Apolipoprotein E prevents the progression of atherosclerosis in Watanabe heritable hyperlipidemic rabbits.

Yamada Nobuhiro, Inoue Ikuo, Kawamura Masako, Harada Kenji, Watanabe Yoshiro, Shimano Hitoshi, Gotoda Takanari, Shimada Masako, Kohzaki Kohichi, Tsukada Toyohiro, Shiomi Masashi, Watanabe Yoshio, Yazaki Yoshio

*The Journal of Clinical Investigation*

Volume 89

Number 2

Page range 706-711

Year 1992-02

(C) The American Society for Clinical Investigation

URL http://hdl.handle.net/2241/102314

doi: 10.1172/JCI115639
Apolipoprotein E Prevents the Progression of Atherosclerosis in Watanabe Heritable Hyperlipidemic Rabbits

Nobuhiro Yamada, Ikuo Inoue, Masako Kawamura, Kenji Harada, Yoshiro Watanabe, Hitoshi Shimano, Takanari Gotoda, Masako Shimada, Kohichi Kohzaki, Toyohiro Tsukada,† Masashi Shiomizu,‡ Yoshiro Watanabe,‡ and Yoshiho Yazaki

3rd Department of Internal Medicine, University of Tokyo, Hongo, Tokyo, Japan; †Sanraku Hospital, Tokyo, Japan; and ‡Institute for Experimental Animals, Kobe University, School of Medicine, Kobe, Japan

Abstract

Apo E plays an important role in plasma lipoprotein metabolism through its high affinity binding to cell surface LDL receptor. In the present study, we studied the effects of apo E on the atherogenic process in Watanabe heritable hyperlipidemic rabbits which are deficient in LDL receptor and an animal model for familial hypercholesterolemia. We isolated apo E from plasma of 1% cholesterol-fed rabbits and administered 10 mg of purified apo E intravenously into five Watanabe heritable hyperlipidemic rabbits three times a week from their age of 2.5 months to 11 months for 8.5 months. After sustained administration of apo E, we found a significant reduction in the accumulation of cholesterol ester in aortae (1.55±0.07 mg/g tissue) as compared to control rabbits (4.32±0.61 mg/g tissue). Supporting this, the percentage of the surface area of the aorta with macroscopic plaque was remarkably decreased in apo E–treated animals (18.8±5.1% vs. 38.8±8.0% in control). Thus, apo E definitely prevented the progression of atherosclerosis in Watanabe heritable hyperlipidemic rabbits. (J. Clin. Invest. 1992; 89:706–711.) Key words: apolipoprotein E • atherosclerosis • Watanabe heritable hyperlipidemic rabbit • low density lipoprotein receptor • cholesterol

Introduction

Apo E functions in plasma lipoprotein metabolism through its interaction with cell surface receptors such as LDL receptor (1–5) and apo E receptor (LDL receptor–related protein) (6, 7). Recently, we and others demonstrated a reduction in plasma cholesterol levels after intravenous administration of apo E into Watanabe heritable hyperlipidemic (WHHL) rabbits (8, 9). Exogenous apo E was incorporated into plasma lipoproteins including VLDL, then VLDL enriched in apo E was removed rapidly from the blood through mutant LDL receptor or apo E receptor in WHHL rabbits, and induced the reduction in plasma cholesterol levels. This function of apo E is supposed to reduce the rate of atherogenesis because many studies have proved the relationship between plasma cholesterol level and the incidence of coronary heart disease (10, 11).

Many studies have suggested important roles of apo E in lipid transport in various tissues, including liver, brain, peripheral nerves, and arterial walls, as well as in plasma (3, 12–14). However, in vivo effects of apo E are not fully understood; in particular, its effects on atherogenesis are controversial. Apo E secreted from macrophages is proposed to function in cholesterol efflux from cholesterol-loaded cells collaborating with HDLs in the interstitial fluid (15, 16); this function is considered to be related to the regression of atherosclerosis by reducing intracellular cholesterol of foam cells. In contrast, apo E–rich lipoproteins such as β-VLDL and hypertriglyceridemic VLDL have been considered to be atherogenic from in vitro studies (17, 18), because macrophages actively endocytose lipoproteins containing several molecules of apo E, and transfer to foam cells in vitro. Our recent study also demonstrated the enhanced uptake of apo E–rich VLDL in human monocyte-derived macrophages (19). The apo E secreted from macrophages can be incorporated into VLDL, and VLDL enriched in apo E is actively taken up by macrophages with a high affinity to cell surface receptors. Both functions of apo E that are related to cholesterol efflux from macrophages and cholesterol uptake by macrophages, may play an important role in pathophysiology of atherosclerosis. To understand an overall effect of apo E on the process of atherosclerosis in vivo, we administered rabbit apo E into WHHL rabbits, an animal model of early atherosclerosis, for 8.5 months in the present study.

Methods

Animal experiments. 10 male WHHL rabbits from six litters, weighing ~ 1.5 kg, were fed laboratory chow ad lib. and housed individually before and during the experiments.

Apo E. Apo E was purified from plasma of rabbits fed a diet containing 1% cholesterol by weight for > 1 mo. Plasma from blood containing 0.1% EDTA and 0.02% sodium azide was ultracentrifuged at 100,000 rpm for 20 h at 16°C in a 50.2 rotor of an ultracentrifuge at 45,000 rpm (Beckman Instruments, Inc., Palo Alto, CA); the supernatant lipoproteins were recentrifuged under the same condition to obtain lipoproteins rich in apo E. All steps of purification were carried out at 4°C. After delipidation of the lipoproteins by extraction with 200
vol of ethanol-ether (1:1) four times and evaporating the solvent under N₂, apolipoproteins were dissolved in 2 mM sodium phosphate, pH 7.4 containing 6 M urea, dialyzed against 2 mM sodium phosphate containing 50 mM NaCl and 1 M urea, pH 7.4 (buffer A), and applied to a column (2 × 16 cm) of heparin-Sepharose 4B (Pharmacia Fine Chemicals, Uppsal, Sweden) preequilibrated with 2 mM sodium phosphate containing 50 mM NaCl, pH 7.4 (buffer B). The column was first washed with 100 ml of buffer A, then with 1,200 ml of buffer B, and finally with 2 mM sodium phosphate containing 500 mM NaCl and 6 M urea, pH 7.4, to elute apo E. The purity of apo E was evaluated by SDS polyacrylamide gel electrophoresis (8) and purity was > 99%. After dialysis against water and lyophilization, purified apo E was stored at −80°C. One day before use, apo E was dissolved in 6 M guanidine chloride, pH 7.4, at a concentration of ~ 20 mg/ml, and finally dialyzed extensively against 150 mM NaCl. We prepared ~ 5 g of apo E for the current study, which was pooled and used as a single lot.

In vivo studies. The experiment was started at the age of 2.5 mo in WHHL rabbits. 10 mg of sterile rabbit apo E in 0.5 ml of 0.15 M NaCl was administered intravenously by a bolus injection through a marginal ear vein every Monday, Wednesday, and Friday morning for 8.5 mo. In control rabbits, 0.5 ml of 0.15 M NaCl was injected to estimate the natural course of atherosclerosis. Each animal group consisted of five rabbits from five litters. Before starting the experiment, and 2 and 6 mo after starting the experiment, 2 ml of blood was taken to measure plasma cholesterol (20), triglyceride levels by enzymatic methods (21), and HDL-cholesterol level by precipitation method (22), one day after apo E injection. After 8.5 mo of the experiment, 5 ml of blood was drawn to evaluate blood cell counts, liver functions, kidney functions, and plasma lipid levels.

Extent of atherosclerotic lesions. Animals were killed under deep anesthesia with xylazine at 7 mg/kg and ketamine at 80 mg/kg intramuscularly. The entire aorta from the aortic valve to the iliac bifurcation was removed and cleaned of loose adventitial tissue. For the analysis, the aorta was divided into three segments at the levels of the first intercostal artery and celiac artery (aortic arch, descending thoracic aorta, and abdominal aorta). We measured contents of cholesteryl ester in each segment of aorta to confirm macroscopic observations. Each segment of aorta was dried by lyophilization and stored at −20°C until analysis. Total lipid extracts were prepared by cutting each aorta into small pieces and homogenizing three times in chloroform/methanol (2:1) in a tapered glass–glass homogenizer. Determinations of total and free cholesterol in the extracts were performed in triplicate by enzymatic assay.

Furthermore, we determined the extent of lesion area with macroscopic atherosclerotic plaques. The aorta was opened longitudinally, covered with a clear plastic sheet, and copied with a magnification of 2, then the areas corresponding to atheromatous plaques on the copied paper were delineated. The delineated areas of atheromatous lesions were scanned and estimated by computer. The lesion areas within each segment were summed and the extent of lesions was expressed as percentage of surface area with atheromatous plaque.

Statistical analysis. All data were expressed as mean±SD. We compared data between groups by analysis of variance, and the statistical difference was determined by Student’s t tests.

Results

Plasma lipid levels. Plasma cholesterol levels and triglyceride levels were over 400 mg/dl and 300 mg/dl, respectively, and plasma HDL-cholesterol levels were 6–9 mg/dl as shown in Table I. There were no significant differences between the two groups, but we found decrease in plasma cholesterol levels in both animal groups after the experiment as compared to before the experiment. Mean plasma apo E level increased 54% 3 min after apo E injection as compared to preinjection level, and the half-life of injected apo E was 1.5 h in a different set of studies using WHHL rabbits (n = 3). We checked daily intake of diet every day; there were no significant differences in the mean food consumption. Body weight gains during the experiment were not significantly different between the two groups. We did not find any differences in peripheral blood cell counts, liver functions, kidney functions and plasma lipid levels.

Table I. Changes in Plasma Cholesterol, Triglyceride, and HDL-Cholesterol Levels (mg/dl) after the Administration of Rabbit apo E (n = 5) and Saline (n = 5) in Male WHHL Rabbits

<table>
<thead>
<tr>
<th>Group</th>
<th>0</th>
<th>2</th>
<th>6</th>
<th>8.5</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cholesterol (mg/dl)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Apo E</td>
<td>502±33</td>
<td>490±39</td>
<td>431±83</td>
<td>408±42</td>
</tr>
<tr>
<td>Control</td>
<td>518±15</td>
<td>512±19</td>
<td>440±65</td>
<td>418±46</td>
</tr>
<tr>
<td>Triglyceride (mg/dl)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Apo E</td>
<td>415±91</td>
<td>422±116</td>
<td>325±67</td>
<td>328±59</td>
</tr>
<tr>
<td>Control</td>
<td>365±82</td>
<td>368±122</td>
<td>387±93</td>
<td>378±181</td>
</tr>
<tr>
<td>HDL-cholesterol (mg/dl)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Apo E</td>
<td>7.0±2.2</td>
<td>6.0±1.7</td>
<td>6.0±0.86</td>
<td>6.0±1.5</td>
</tr>
<tr>
<td>Control</td>
<td>8.2±1.6</td>
<td>6.8±2.0</td>
<td>6.7±1.9</td>
<td>7.9±1.0</td>
</tr>
</tbody>
</table>

Serum lipid levels were measured by enzymatic assay. Male WHHL rabbits weighing ~1.5 kg were fed laboratory chow ad lib. Apo E was purified from plasma of rabbits fed a diet containing 1% cholesterol by weight for more than one month (6). We prepared ~ 5 g of apo E for the current study, which was pooled and used as a single lot. The experiment was started at the age of 2.5 months in WHHL rabbits. We checked the daily intake of diet every day; there were no significant differences in the mean food consumption. Body weight gains during the experiment were not significantly different between the two groups. We did not find any differences in peripheral blood cell counts, liver functions or kidney functions between the groups after 8.5 months of treatment.

Values are expressed as mean ± SD.

Apolipoprotein E Prevents the Progression of Atherosclerosis in Watanabe Heritable Hyperlipidemic Rabbits
specific for smooth muscle cells (HHF 35) and macrophages (RAM 11) (23), we found the similar distribution of both cell types in the atherosclerotic lesions of both groups. As compared to the percentage lesion area, the cholesterol ester content of the aortae were extremely reduced in animals treated with apo E, and those variations among animals were much smaller. When the ratios of cholesterol ester accumulation to lesion area were compared between two groups, we could not find significant differences in the ratios between the two groups as shown in Fig. 2.

**Discussion**

In recent studies, we have suggested the possible involvement of apo E in the process of atherosclerosis (8, 19). The present study tried to determine the effects of apo E on its process in vivo. One of the best ways to evaluate the effects of agents that can affect the rate of atherogenesis in vivo is long-term administration of such agents to young WHHL rabbits that have not yet developed atherosclerotic lesions (24, 25). Plasma cholesterol levels in WHHL rabbits are > 400 mg/dl, and severe, progressive atherosclerosis occurs after the age of 4 months (26). In the present study, we injected a relatively low dose of apo E (10 mg/animal per day) into young WHHL rabbits (2.5 months) as compared to the previous study, in which apo E was shown to significantly reduce plasma cholesterol levels after treatment with 30–40 mg of apo E. Minimal effects on plasma cholesterol level have been demonstrated in WHHL rabbits after intravenous injection of 10 mg of apo E (27). During the experimental period, there were no significant differences in plasma lipid levels between the two groups.

Apo E definitely reduced the rate of atherogenesis after 8.5 months of treatment in WHHL rabbits. The cholesterol accumulation and lesion area were reduced approximately one-third and one-half, respectively, in animals treated with apo E as compared to those with saline. The cholesterol ester content of each aortic segment was well correlated to percentage lesion area ($r = 0.884$). Similar retarded progression of atherosclerosis without significant change in plasma cholesterol level was dem-

**Table II. Contents of Cholesterol Ester of Each Aortic Arch, Descending Thoracic Aorta, Abdominal Aorta, and Total Aorta in WHHL Rabbits Treated with Rabbit apo E or Saline**

<table>
<thead>
<tr>
<th>Group</th>
<th>Aortic arch</th>
<th>Thoracic</th>
<th>Abdominal</th>
<th>Total aorta</th>
<th>Cholesterol ester/dry tissue</th>
</tr>
</thead>
<tbody>
<tr>
<td>Apo E-1</td>
<td>0.20</td>
<td>0.10</td>
<td>0.08</td>
<td>0.38</td>
<td>1.58</td>
</tr>
<tr>
<td>Apo E-2</td>
<td>0.21</td>
<td>0.11</td>
<td>0.03</td>
<td>0.35</td>
<td>1.52</td>
</tr>
<tr>
<td>Apo E-3</td>
<td>0.21</td>
<td>0.11</td>
<td>0.10</td>
<td>0.42</td>
<td>1.50</td>
</tr>
<tr>
<td>Apo E-4</td>
<td>0.19</td>
<td>0.08</td>
<td>0.02</td>
<td>0.31</td>
<td>1.48</td>
</tr>
<tr>
<td>Apo E-5</td>
<td>0.20</td>
<td>0.18</td>
<td>0.05</td>
<td>0.43</td>
<td>1.65</td>
</tr>
<tr>
<td>Mean±SD</td>
<td>0.20±0.01*</td>
<td>0.12±0.04*</td>
<td>0.06±0.03*</td>
<td>0.38±0.05*</td>
<td>1.55±0.07*</td>
</tr>
</tbody>
</table>

* P < 0.001. Cholesterol ester/dry tissue indicates contents of cholesterol ester (in milligrams) per weight of dry total aorta (in grams). The apo E group was compared to the saline group. Each segment of aorta was dried by lyophilization and stored at −20°C until analysis. Total lipid extracts were prepared by cutting each aorta into small pieces and homogenizing it three times in chloroform/methanol (2:1) in a tapered glass-glass homogenizer. Determinations of total and free cholesterol in the extracts were performed in triplicate by enzymatic assay.

**Table III. Percentages of Intimal Surface Area with Atheromatous Plaque of Each Aortic Arch, Descending Thoracic Aorta, Abdominal Aorta, and Total Aorta in WHHL Rabbits Treated with Rabbit apo E or Saline**

<table>
<thead>
<tr>
<th>Group</th>
<th>Aortic arch</th>
<th>Thoracic</th>
<th>Abdominal</th>
<th>Total aorta</th>
</tr>
</thead>
<tbody>
<tr>
<td>Apo E-1</td>
<td>30</td>
<td>9</td>
<td>9</td>
<td>14</td>
</tr>
<tr>
<td>Apo E-2</td>
<td>33</td>
<td>11</td>
<td>5</td>
<td>14</td>
</tr>
<tr>
<td>Apo E-3</td>
<td>41</td>
<td>15</td>
<td>9</td>
<td>18</td>
</tr>
<tr>
<td>Apo E-4</td>
<td>34</td>
<td>30</td>
<td>13</td>
<td>25</td>
</tr>
<tr>
<td>Apo E-5</td>
<td>42</td>
<td>24</td>
<td>8</td>
<td>23</td>
</tr>
<tr>
<td>Mean±SD</td>
<td>36.0±5.2*</td>
<td>17.8±8.9*</td>
<td>8.8±2.9*</td>
<td>18.8±5.1*</td>
</tr>
</tbody>
</table>

* P < 0.001; † P < 0.02; ‡ P < 0.002. The apo E group was compared to the control group. The entire aorta from the aortic valve to the iliac bifurcation was removed and opened longitudinally. The aorta, covered with a clear plastic sheet, was copied with a magnification of 2 and the area corresponding to atheromatous plaques on the copied paper was delineated. The delineated areas of atheromatous lesions were estimated as lesioned area. The aorta was divided into three segments at the levels of the first intercostal artery and celiac artery (aortic arch, descending thoracic aorta, and abdominal aorta).
Figure 1. Aortic specimens obtained from WHHL rabbits. The upper two aortae are from rabbits (saline-2 and saline-3) in the saline group. The lower two aortae are from rabbits (apo E-1 and apo E-2) in the apo E group. 10 mg of sterile rabbit apo E in 0.5 ml of 0.15 M NaCl was administered intravenously by a bolus injection through a marginal ear vein every Monday, Wednesday, and Friday morning for 8.5 mo. Animals were killed under deep anesthesia with xylazine at 7 mg/kg and ketamine at 80 mg/kg intramuscularly. The entire aorta from the aortic valve to the iliac bifurcation was removed and cleaned of loose adventitial tissue. In the saline group, 0.5 ml of 0.15 M NaCl was injected as control.

Apolipoprotein E Prevents the Progression of Atherosclerosis in Watanabe Heritable Hyperlipidemic Rabbits

Apolipoprotein E (apo E) has been found to be effective in correcting the imbalance of factors which determine the rate of atherogenesis. These factors include accumulation of cholesterol in and efflux of cholesterol from macrophages, recruitment of circulating monocytes, and functions of endothelial cells and smooth muscle cells. If there were an imbalance in these factors, such as high plasma cholesterol levels and low HDL cholesterol levels, the rate of atherogenesis would be accelerated. The results of our study suggest that apo E is effective in correcting the imbalance of factors which determine the rate of atherogenesis. Because there were no significant differences in plasma cholesterol levels between the two groups during the experimental period, the effects of apo E on the process of atherosclerosis might not be related to plasma cholesterol levels, although there were minimal and acute effects of low dose apo E on plasma cholesterol levels.

Apo E secreted from macrophages in peripheral tissues is suggested to be involved in cholesterol transport from peripheral tissues to liver, which is called a reverse cholesterol transport system. Cholesterol-loaded cells such as macrophages secrete apo E to facilitate cholesterol transport. This transport system may play a role in the prevention of atherosclerosis by reducing the amount of cholesterol available for accumulation in the arterial wall.
phages release their cholesterol to either HDL or other acceptor proteins including apo E and apo AI in the interstitial fluid. In vitro studies showed that the presence of apo E facilitates the acquisition of cholesterol by HDL (15), and apo E itself actively accepts cholesterol from cholesterol-loaded cells (28). HDL with apo E or apo E–lipid complex, after accepting cellular cholesterol, can bind to hepatic receptors including LDL receptors and apo E receptors by virtue of the presence of apo E on lipoproteins. Thus, the extracellular cholesterol accumulating in the subendothelial foam cells can be transported with apo E to the liver, where it is removed by receptor-mediated endocytosis. Because HDL concentrations are low in WHHL rabbits (6–9 mg/dl) as compared to normal rabbits (20–25 mg/dl) (29), exogenous apo E either associated with or without HDL (HDL with apo E or free form of apo E) may help the function of HDL in the reverse cholesterol transport system, and may accelerate the turnover of this system.

In addition to the effects of apo E on lipoprotein metabolism, apo E influences the functions of lymphocytes and smooth muscle cells, which are major cell components of arterial walls. Apo E inhibits mitogenic stimulation of lymphocytes by binding to specific sites on the surface of lymphocytes (30), and apo E produced by smooth muscle cells may be involved in modulating cell proliferation and differentiation (3). It is possible that cellular functions influenced by the presence of apo E play important roles in reducing the rate of atherogenesis in WHHL rabbits. In addition, macrophage function is important in foam cell formation because an early atherosclerotic lesion is characterized by the presence of macrophage-derived foam cells. Recently, it has been demonstrated that LDL receptor-related protein, a candidate for apo E receptor, is identical to α2 macroglobulin receptor that appears on macrophages (31). The function of macrophages can be altered by the presence of apo E through its binding to α2 macroglobulin receptor. The altered function of macrophages may influence the mechanism of foam cell generation in the arterial wall.

In vitro studies demonstrated that lipoproteins rich in apo E can be actively taken up by macrophages and generate foam cells (17, 18). When we studied the uptake of VLDL enriched in apo E which had been incubated with excess apo E in human monocyte-derived macrophages and hep G2 cells, enrichment of apo E doubled the lipoprotein uptake to a similar extent in both cell types, indicating no preferential uptake of lipoproteins enriched in apo E in macrophages (32). Current results demonstrating no acceleration of the rate of atherogenesis in WHHL rabbits, also suggest that sustained apo E injection does not specifically enhance foam cell formation through the mechanism of the uptake of apo E–rich lipoproteins by macrophages. In the present study, we have investigated an overall effect of apo E in vivo. Although the precise mechanism of apo E action in the prevention of atherosclerosis is yet unknown, exogenous apo E may influence the rate of atherogenesis by not only activating the reverse cholesterol transport system but also modifying cellular functions in the arterial wall, rather than by enhancing the cholesterol accumulation in the arterial wall.

Acknowledgments

We thank Dr. F. Takaku, Dr. T. Murase, and Dr. Y. Akanuma for encouragement and Dr. S. Ishibashi, Dr. H. Mokuno, Dr. K. Takahashi, and Dr. N. Mori for helpful discussions. This study was supported by Terumo foundation.

References


