

Supplementary Data

Title:

Activating glutamate decarboxylase activity by removing the autoinhibitory domain leads to hyper γ -aminobutyric acid (GABA) accumulation in tomato fruit

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>SIGAD3 (Solyc01g005000)

ATGGTTCTCTCAAAAACCTCCTCTGATGATTCTGTACACTCCACATTGCTTCGCTATGTTCGAAGCTT
SIGAD3-F →
CACTACCAAGGTTGAGATGCTAGAGAAGTCTATAACCAAAGAGGCAGCATAACAAATGATTAATGATGA
GTTAATGCTTGATGGGAATCCAAGGTTAAATTGGCATCATTGTAACCACATGGATGAAACCAGAACATG
GATAAGCTTATGATGGCTTCAATTAACAAGAATTATGTTGACATGGATGAATACCCGTCAACCACTGAGC
TTCAGAATCGATGTGTAACATGATAGCGCGTTATTCAATGCGCCTTGAAAGAGGAAGAAATAGGAAT
TGGTGTGGGACAGTGGGTCACTCAGAGGCCATAATGTTAGCAGGGCTGGCCTCAAGAGGAACGGCAA
AACAAACGCAAAGCTGAGGGAAAGCCTATGATAAGCCAACATTGTCACTGGTCTAATGTCAGGTGT
GTTGGGAGAAATTGCAAACACTTTGAAGTGGATTGAAACAAGTCAAGTTAAGTGAAGGGTACTATGT
GATGGACCCAATCAAAGCTGTGAAATGGTAGATGACAACACTATTTGTGTTGCTGCTATTGGTCA
ACACTTAATGGAGAATTGAAAGATGTCAAACACTTGAATGATCTTGATTGAAAAGAATAAACAAACTG
GATGGGACACACCTATTGATGGATGCAGCAAGTGGATTGACCACTTTATCCAGAGTT
GGAATGGGATTAGGCTTCTTAGTGAAGGATATTGAGAACTAAACAAGACTTGCCTCAACAACTCATTGAT
GCTGGTATTGGTTGGTTATTGGAGAACTAAACAAGACTTGCCTCAACAACTCATTGAT
ATCTTGGTGTGATCAGCCTACTTTACTCTCAATTCTAAAGGTTCAAGTCAGTCATTGCTCAATA
TTATCAGCTTATCCGCTTGGCTATGAGGGATATCGAAATGTAATGGAAAATTGTCGTGAAAATGCAATT
GTGCTAAGAAAAGGACTTGAAAAAACAGGACGTTCAATATAATCTCAAAGATGAAGGTATACCCTGG
TGGCATTTCCTCAAAGACAATAGCCTCCACAACGAATTGAGGTCTTGAGACCCCTCCGTAGTTGG
GTGGATTGTCCCAGCCTACACTATGCCAGCTGACCTGCAACATGTTACAGTGTGCGCTGTGATTAGA
GAGGACTTCTCCGAACCCCTAGCAGATCGTCTGCTCTGACATCGTCAAGGTCCACGAGCTCCGA
ATGCCAAAAAGTGGAGGATAATTGATGATCAATAATGAGAAGAAACAGAAATTGAAGTTCAAAGGGC
← **SIGAD3ΔC-R** ←
AATTGCTGAGTTTGGAGAAAGAATATGTTTAGCTAGGAAAGCATCTATTGTTAG ←
← **SIGAD3-R**

Fig. S1

Full-length coding sequence (CDS) of *SIGAD3*. Arrows indicate the position of primers that were used for vector construction of *SIGAD3^{ox}* and *SIGAD3ΔC^{ox}*. The sequence deleted in *SIGAD3ΔC^{ox}* construct is highlighted in yellow.

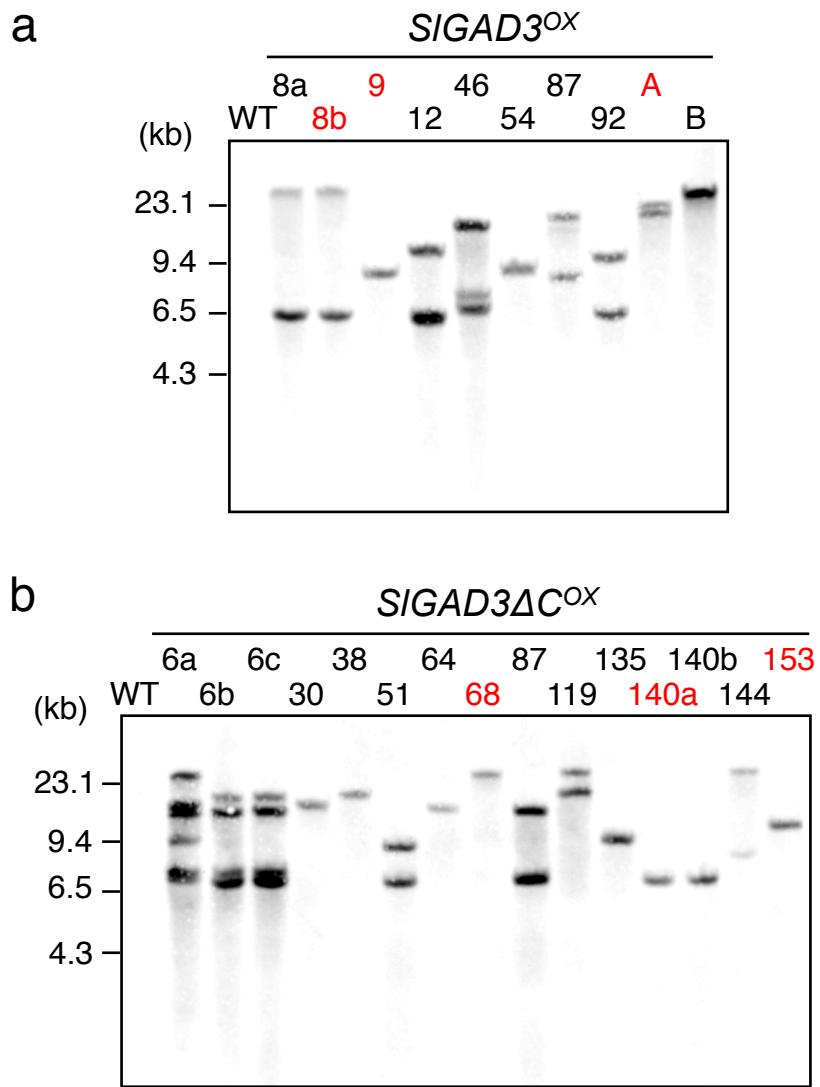


Fig. S2

Southern blot analysis of T_0 transgenic plants. (a) *S/GAD3^{OX}*. (b) *S/GAD3ΔC^{OX}*. Genomic DNA (10 μ g) was digested with *Eco*RI and detected with the *NPTII* probe. WT was also tested as a negative control. The lines used for more detailed analyses are shown in red.

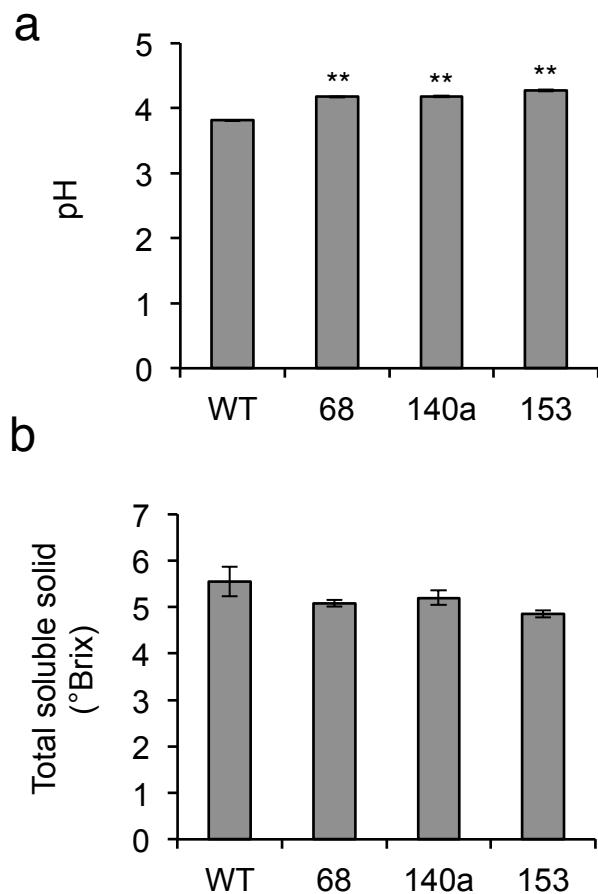


Fig. S3

Effects of *SIGAD3ΔC* over expression on the fruit pH and total soluble solid in T₁ *SIGAD3ΔC^{OX}* lines. pH (a) and total soluble solids (b) were determined using Br+10 fruits of WT and in T₁ *SIGAD3ΔC^{OX}* lines. The mean ± SE of three biological replicates are shown. Asterisks indicate a significant difference between WT and transgenic lines according to Student's *t*-test (**P* < 0.05 and ***P* < 0.01).

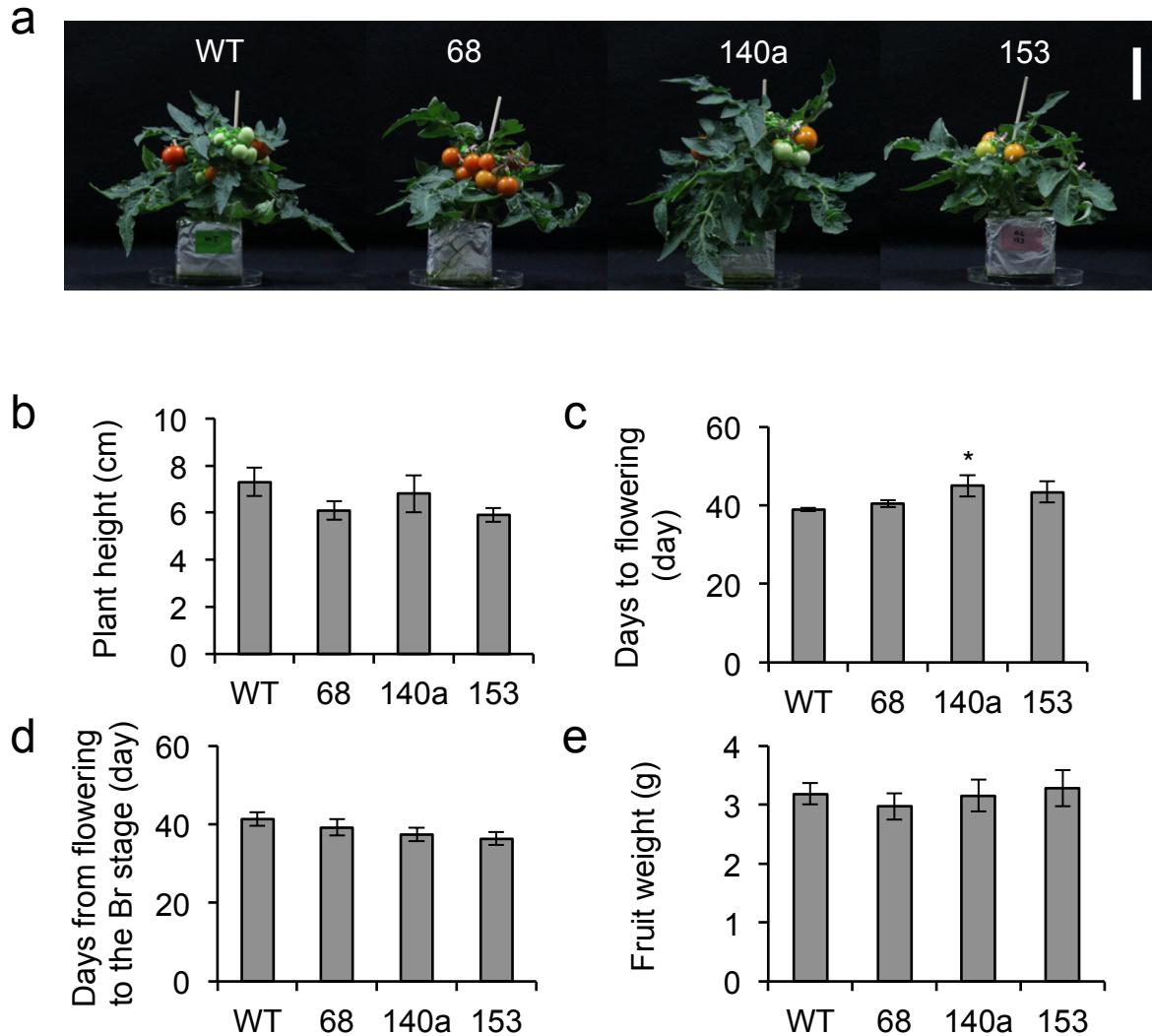


Fig. S4

Plant growth and fruit development of WT and $T_1 S/GAD3\Delta C^{OX}$ lines. (a) Appearance of 3-month-old plants. Bar = 5 cm. (b) Plant height when the first flower opened. Values are the mean \pm SE ($n = 5$). (c) Days to flowering. (d) Days from flowering to the Br stage. (e) Fruit weight. (c–e) Values are the mean \pm SE ($n = 13$). Asterisks indicate a significant difference between WT and transgenic lines according to Student's *t*-test (* $P < 0.05$ and ** $P < 0.01$). Br, breaker

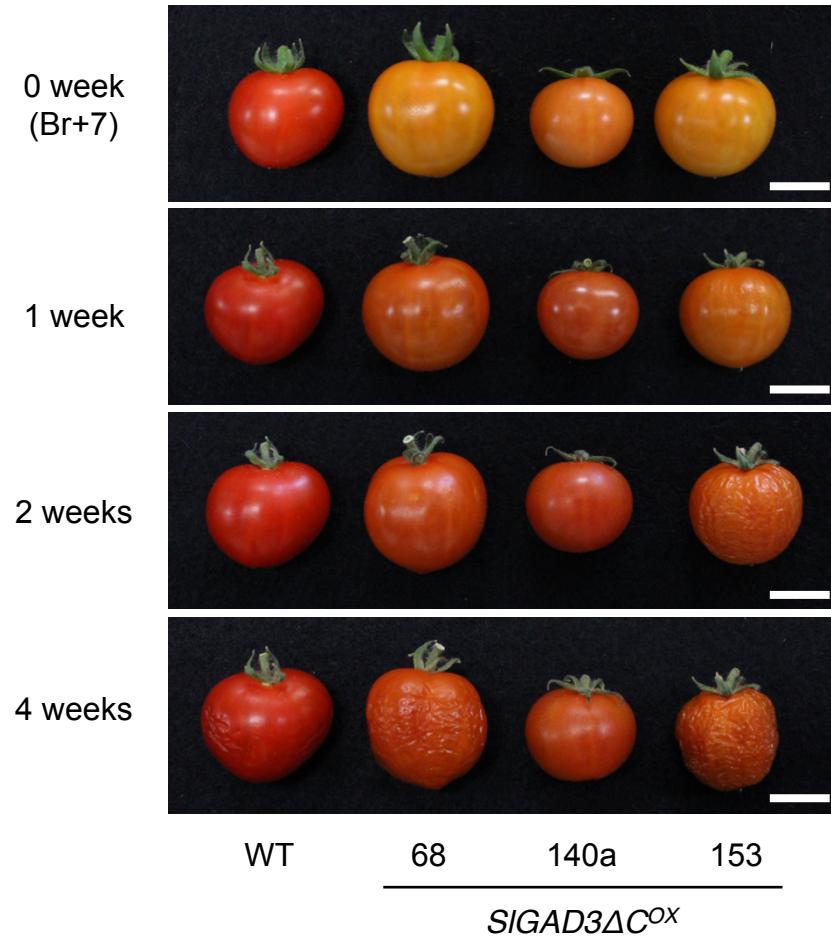


Fig. S5

Fruit shelf life of WT and T₁ *SIGAD3ΔC^{OX}* lines. Fruits were harvested at the Br+7 stage and stored at 25°C for 4 weeks. Bar = 1 cm.

Table S1 List of primers used in this study

Primer name	Primer sequence (5'→3')	Note
SIGAD3-F (SIGAD3ΔC-F)	GCAAAAGAAG<u>GGATCC</u>ATGGTTCTCTCAA AAACTCCTTCTG	Used for vector construction. The last 10 bases of the E8 promoter sequence (Underlined) and the <i>Bam</i> H site (Bold letters) were added.
SIGAD3-R	<u>CTTCATCTCATAT</u> GAGCTC CTAACAAATAG ATGCTTT	Used for vector construction. The first 14 bases of the HSP terminator sequence (Underlined) and the <i>Sac</i> I site (Bold letters) were added.
SIGAD3ΔC-R	<u>CTTCATCTCATAT</u> GAGCTC CTAACATTGATCA TCAAATTATCCTCCAC	Used for vector construction. The first 14 bases of the HSP terminator sequence (Underlined) and the <i>Sac</i> I site (Bold letters) were added.
SIUbiquitin-F SIUbiquitin-R	CACCAAGCCAAAGAAGATCA TCAGCATTAGGGCACTCCTT	Used for qRT-PCR of <i>SIUbiquitin</i> (Takayama et al. 2015)
SIGAD3-F SIGAD3-R	CAGGACGTTCAATATAATC CCTACGGAGGGTCTCAGAG	Used for qRT-PCR of <i>SIGAD3</i> (Takayama et al. 2015)
ACS2-F ACS2-R	GGAGGTTCGTAGGTGTTGAG TAATGGTGAGGGAGGAATAGGT	Used for qRT-PCR of <i>ACS2</i> (Mantelin et al. 2013)
ACS4-F ACS4-R	AACAAGCACAATGGAAGAGGA CGCACTACGAGCAAGGAAT	Used for qRT-PCR of <i>ACS4</i> (Mantelin et al. 2013)
ACO1-F ACO1-R	ACCATGTCTAAGCCCCATT ATTCTGTCCCCGTCTGTTGT	Used for qRT-PCR of <i>ACO1</i> (Shinozaki et al. 2015)
PSY1-F PSY1-R	AACTTGTGATGGCCCAAAC CTGTATGGACAAAGCACCA	Used for qRT-PCR of <i>PSY1</i> (Ariizumi et al. 2014)
PDS-F PDS-R	AGCCGGTGACTACACGAAAC GCTTGCTTCCGACAACATTCT	Used for qRT-PCR of <i>PDS</i> (Ariizumi et al. 2014)
ZDS-F ZDS-R	CATGTCAAAGGCCACTCAGA ACGGTAACAACAGGCACTCC	Used for qRT-PCR of <i>ZDS</i> (Ariizumi et al. 2014)
CRTISO-F CRTISO-R	CCTGGGAATGCCTTCATA AACTCAGCTGCAACACGATG	Used for qRT-PCR of <i>CRTISO</i> (Ariizumi et al. 2014)
LCYB-F LCYB-R	GGACCCCATTGAAAGTTTC AACCATGATGTGGGTTCAGA	Used for qRT-PCR of <i>LCY-B</i> (Guo et al. 2013)
CYCB-F CYCB-R	CTTTCGGACATGGCTAAAC GCTAGATTCCAATCAGTCTAACCA	Used for qRT-PCR of <i>CYC-B</i> (Guo et al. 2013)
RIN-F RIN-R	ATGGCATTGTGGTGAGCAAAG GTTGATGGCTGCATTTCG	Used for qRT-PCR of <i>RIN</i> (Shima et al. 2013)
TAGL1-F TAGL1-R	TGCCTGTAATCTCCTGGAAC AGAATACCTGCTCCATGATTATCAGA	Used for qRT-PCR of <i>TAGL1</i> (Itkin et al. 2009)
FUL1-F FUL1-R	CAACAACTGGACTCTCCTCACCTT TCCTTCCACTCCCCATTATCTATT	Used for qRT-PCR of <i>FUL1</i> (Shima et al. 2013)
FUL2-F FUL2-R	CACACCCCTTAACAACTTCACA GCGATGATCCTCTACTTCTCCAT	Used for qRT-PCR of <i>FUL2</i> (Shima et al. 2013)
SIERF6-F SIERF6-R	CCATGGAAATGCCCATAGTT TCAGTAGAACTGATGAGTTG	Used for qRT-PCR of <i>SIERF6</i> (Lee et al. 2012)