptor for the kinesin motor proteins [22]. Recent studies have shown that RUN domain proteins function as effectors of Rap and Rab GTPases in the control of membrane trafficking [54], highlighting the importance of vesicle trafficking in the regulation of axonal growth and guidance. Several studies have suggested that Unc-51 plays an essential role in the delivery of specific receptors for axonal guidance molecules [19,20,21,23,55].

Together with Unc-14, Unc-51 regulates the subcellular localization of the Netrin receptor/Unc-5 in *C. elegans* [23]. Thus, in unc-14 and unc-51 mutants, the Netrin receptor accumulates in neural cell bodies rather than at axonal termini, causing severe guidance defects in the DD/DV neurons [23]. Unc-51 also interacts with the kinesin-related Vab-8 protein, which regulates anterior-posterior migration of *C. elegans* mechanosensory neurons [55] through the regulation of another Netrin receptor Unc-40/Dcc and the Slit receptor Sax-3/Robo [19,20,21]. Vab-8 controls cell-surface expression of Sax-3/Robo in the growth cones of touch neurons through interaction with Unc-73/Trio, a guanine nucleotide exchange factor for Rac. Consequently, peptide-mediated interference with the Vab-8 and Unc-51 interaction in worm neurons blocked axonal outgrowth and posteriorly directed guidance [20].

In mouse, Unc51.1/ULK1 is expressed in granule cells in the cerebellar cortex, and retroviral infection of immature granule cells with a dominant-negative Unc51.1 caused inhibition of neurite outgrowth both in *vitro* and in *vivo* [13]. Subsequent molecular studies showed that Unc51.1 binds to SynGAP and Syntenin [14], the latter of which, in turn, binds Rab5 GTPase to tether the Unc51.1/SynGAP/Rab5 complex to the vesicular membrane. Immunoelectron microscopy of granule cells provided evidence that Unc51.1 indeed associates with membrane vesicles. Moreover, SynGAP stimulates the GTPase activity of Rab5, and overexpression of SynGAP in cultured cerebellar granule neurons leads to truncated neurites and disorganized vesicular compartments [14]. This suggests that the Unc51.1-containing protein complex governs axonal elongation and pathfinding by modulating the Ras-like GTPase signaling pathway and the Rab5-mediated endocytic pathway in developing neurons.

The importance of Unc-51 in the regulation of vesicle trafficking is further supported by the finding that suppression of Unc-51 activity leads to increased neurite branch formation and shortened axons in cultured mouse dorsal root ganglia neurons [17]. Both Unc-51.1 and Unc-51.2 are localized to vesicular structures in growth cones in sensory axons, in which Unc-51.1 promotes endocytosis of the neurotrophic tyrosine kinase receptor TrkA by a non-clathrin mediated pathway, presumably through the interaction of Unc-51.1 with SynGAP and Rab5 [17]. Moreover, Unc-51.1 also interacts with the Golgi-associated ATPase enhancer of 16 kD (Gafterp2/Gate-16), which is an essential factor for intra-Golgi transport [56]. Unc-51.1 also interacts with the gamma-2 subunit of the GABA-A receptor associated protein (GABARAP), which is again involved in the regulation of receptor trafficking [56]. Together with our findings in the *Drosophila* brain, these studies highlight the functional importance of the Unc-51 family proteins in the endocytic processes that regulate diverse signaling events during axonal elongation, fasciculation, and guidance. Loss of the Unc-51 activity is likely to perturb the trafficking of multiple types of post-Golgi vesicles and lead to severe disruption of the controlled delivery of essential axonal growth/guidance receptors to the different compartments of growing neurons. Elucidation of the exact molecular components that are involved in Unc-51-mediated regulation of vesicle transport is an important subject for future studies. We envisage that studies in *Drosophila* will continue to provide critical insights into the conserved molecular mechanisms of coordinate regulation of membrane vesicle trafficking and axon growth/guidance in developing neurons.

**Materials and Methods**

**Drosophila strains**

Fly strains were maintained on standard medium at 25°C. Construction of the unc-51 null mutants, unc-511 and unc-512, as well as unc-511NT and unc-512K38A transgenic flies has been described previously [16]. The following fly stocks were used: *Khc-eGFP* [44] (a gift from Joseph G. Gindhart), *Klc-mRFP* [45], UAS- *Fas II::TFP* [57] (a gift from Akinao Nose), UAS- *Deam/TM11::GFP* [47] (a gift from Tzumin Lee), UAS- *mCD8::DsRed* [a gift from Yuh Nung Jan], *GHI16-GAL4* [34] (a gift from Reinhard F. Stocker), UAS- *Lamp1::GFP* [58] (a gift from Helmut Kramer), OK107-GAL4 [59,60], 2011-GAL4 [61], elav-GAL4 [62], elav1555 [63], tub-Gal80 FRT24 [32], elav1555 UAS- *mCD8::GFP* hs- hsp [32], UAS-unc- *GFP* [30], UAS- *Nod::β-Gal* [50], UAS- *Synt::GFP* [64], UAS-a-Syb::GFP [64], UAS-Rab3::TFP [65], UAS-mito::GFP [36], UAS-hLC3::GFP [39] and UAS- *Khc::eGFP* (Bloomington Stock Center). The UAS- *Klc-mRFP* stock was newly constructed for this work.

**MARCM mosaic analysis**

Clones were generated using the MARCM method [32]. Egg collection was performed for 4 h on standard food at 25°C. Twenty-four hours after the end of egg collection, newly emerged larvae were heat shocked for 0.5 h (for the induction of MB clones) or 1 h (for the induction of AL-PN clones) at 37°C. Animals were fixed for examination at the wandering larval stage for the larval MB clones or at 2–3-day-old adult stage for the AL-PN clones.
The following genotypes were examined: (1) wild-type MBs: *elav*<sup>1.55</sup> hs-FLP *UAS-mCD8::GFP/+; Gal80 FRT2A/FRT2A; (2) *unc-51*<sup>3/3</sup> MBs: *elav*<sup>1.55</sup> hs-FLP *UAS-mCD8::GFP/+; Gal80 FRT2A/*unc-51<sup>3</sup> FRT2A; (3) wild-type PNs: hs-FLP *UAS-mCD8::GFP/+; *GH146-GAL4*; *Gal80 FRT2A/FRT2A; (4) *unc-51<sup>3/3</sup> PNs: hs-FLP *UAS-mCD8::GFP/+; *GH146-GAL4*; *Gal80 FRT2A/*unc-51<sup>3</sup> FRT2A; (5) *Khc<sup>–/–</sup> MBs: *elav*<sup>1.55</sup> hs-FLP *UAS-mCD8::GFP/+; *Gal80 FRT2A*<sup>25/25</sup> FRT2A.

**Immunostaining with anti-unc-51**

Immunological staining of larval and adult brains was performed as described in Kurosu et al. [20]. The primary antibodies used were: rabbit anti-SYT [60] (1:500; a gift from Troy Littleton), guinea pig anti-Unc-51 (1:100) [16], rabbit anti-β-Gal (1:1000; Chemicon International), mouse anti-nc82 (1:20; Developmental Studies Hybridoma Bank (DSHB)), mouse anti-Fas II (1:5; DSHB), rat anti-N-Cadherin (1:50; DSHB), mouse anti-Dac (1:250; DSHB), rat anti-mCD8A (1:100; Caltag), chicken anti-GFP (1:500; Abcam), fluorescein isothiocyanate-conjugated horseradish peroxidase (1:100; Jackson ImmunoResearch), Alexa 633-conjugated phalloidin (1:100; Invitrogen). The secondary antibodies used were: Alexa 488-conjugated goat anti-rabbit, Alexa 543-conjugated goat anti-mouse (1:500; Molecular Probes). Confocal images were captured using a Zeiss LSM510 confocal microscope and processed with Adobe Photoshop.

**Supporting Information**

**Figure S1 Localization of transport machinery proteins in *unc-51* mutant MBs.** (A, B) Localization of Khc as revealed with *UAS-Khc::GFP*. Arrows indicate accumulations. Arrowhead in B2 denotes mislocalization at the proximal peduncle. (C, D) Localization of Klc as revealed with *UAS-Klc::mRFP*. (E, F) Localization of Tau as revealed with *UAS-tau::GFP*. (G, H) Localization of Futsch as revealed by anti-Futsch staining. The outline of the lobe is demarcated with yellow dashed lines in G3 and H3. Note that Futsch was detected in the cell bodies and the calyx but absent in lobes in both wild type and mutant MBs. (A, C, E, G) wild type. (B, D, F, H) *unc-51*<sup>3/3</sup> mutant. Late third instar larval stage. The calyx is demarcated with a yellow dashed circle. In A-F, marker expression was driven with 201Y-GAL4. Scale bars, 10 μm. (I) Number of Khc::GFP clogs in the lobes. The background accumulation in the wild type lobes, only large clogs (>2 μm<sup>2</sup>) were counted. (J) Number of Klc::GFP clogs in the cell bodies. (K) Number of Klc::mRFP clogs in the lobes. (L) Number of Klc::mRFP clogs in the cell bodies. (TIF)

**Figure S2 Loss of *unc-51* causes axonal targeting defects in adult olfactory PNs.** (A–F) Axonal targeting phenotypes of wild-type (A, C, E) and *unc-51*<sup>3/3</sup> (B, D, F) clones. Arrows indicate accumulations. Arrowhead in B2 indicates mislocalization at the proximal peduncle. (A, B) Rab5::YFP. (C, D) Lamp1::GFP. (E, F) α-Man-II::GFP. Late third instar larval stage. The VA2 glomerulus (white letters in A) is normally innervated by ad-PNs born in the embryonic stage, and is thus uninnervated in the wild-type clone. The DA1, DM5 and DM2 glomeruli (white letters) are normally innervated by the lateral PNs but not the ad-PNs. While denticid targeting to the anterodorsal-type on-target glomeruli was normal, the lateral-type off-target glomeruli (*DA1, DM5 and DM2*) were occasionally innervated by the *unc-51*<sup>3/3</sup> ad-PNs (yellow arrows in B and F). Scale bar, 10 μm. (K–N) Quantification of dendritic phenotypes of ad-PNs. (K, M) Dendritic innervation of on-target glomeruli. Both wild-type and *unc-51*<sup>3/3</sup> clones exhibited complete innervation of all the on-target glomeruli. (L, N) Dendritic innervation of off-target glomeruli. The extent of innervation of the non-target glomeruli was classified into one of three groups: irregular, weak and no input. Sample sizes: wild type (*n* = 9) and *unc-51*<sup>3/3</sup> mutant (*n* = 10). (TIF)

**Figure S3 Loss of *unc-51* causes aberrant dendritic overextension of the olfactory PNs.** (A–F) Dendritic projection patterns of olfactory PNs in the adult brain. (A, B) anterodorsal neuroblast (adNb) clones. (C, D) lateral neuroblast (lNb) clones. (E, F) ventral neuroblast (vNb) clones. (G) Quantification of the dendrite extent of innervation of the non-target glomeruli was classified into one of three groups: irregular, weak and no input. Sample sizes: wild type (*n* = 14), *unc-51* mutant (*n* = 22). Scale bar, 30 μm. (TIF)

**Figure S4 Loss of *unc-51* causes moderate dendritic targeting defects in adult PNs.** (A–F) Dendritic targeting phenotypes of wild-type (A, C, E) and *unc-51*<sup>3/3</sup> (B, D, F) clones. Arrows indicate accumulations. Arrowhead in B2 indicates mislocalization at the proximal peduncle. (A, B) Rab5::YFP. (C, D) Lamp1::GFP. (E, F) α-Man-II::GFP. Late third instar larval stage. The VA2 glomerulus (white letters in A) is normally innervated by ad-PNs born in the embryonic stage, and is thus uninnervated in the wild-type clone. The DA1, DM5 and DM2 glomeruli (white letters) are normally innervated by the lateral PNs but not the ad-PNs. While denticid targeting to the anterodorsal-type on-target glomeruli was normal, the lateral-type off-target glomeruli (*DA1, DM5 and DM2*) were occasionally innervated by the *unc-51*<sup>3/3</sup> ad-PNs (yellow arrows in B and F). Scale bar, 10 μm. (K–N) Quantification of dendritic phenotypes of ad-PNs. (K, M) Dendritic innervation of on-target glomeruli. Both wild-type and *unc-51*<sup>3/3</sup> clones exhibited complete innervation of all the on-target glomeruli. (L, N) Dendritic innervation of off-target glomeruli. The extent of innervation of the non-target glomeruli was classified into one of three groups: irregular, weak and no input. Sample sizes: wild type (*n* = 9) and *unc-51*<sup>3/3</sup> mutant (*n* = 10). (TIF)

**Figure S5 Accumulation of endosomes and Golgi outposts in *unc-51* mutant MBs.** (A, B) Rab5::YFP. (C, D) Lamp1::GFP. (E, F) α-Man-II::GFP. Late third instar larval stage. The VA2 glomerulus (white letters in A) is normally innervated by ad-PNs born in the embryonic stage, and is thus uninnervated in the wild-type clone. The DA1, DM5 and DM2 glomeruli (white letters) are normally innervated by the lateral PNs but not the ad-PNs. While denticid targeting to the anterodorsal-type on-target glomeruli was normal, the lateral-type off-target glomeruli (*DA1, DM5 and DM2*) were occasionally innervated by the *unc-51*<sup>3/3</sup> ad-PNs (yellow arrows in B and F). Scale bar, 10 μm. (K–N) Quantification of dendritic phenotypes of ad-PNs. (K, M) Dendritic innervation of on-target glomeruli. Both wild-type and *unc-51*<sup>3/3</sup> clones exhibited complete innervation of all the on-target glomeruli. (L, N) Dendritic innervation of off-target glomeruli. The extent of innervation of the non-target glomeruli was classified into one of three groups: irregular, weak and no input. Sample sizes: wild type (*n* = 9) and *unc-51*<sup>3/3</sup> mutant (*n* = 10). (TIF)

**Figure S6 Distribution of mitochondria and autophagosomes in wild-type and *unc-51* mutant MBs.** (A, B) mito::GFP. (C, D) hLC3::GFP. Late third instar larval stage. (A, C) wild type. (B, D) *unc-51*<sup>3/3</sup> mutant. MB expression was driven by 201Y-GAL4. Arrows, aberrant accumulations of the marker proteins. Arrowhead, ectopic accumulation in the proximal peduncle. The calyx is demarcated with a yellow dashed circle. Scale bar, 10 μm. (TIF)
Figure S7 Loss of Klc activity causes dendritic and axonal defects in MB neurons. (A–D) Larval MB neuroblast clones generated by the MARCM technique. (A, C) Wild type. (B, D) Klc8e94/8e94 mutant. (C, D) Peduncle sections. Note severe dendritic overextension (arrowheads) in the mutant. Clones were induced by a heat shock at the early first instar stage and labeled with mCD8::GFP (green) driven by elav155 GALA. Mature axons were stained with anti-Fas II (magenta). (E–G) Quantification of the Klc mutant phenotype. (E) Quantification of the calyx (Cx) dendrite overextension phenomenon. Sample sizes: wild type (n = 17), Klc8e94/8e94 wild type (n = 7). (F) Quantification of peduncle fasciculation. Sample sizes: wild type (n = 8), Klc8e94/8e94 mutant (n = 7). (G) Quantification of Fas II mislocalization in the proximal peduncle. Sample sizes: wild type (n = 11), Klc8e94/8e94 mutant (n = 7), and unc-51+/− wild type (n = 10). Scale bars, 10 μm. (TIF)

Figure S8 unc-51 does not interact with RPTPs in axonal fasciculation. Larval MB lobes and peduncle sections (inset) from the late third instar stage. MBs were labeled with anti-Fas II (green) and anti N-Cadherin (magenta) (A, B, C). None of the double heterozygotes unc-51(ptp10D) larval MB neuroblast clones generated by the MARCM technique. (A, C) Wild type. (B, D) Klc8e94/8e94 mutant. (C, D) Peduncle sections. Note severe dendritic overextension (arrowheads) in the mutant. Clones were induced by a heat shock at the early first instar stage and labeled with mCD8::GFP (green) driven by elav155 GALA. Mature axons were stained with anti-Fas II (magenta). (E–G) Quantification of the Klc mutant phenotype. (E) Quantification of the calyx (Cx) dendrite overextension phenomenon. Sample sizes: wild type (n = 17), Klc8e94/8e94 wild type (n = 7). (F) Quantification of peduncle fasciculation. Sample sizes: wild type (n = 8), Klc8e94/8e94 mutant (n = 7). (G) Quantification of Fas II mislocalization in the proximal peduncle. Sample sizes: wild type (n = 11), Klc8e94/8e94 mutant (n = 7), and unc-51+/− wild type (n = 10). Scale bars, 10 μm. (TIF)

Figure S9 Localization of Dscam and Nod in unc-51 mutant MBs. (A–F) Localization of Dscam as revealed with Dscam::GFP antibodies. (B–L) Localization of Nod as revealed with UAS-Nod::bGal. Late third instar stage. MBs were labeled with UAS-mCD8::DRed (A–F) or UAS-mCD8::GFP (G–L) in conjunction with 21F-GALA. The arrowheads in (K) indicate Nod::bGal localization in calyx overextensions. Scale bar, 10 μm. (M, N) Quantification of the number of the Dscam::GFP clogs. (O, P) Quantification of the number of the Nod::bGal clogs. Clog accumulations in the lobes (M, O) or the cell bodies (N, P) were counted. (TIF)

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Author Contributions

Conceived and designed the experiments: HM TT KF-T. Performed the experiments: HM HT MA. Analyzed the data: HM HT MA TT KF-T. Contributed reagents/materials/analysis tools: HT MK TT KF-T. Wrote the paper: HM TT KF-T.

References