

Elucidating the Significance of Rhopty Proteins  
Associated with the Survival Strategy of *Toxoplasma*  
*gondii* in the Host

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## **Abstract**

Most intracellular pathogens modify host cell functions to avoid host's defense system and replicate in the host. *Toxoplasma gondii*, which is an intracellular apicomplexan parasite, also changes the function of host cells. These modifications are represented by the inhibition of parasite clearance, interpretation of host gene expression and recruitment of host organelles. Rhopty kinase family proteins (ROPs), which are secreted into host cells from apicomplexan specialized organelles rhoptries, are involved in clearance inhibition of parasites and genetic interpretation in host nuclei. In this study, I analyzed ROPs associated with the functional modifications of host cells in Japanese isolates of *T. gondii* and searched ROPs participated in host mitochondrial recruitment. I could explain abnormal virulence of Japanese *T. gondii* in mice by revealing genetic structure of the ROP5, ROP16 and ROP18, which are related to virulence of parasites by inhibiting clearance of parasites and controlling host gene expression, and identify ROP39 as the novel factor of host mitochondrial recruitment. ROPs are possibly promising drug targets of *T. gondii* considering the importance in *T. gondii* infections.

## **Chapter 1. General Introduction**

*Toxoplasma gondii* is an apicomplexan parasite that can infect almost all warm-blooded animals, and nearly one-third of the world's population is infected with this parasite (Grigg and Sundar, 2009). *T. gondii* causes a latent infection in most humans, but leads to lethal diseases, including encephalitis, only in immunosuppressed people due to acquired immunodeficiency syndrome (AIDS) or organ transplantation. In pregnant women, initial infection with *T. gondii* may cause fetuses to encounter the parasites through vertical transmission, and this can result in serious symptoms such as retinochoroiditis, hydrocephalus and psychomotor retardation (Montoya and Liesenfeld, 2004). When *T. gondii* invades host cells, it secretes rhoptry kinase family proteins (ROPs) from rhoptry organelles into the cells. Secreted ROPs are localized and function in host nuclear or on parasitophorous vacuole (PV), which contains an intracellular parasite. About 60 ROPs have been identified and they commonly possess a N-terminal signal peptide and a serine/threonine kinase domain. *T. gondii* modifies various functions of host cells possibly for establishing ideal environment to replicate in the cells. The inhibition of parasite clearance, interpretation of host gene expression and recruitment of host organelles are representative of the functional modification. ROP18 phosphorylates immunity-related GTPases (IRGs) on PV with the aid of ROP5 to inhibit parasite clearance (Fentress *et al.*, 2010; Behnke *et al.*, 2012). ROP16 interprets host gene expression associated with immune response through the phosphorylation of STAT3 and STAT6 (Saeij *et al.*, 2007). The phenotypes of *T.*

*gondii* is classified into three types (type I, type II and type III). *T. gondii* of each type shows different virulence to mice (Sibley and Boothroyd, 1992). The differences of these virulences are attributed to distinct genetic variations of ROP5, ROP16 and ROP18 in the parasites' genome (Saeij *et al.*, 2006; Taylor *et al.*, 2006; Saeij *et al.*, 2007; Behnke *et al.*, 2011; Reese *et al.*, 2011). Elucidating molecular mechanism associated with survival strategy of *T. gondii* in the host, I dissected the function of ROPs, which are thought to be essential factors for parasite's survival.

#### **4. General Discussion**

The identification of TgCatJpOk3 and TgCatJpOk4 showed that traditional phenotyping based on type I-III could not estimate the virulence of parasite accurately. In this study, the analysis for ROPs enhancing virulence of *T. gondii* worked well to reveal the cause of high virulence of TgCatJpOk4. Therefore, the information of ROPs are thought to be effective for predicting virulence of *T. gondii*.

The potency of host mitochondrial recruitment varies in type I-III parasites, in order of type I  $\geq$  type II  $\geq$  type III. Considering that the genetic structures of ROP5, ROP16 and ROP18 contribute to virulence of parasites in mice, I reasoned that the genetic structure of ROP39 in each type of *T. gondii* might contribute to different strength of the mitochondrial recruitment between type I-III strains. I searched the amino acid sequence of type I-III ROP39 and found the amino acid sequence identities are 95.07% (type I vs type II), 95.24% (type II vs type III) and 99.83% (type III vs type I), which indicates the amino acid sequence of type II is different from those of type I and III (Fig. 14). Furthermore, I discovered that transcription level of ROP39 was lower in type II strains than in type I and type III strains by referring ToxoDB (<http://toxodb.org>), which indicated that transcription levels of ROP39 correlated with the ability of host mitochondrial recruitment. Taken together, amino acid substitutions or/and transcription levels of ROP39 may determine the strength of host mitochondrial recruitment.

ROPs are participated in various process indispensable for establishing parasite infection. There are still a lot of functionally unknown ROPs. Therefore, understanding comprehensively the function of ROPs in host cells may lead to elucidating the mechanism to establish the infection of *T. gondii*. ROPs are fascinating drug targets because human beings do not have these molecules. In fact, inhibitors of kinase activity of ROPs are studied to be used for the treatment of *T. gondii* (D. Sibley, personal communication). Therefore, the accumulation of knowledge related to ROPs may provoke a development of effective drugs against *T. gondii*.

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## Reference

- Ajzenberg D, Bañuls AL, Su C, Dumètre A, Demar M, Carme B, et al. Genetic diversity, clonality and sexuality in *Toxoplasma gondii*. *Int J Parasitol.* 2004;34: 1185–1196.
- Ajzenberg D. 1995–2015: It is time to celebrate 20 years of (intensive) genotyping of *Toxoplasma gondii* strains. *Future Microbiol.* 2015;10: 689–691.
- Behnke MS, Khan A, Wootton JC, Dubey JP, Tang K, Sibley LD. Virulence differences in *Toxoplasma* mediated by amplification of a family of polymorphic pseudokinases. *Proc Natl Acad Sci.* 2011;108: 9631–9636.
- Behnke MS, Fentress SJ, Mashayekhi M, Li LX, Taylor GA, Sibley LD. The Polymorphic pseudokinase ROP5 controls virulence in *Toxoplasma gondii* by regulating the active kinase ROP18. *PLoS Pathog.* 2012;8. doi:10.1371/journal.ppat.1002992
- Bendtsen JD, Nielsen H, von Heijne G, Brunak S. Improved prediction of signal peptides: SignalP 3.0. *J Mol Biol.* 2004;340: 783–795.
- Boothroyd JC, Dubremetz JF. Kiss and spit: the dual roles of *Toxoplasma* rhoptries. *Nat Rev Microbiol.* 2008;6: 79–88.
- Bossi P, Caumes E, Paris L, Dardé ML, Bricaire F. *Toxoplasma gondii*-associated Guillain-Barré syndrome in an immunocompetent patient. *J Clin Microbiol.* 1998;36: 3724–3725.
- Carruthers VB, Håkansson S, Giddings OK, Sibley LD. *Toxoplasma gondii* uses sulfated proteoglycans for substrate and host cell attachment. *Infect Immun.* 2000;68: 4005–4011.
- Cong L, Ran FA, Cox D, Lin S, Barretto R, Habib N, et al. Multiplex genome engineering using CRISPR/Cas systems. *Science.* 2013;339: 819–824.
- Crawford MJ, Thomsen-Zieger N, Ray M, Schachtner J, Roos DS, Seeber F. *Toxoplasma gondii* scavenges host-derived lipoic acid despite its de novo synthesis in the apicoplast. *EMBO J.* 2006;25: 3214–3222.
- Dardé ML, Bouteille B, Pestre-Alexandre M. Isoenzyme analysis of 35 *Toxoplasma gondii* isolates and the biological and epidemiological implications. *J Parasitol.* 1992;78: 786–794.
- Dardé M. Biodiversity in *Toxoplasma gondii*. *Curr. Top. Microbiol. Immunol.* 1996;219: 27–41.
- Dardé ML, Villena I, Pinon JM, Beguinot I. Severe toxoplasmosis caused by a *Toxoplasma gondii* strain with a new isoenzyme type acquired in French Guyana. *J Clin Microbiol.* 1998;36: 324.

- Demar M, Ajzenberg D, Serrurier B, Darde ML, Carme B, Dardé ML. Atypical *Toxoplasma gondii* strain from a free-living jaguar (*Panthera onca*) in French Guiana. Am J Trop Med Hyg. 2008;78: 195–197.
- Dubey JP, Shen SK, Kwok OC, Frenkel JK. Infection and immunity with the RH strain of *Toxoplasma gondii* in rats and mice. J Parasitol. 1999;85, 657-662.
- Dubey JP, Navarro IT, Sreekumar C, Dahl E, Freire RL, Kawabata HH, et al. *Toxoplasma gondii* Infections in Cats from Paraná, Brazil: Seroprevalence, tissue distribution, and biologic and genetic characterization of isolates. J Parasitol. 2004;90: 721–726.
- Dubey JP, Sundar N, Pineda N, Kyvsgaard NC, Luna LA, Rimbaud E, et al. Biologic and genetic characteristics of *Toxoplasma gondii* isolates in free-range chickens from Nicaragua, Central America. Vet Parasitol. 2006a;142: 47–53.
- Dubey JP, Su C, Cortés JA, Sundar N, Gomez-Marin JE, Polo LJ, et al. Prevalence of *Toxoplasma gondii* in cats from Colombia, South America and genetic characterization of *T. gondii* isolates. Vet Parasitol. 2006b;141: 42–47.
- Dubey JP, Cortés-Vecino JA, Vargas-Duarte JJ, Sundar N, Velmurugan G V., Bandini LM, et al. Prevalence of Toxoplasma gondii in dogs from Colombia, South America and genetic characterization of *T. gondii* isolates. Vet Parasitol. 2007a;145: 45–50.
- Dubey JP, Applewhaite L, Sundar N, Velmurugan GV, Bandini LA, Kwok OCH, et al. Molecular and biological characterization of *Toxoplasma gondii* isolates from free-range chickens from Guyana, South America identified several unique and common parasite genotypes. Parasitol. 2007b;134: 1559-1565.
- Dubey JP, Sundar N, Gennari SM, Minervino AHH, Farias NA da R, Ruas JL, et al. Biologic and genetic comparison of *Toxoplasma gondii* isolates in free-range chickens from the northern Pará state and the southern state Rio Grande do Sul, Brazil revealed highly diverse and distinct parasite populations. Vet Parasitol. 2007c;143: 182–188.
- Dubey JP, Zhu XQ, Sundar N, Zhang H, Kwok OCH, Su C. Genetic and biologic characterization of *Toxoplasma gondii* isolates of cats from China. Vet Parasitol. 2007d;145: 352–356.
- Dubey JP, Sundar N, Hill D, Velmurugan G V., Bandini LA, Kwok OCH, et al. High prevalence and abundant atypical genotypes of Toxoplasma gondii isolated from lambs destined for human consumption in the USA. Int J Parasitol. 2008;38: 999–1006.
- Dubey JP, Velmurugan G V, Morales J a, Arguedas R, Su C. Isolation of *Toxoplasma gondii* from the keel-billed toucan (*Ramphastos sulfuratus*) from Costa Rica. J Parasitol. 2009;95: 467–468.

- Escoll P, Song OR, Viana F, Steiner B, Lagache T, Olivo-Marin JC, *et al.* *Legionella pneumophila* modulates mitochondrial dynamics to trigger metabolic repurposing of infected macrophages. *Cell Host Microbe*. 2017;22: 302-316.
- Falush D, Stephens M, Pritchard JK. Inference of population structure using multilocus genotype data: Linked loci and correlated allele frequencies. *Genetics*. 2003;164: 1567–1587.
- Fentress SJ, Behnke MS, Dunay IR, Mashayekhi M, Rommereim LM, Fox BA, *et al.* Phosphorylation of immunity-related GTPases by a *Toxoplasma gondii*-secreted kinase promotes macrophage survival and virulence. *Cell Host Microbe*. 2010;8: 484–495.
- Fukasawa Y, Tsuji J, Fu SC, Tomii K, Horton P, Imai K. MitoFates: Improved prediction of mitochondrial targeting sequences and their cleavage sites. *Mol Cell Proteomics*. 2015;14: 1113–1126.
- Grigg ME, Sundar N. Sexual recombination punctuated by outbreaks and clonal expansions predicts *Toxoplasma gondii* population genetics. *Int J Parasitol*. 2009;39: 925–933.
- Guerra AJ, Zhang O, Bahr CME, Huynh MH, DelProposto J, Brown WC, *et al.* Structural basis of *Toxoplasma gondii* perforin-like protein 1 membrane interaction and activity during egress. *PLoS Pathog*. 2018;14. e1007476
- Heigwer F, Kerr G, Boutros M. E-CRISP: fast CRISPR target site identification. *Nat Methods*. 2014;11: 122–123.
- Horwitz MA, John A, Hartford GL, Fellowship F. Formation of a novel phagosome by the Legionnaires' disease bacterium (*Legionella pneumophila*) in the human monocytes. *J Exp Med*. 1983;158: 1319-1331.
- Howe DK, Sibley LD. *Toxoplasma gondii* comprises of parasite three clonal lineages: Correlation with Human Disease Genotype. *J Infect Dis*. 1995;172: 1561–1566.
- Jinek M, Chylinski K, Fonfara I, Hauer M, Doudna JA, Charpentier E. A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity. *Science*. 2012;337: 816–822.
- Kafsack BF, Pena JD, Coppens I, Ravindran S, Boothroyd JC, Carruthers VB. Rapid membrane disruption by a perforin-like protein facilitates parasite exit from host cells. *Science*. 2009;323: 530-533.
- Katoh K, Standley DM. MAFFT multiple sequence alignment software version 7: Improvements in performance and usability. *Mol Biol Evol*. 2013;30: 772–780. 2017;2: 1–14.
- Khan A, Jordan C, Muccioli C, Vallochi AL, Rizzo L V., Belfort R, *et al.* Genetic divergence

- of *Toxoplasma gondii* strains associated with ocular toxoplasmosis, Brazil. *Emerg Infect Dis.* 2006;12: 942–949.
- Khan A, Taylor S, Ajioka JW, Rosenthal BM, Sibley LD. Selection at a single locus leads to widespread expansion of *Toxoplasma gondii* lineages that are virulent in mice. *PLoS Genetics.* 2009;5. doi:10.1371/journal.pgen.100040436
- Khan A, Dubey JP, Su C, Ajioka JW, Rosenthal BM, Sibley LD. Genetic analyses of atypical *Toxoplasma gondii* strains reveal a fourth clonal lineage in North America. *Int J Parasitol.* 2011;41: 645–655.
- Kyan H, Taira M, Yamamoto A, Inaba C, Zakimi S. Isolation and characterization of *Toxoplasma gondii* genotypes from goats at an abattoir in Okinawa. *Jpn J Infect Dis.* 2012;65: 167–170.
- Lagal V, Binder EM, Huynh MH, Kafsack BF, Harris PK, Diez R, et al. *Toxoplasma gondii* protease TgSUB1 is required for cell surface processing of micronemal adhesive complexes and efficient adhesion of tachyzoites. *Cell Microbiol.* 2010;12: 1792–1808.
- Langmead B, Salzberg SL. Fast gapped-read alignment with Bowtie 2. *Nat Methods.* 2012;9: 357–359.
- Lehmann T, Marcet PL, Graham DH, Dahl ER, Dubey JP. Globalization and the population structure of *Toxoplasma gondii*. *Proc Natl Acad Sci.* 2006;103: 11423–11428.
- Letunic I, Doerks T, Bork P. SMART: Recent updates, new developments and status in 2015. *Nucleic Acids Res.* 2015;43: 257–260.
- Letunic I, Bork P. 20 years of the SMART protein domain annotation resource. *Nucleic Acids Res.* 2018;46: 493–496.
- Lorenzi H, Khan A, Behnke MS, Namasivayam S, Swapna LS, Hadjithomas M, et al. Local admixture of amplified and diversified secreted pathogenesis determinants shapes mosaic *Toxoplasma gondii* genomes. *Nat Commun.* 2016;7. doi:10.1038/ncomms10147
- Matsumoto A, Bessho H, Uehira K, Suda T. Morphological studies of the association of mitochondria with chlamydial inclusions and the fusion of chlamydial inclusions. *J Electron Microsc.* 1991;40: 356–363.
- Matui T, Morii T, Iijima T, Kobayashi F, Fujino T. Surveys of coccidium infection in kittens and puppies in Tama area, Tokyo. *J. Kyorin Med. Soc.* 1986;17: 19–23.
- Mercier A, Devillard S, Ngoubangoye B, Bonnabau H, Bañuls AL, Durand P, et al. Additional haplogroups of *Toxoplasma gondii* out of Africa: Population structure and mouse-virulence of strains from Gabon. *PLoS Negl Trop Dis.* 2010;4. doi:10.1371/journal.pntd.0000876

- Miller MA, Grigg ME, Kreuder C, James ER, Melli AC, Crosbie PR, *et al.* An unusual genotype of *Toxoplasma gondii* is common in California sea otters (*Enhydra lutris nereis*) and is a cause of mortality. *Int J Parasitol.* 2004;34: 275–284.
- Montoya JG, Liesenfeld O. Toxoplasmosis. *Lancet.* 2004;363: 1965–1976.
- Nagamune K, Beatty WL, Sibley LD. Artemisinin induces calcium-dependent protein secretion in the protozoan parasite *Toxoplasma gondii*. *Eukaryot Cell.* 2007;6: 2147–2156.
- Pena HFJ, Gennari SM, Dubey JP, Su C. Population structure and mouse-virulence of *Toxoplasma gondii* in Brazil. *Int J Parasitol.* 2008;38: 561–569.
- Pernas L, Boothroyd JC. Association of host mitochondria with the parasitophorous vacuole during *Toxoplasma* infection is not dependent on rhoptry proteins ROP2/8. *Int J Parasitol.* 2010;40: 1367–1371.
- Pernas L, Adomako-Ankomah Y, Shastri AJ, Ewald SE, Treeck M, Boyle JP, Boothroyd JC. *Toxoplasma* effector MAF1 mediates recruitment of host mitochondria and impacts the host response, *PLoS Biol.* 2014;12. e1001845
- Pernas L, Bean C, Boothroyd JC, Scorrano L. Mitochondria restrict growth of the intracellular parasite *Toxoplasma gondii* by limiting its uptake of fatty acids, *Cell Metab.* 2018;27:886–897.
- Reese ML, Zeiner GM, Saeij JPJ, Boothroyd JC, Boyle JP. Polymorphic family of injected pseudokinases is paramount in *Toxoplasma* virulence. *Proc Natl Acad Sci U S A.* 2011;108: 9625–9630.
- Saeij JPJ, Boyle JP, Coller S, Taylor S, Sibley LD, Brooke-Powell ET, *et al.* Polymorphic secreted kinases are key virulence factors in toxoplasmosis. *Science* 2006;314: 1780–1783.
- Saeij JPJ, Coller S, Boyle JP, Jerome ME, White MW, Boothroyd JC. *Toxoplasma* co-opts host gene expression by injection of a polymorphic kinase homologue. *Nature.* 2007;445: 324–327.
- Scanlon M, Leitch GJ, Visvesvara GS, Shaw AP. Relationship between the host cell mitochondria and the parasitophorous vacuole in cells infected with *Encephalitozoon microsporidia*. *J Eukaryot Microbiol.* 2004;51: 81–87.
- Shapiro RS, Chavez A, Collins JJ. CRISPR-based genomic tools for the manipulation of genetically intractable microorganisms, *Nat Rev Microbiol.* 2018;16: 333–339.
- Sharma SP, Dubey JP. Quantitative survival of *Toxoplasma gondii* tachyzoites and bradyzoites in pepsin and in trypsin solutions. *Am J Vet Res.* 1981;42: 128-130.

- Shen B, Brown KM, Lee TD, Sibley LD. Efficient gene disruption in diverse strains of *Toxoplasma gondii* using CRISPR/CAS9. MBio. 2014;5: 1–11.
- Shwab EK, Zhu XQ, Majumdar D, Pena HFJ, Gennari SM, Dubey JP, et al. Geographical patterns of *Toxoplasma gondii* genetic diversity revealed by multilocus PCR-RFLP genotyping. Parasitology. 2014;141: 453–461.
- Sibley LD, Boothroyd JC. Virulent strains of *Toxoplasma gondii* comprise a single clonal lineage. Nature. 1992;359: 82–85.
- Sidik SM, Huet D, Lourido S. CRISPR-Cas9-based genome-wide screening of *Toxoplasma gondii*. Nat Protoc. 2018;13: 307–323.
- Sigrist CJ, de Castro E, Cerutti L, Cuche BA, Hulo N, Bridge A, et al. New and continuing developments at PROSITE. Nucleic Acids Res. 2013;41: 344–347.
- Sinai AP, Webster P, Joiner KA. Association of host cell endoplasmic reticulum and mitochondria with the *Toxoplasma gondii* parasitophorous vacuole membrane: a high affinity interaction. J Cell Sci. 1997;110: 2117–2128.
- Sinai AP, Joiner KA. The *Toxoplasma gondii* protein ROP2 mediates host organelle association with the parasitophorous vacuole membrane. J Cell Biol. 2001;154: 95–108.
- Stamatakis A. RAxML version 8: A tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics. 2014;30: 1312–1313.
- Su C, Zhang X, Dubey JP. Genotyping of *Toxoplasma gondii* by multilocus PCR-RFLP markers: A high resolution and simple method for identification of parasites. Int J Parasitol. 2006;36: 841–848.
- Su C, Khan A, Zhou P, Majumdar D, Ajzenberg D, Darde M-L, et al. Globally diverse *Toxoplasma gondii* isolates comprise six major clades originating from a small number of distinct ancestral lineages. Proc Natl Acad Sci. 2012;109: 5844–5849.
- Tahara M, Andrabi SBA, Matsubara R, Aonura H, Nagamune K. A host cell membrane microdomain is a critical factor for organelle discharge by *Toxoplasma gondii*. Parasitol Int. 2016;65: 378–388.
- Taniguchi Y, Appiah-Kwarteng C, Murakami M, Fukumoto J, Nagamune K, Matsuo T, et al. Atypical virulence in a type III *Toxoplasma gondii* strain isolated in Japan. Parasitol Int. 2018;67: 587–592.
- Taylor S, Barragan A, Su C, Fux B, Fentress SJ, Tang K, et al. A secreted serine-threonine kinase determines virulence in the eukaryotic pathogen *Toxoplasma gondii*. Science 2006;314: 1776–1780.
- Wiese S, Reidegeld KA, Meyer HE, Warscheid B. Protein labeling by iTRAQ: A new tool for

- quantitative mass spectrometry in proteome research. *Proteomics*. 2007;7: 340–350.
- Yamamoto M, Standley DM, Takashima S, Saiga H, Okuyama M, Kayama H, *et al.* A single polymorphic amino acid on *Toxoplasma gondii* kinase ROP16 determines the direct and strain-specific activation of Stat3. *J Exp Med*. 2009;206: 2747–2760.
- Yamamoto M, Ma JS, Mueller C, Kamiyama N, Saiga H, Kubo E, *et al.* ATF6 $\beta$  is a host cellular target of the *Toxoplasma gondii* virulence factor ROP18. *J Exp Med*. 2011;208: 1533–1546.
- Zakimi S, Kyan H, Oshiro M, Sugimoto C, Xuenan X, Fujisaki K. Genetic characterization of GRA6 genes from *Toxoplasma gondii* from pigs in Okinawa, Japan. *J Vet Med Sci*. 2006;68: 1105–1107.