Effects of Platelet-Rich Plasma on Intestinal Anastomotic Healing in Rats: PRP Concentration is a Key Factor

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Title: Effects of platelet-rich plasma on intestinal anastomotic healing in rats: PRP concentration is a key factor

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Running title: PRP has a dose-specific effect

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ABSTRACT

Introduction. There have been few studies that examine the effects of platelet-rich plasma (PRP) on intestinal anastomotic healing. The applied preparation methods and PRP concentrations used in the few studies that have been carried out varied markedly. Therefore, the positive effects of PRP on the anastomotic healing process remain unclear.

The aim of this study is to examine the effects of different concentrations of PRP on intestinal anastomotic healing.

Material and Methods. From SD rat blood, three different concentrations of plasma were prepared: high-concentrated PRP (H-PRP: platelet count $5 \times 10^6$/mm$^3$), low-concentrated PRP (L-PRP: $2 \times 10^6$/mm$^3$), and platelet-poor plasma (PPP). Male SD rats underwent proximal jejunal anastomosis and central venous catheterization. Rats were divided into four groups (N=12 for each group): control, PPP, L-PRP, and H-PRP groups. Two types of PRP and PPP (0.21 ml) were applied to each anastomosis line, with the exception of the control group. Total parenteral nutrition (TPN) solutions were administered (151 kcal/kg/day). Five days after surgery, anastomotic bursting pressure (ABP) in situ and hydroxyproline concentration (HYP) in anastomotic tissue were evaluated.
**Results.** The ABP values of control, PPP, L-PRP, and H-PRP groups were 171±20, 174±23, 189±17, and 148±25 mmHg, respectively. The HYP values of each group were 516±130, 495±123, 629±120, and 407±143 μg/g dry tissue. Compared with the other groups, the L-PRP group exhibited a significant increase in both ABP and HYP, while the H-PRP group exhibited a significant decrease in these two variables. As a result, L-PRP was considered to promote anastomotic wound healing, but H-PRP was considered to inhibit it. There was no significant difference between the PPP group and the control group.

**Conclusions.** PRP concentration plays a crucial role in the efficacy of PRP. PRP might exert positive effects on intestinal anastomotic healing in a dose-dependent manner up to a certain level, but adverse effects occur when it is highly concentrated. The essential PRP action appears to be driven by the platelets themselves.

Key Words: platelet-rich plasma, platelet-poor plasma, gastrointestinal anastomosis, wound healing, PRP concentration, bimodal effect, anastomotic bursting pressure, hydroxyproline, rat.
INTRODUCTION

Despite major advances in surgical techniques and perioperative management, the rates of gastrointestinal anastomotic leakage have still not been reduced to a negligible level. Anastomotic leakage has large negative impacts not only on mortality but also on long-term survival [1][2][3]. Hence, innovative methods to avoid anastomotic leakage are deemed necessary and are much anticipated.

There is a growing body of evidence to demonstrate that platelet concentrates such as platelet-rich plasma (PRP) enhance wound healing in a variety of clinical fields on the basis of the premise that higher growth factor content should promote better healing [4][5][6][7][8]. Platelets contain high quantities of key growth factors, such as platelet-derived growth factor (PDGF) and transforming growth factor-β1 (TGF-β1), which regulate cell proliferation and matrix remodeling [9]. PRP is regarded as a storage vehicle of growth factors. PRP application to gastrointestinal anastomosis is one of the most useful methods to deliver concentrated amounts of growth factors throughout the surgical site, and expected to accelerate the wound healing process. There have been only two reports regarding the effect of PRP application on intestinal wound healing [10][11]. However, these two reports provided contrasting results. Yol et al. showed that PRP has a
positive effect on wound healing in colonic anastomosis [10]. On the other hand, Fresno et al. indicated that PRP application only increases granulation tissue and fibrosis without improving jejunal anastomotic breaking strength [11]. Similarly, the effect of PRP remains a controversial subject in other research fields [12][13]. Behind the controversy is the fact that PRP preparation methods vary between studies, which results in significantly different platelet concentrations. Different platelet concentrations inevitably lead to different concentrations of various growth factors. Consequently, the platelet concentrations might be a crucial factor in determining whether PRP achieves the desired effects on wound healing. In addition, there is the possibility that the optimal concentration of PRP depends on the type of wound to which it is applied and adverse effects can be exerted when too high a concentration is used.

The aim of this study is to clarify the causal relationship between PRP concentration and its effect on intestinal anastomotic healing.

MATERIAL AND METHODS

Animals

Seventy-seven male Sprague-Dawley rats (Nippon Clea, Tokyo, Japan), weighing 192 g
(172-252 g), were used in the experiment. The animals were maintained at 21°C with a 12 h light/dark cycle and allowed free access to water and standard chow for 3 days. The experimental protocol was carried out in a humane manner after receiving approval from the Institutional Animal Experiment Committee of the University of Tsukuba, and in accordance with the Regulations for Animal Experiments of our university and the Fundamental Guidelines for Proper Conduct in Animal Experiments and Related Activities in Academic Research Institutions under the Jurisdiction of the Ministry of Education, Culture, Sports, Science, and Technology.

**Preparation of Platelet-Rich Plasma**

Ten milliliters of homologous fresh blood treated with 1 ml of ACD-A, anticoagulant solution, was obtained in a sterile tube from the heart of each rat. Blood was centrifuged immediately at 400 ×g and 22°C for 10 minutes. Blood was separated into three layers, red blood cells at the bottom, acellular plasma in the supernatant, and a buffy coat layer in between. The upper layer was transferred with a sterile pipette to another 10 ml centrifuge tube without buffy coat and re-centrifuged at 800 ×g and 22°C for 10 minutes. About 0.5 ml of PRP (platelet count about 5.50×10⁸/mm³) was pipetted from the bottom of the tube and about 1.0 ml of platelet-poor plasma (PPP) was harvested as a supernatant (Fig. 1A).
The prepared PRP, PPP, and whole blood of the rats were subjected to platelet and white blood cell counting with a cell counter (MICROS abc LC-152, HORIBA, ltd., Kyoto, Japan), and PPP was added a little at a time to adjust PRP to different platelet concentrations: low-concentrated PRP (platelet count $2 \times 10^6$/mm$^3$) and high-concentrated PRP (platelet count $5 \times 10^6$/mm$^3$). To produce a viscous coagulum gel that can be applied to the anastomosis, 180 $\mu$l of PRP was mixed with the “activator”, a mixture of 180 units of bovine thrombin (Sigma-Aldrich, St. Louis, MO, USA) and 30 $\mu$l of 10% calcium chloride. Calcium chloride neutralizes the anticoagulant effect of citrate, and bovine thrombin initiates the clotting process. Likewise, PPP was activated by the above-mentioned process.

Finally, each of activated PRP and PPP was prepared as a total amount of 0.21 ml. The mixture, which clots within a few seconds, should be used immediately to allow the release of growth factors.

**Determination of PDGF-BB and TGF-β1 concentrations**

The activated PRP and PPP aliquots were incubated for 30 min at 37°C in a 5% CO$^2$ incubator (Auto-Flow incubator, NuAire, Inc., Plymouth, MN, USA). Samples were centrifuged for 10 min at 15500 $\times$g, and the supernatants were harvested. All supernatants were immediately stored at −80°C until further analysis. After being thawed,
the samples were assayed for platelet-derived growth factor-BB (PDGF-BB) and transforming growth factor-β1 (TGF-β1) by enzyme-linked immunosorbent assay (ELISA) testing (R&D Systems, Minneapolis, MN, USA). The levels of PDGF BB and TGF-β1 are reported in picograms per milliliter.

5 Operative procedure

Prior to surgery, all rats were fasted overnight. They were anesthetized by intraperitoneal injection of pentobarbital sodium (Somnopentyl®, Kyoritsu Seiyaku, Tokyo, Japan) at a dose of 40 mg/kg of body weight. Under aseptic conditions, (1) jugular vein catheterization at a central venous position was carried out. The tip of the catheter was a silicone tube of 0.5 mm inner diameter and 1.0 mm outer diameter (Fuji Systems Co., Tokyo, Japan), and the other part of the catheter was a plastic tube of 0.5 mm inner diameter and 1.0 mm outer diameter (Imamura Co., Chiba, Japan). The distal end of the catheter was tunneled subcutaneously and brought out in the cephalad portion of the interscapular area. The catheters were fixed to the skin using a harness attached to a swivel assembly. (2) A laparotomy was performed through an upper midline incision measuring 3 cm. The proximal jejunum was divided 2.0 cm distal from the duodenum, and single-layer, end-to-end anastomosis was performed with 7-0 Proline sutures
(Ethicon Inc., USA) in an inverted interrupted fashion. To ensure technical uniformity, all anastomoses were sutured by only one surgeon. After that, the animals were divided into four groups: the control group (n=12), in which neither PPP or PRP was applied to the anastomosis line; the PPP group (n=12), in which activated PPP was applied to the anastomosis line; the low-concentrated PRP (L-PRP) group (n=12), in which activated low-concentrated PRP was applied to the anastomosis line; and the high-concentrated PRP (H-PRP) group (n=12), in which activated high-concentrated PRP was applied to the anastomosis line. In the PPP group, the L-PRP group, and the H-PRP group, the activated PRP or PPP was applied as a film layer of around 8 mm width and 3 mm thickness immediately after mixing with bovine thrombin and 10% calcium chloride (Fig. 1B). The fascial layer of the abdominal wall was closed with a continuous suture using 2-0 Silk (Ethicon Inc., USA), and the abdominal skin was closed in the same way. Following these procedures, the rats were maintained in individual metabolic cages for 5 days.

Nutrient solutions

In all groups, nutrient solutions (Neoparen 2, Otsuka Pharmaceutical Co., Ltd., Tokyo, Japan) were administered via the central venous catheter. Following surgical procedures, rats were maintained in individual metabolic cages for 5 days. Identical solutions were
administered to the four groups. Continuous infusion using infusion pumps (SP-115, JMS Co., Ltd., Tokyo, Japan) was initiated immediately after surgery. The full-dose energy intake was 151 kcal/kg body weight per day, equal to an infusion rate of 9 ml/kg body weight per hour. This target value corresponds to about 21 kcal/kg body weight per day for humans [14], which is thought to be reasonable during the acute phase of surgical stress [15]. Twenty-four hours postoperatively, half of the target calories had been administered. The reason why the rats were fed with TPN instead of standard rat chow and water ad libitum was to provide equivalent amounts of fluid volume and energy intake, which allows for leveling of the nutritional backgrounds of all rats.

Nutritional parameters

Urinary output was monitored daily, and body weights were recorded. On postoperative day 5, the rats were deeply anesthetized by an intraperitoneal injection of pentobarbital sodium at a dose of 40 mg/kg of body weight, and the infusion of nutrients was stopped. Blood was drawn from the inferior vena cava to determine the serum levels of total protein, albumin, glucose, and blood urea nitrogen.

Anastomotic bursting pressure

After the animals were anesthetized, the abdomen was opened, and the anastomotic
bursting pressure (ABP) was measured in situ without interruption of the normal mesenteric blood supply or adhesions to the anastomosis. A 16-gauge silicone rubber catheter was inserted 1.5 cm from the oral side of the anastomosis to the distal side. Two ligations were made, 1.5 cm proximal and distal from the anastomosis with 3-0 silk sutures. Normal saline solution was continuously infused through the catheter via a syringe pump at a rate of 1.0 ml/min. Intraluminal pressure was monitored continuously via a transducer (BLPR®, World Precision Instruments Inc., Sarasota, FL, USA) and recorded on a chart recorder (Power Lab®, AD Instruments, Tokyo, Japan). The ABP was defined as the peak pressure attained just before rupture of the anastomosis.

Collagen concentration

Hydroxyproline is useful as an index of collagen concentration because it is an amino acid unique to collagenous proteins [16]. After bursting of the anastomosis, 1 cm of the anastomotic segment, that is, 0.5 cm proximal and distal from the anastomosis, was excised and dissected free of mesenteric fat, intestine, and adherent omentum. Adhesions to the surrounding tissue were not dense, and it was feasible to separate adhesions from the anastomotic site using the blunt technique. The specimens were opened at the mesenteric side, gently washed with saline solution, and cut into three pieces. A strip of
the specimen was used to determine the concentration of hydroxyproline as an index of collagen concentration. The concentration of hydroxyproline was measured as described by Reddy and Enwemeka [17]. The procedure is based on alkaline hydrolysis of the tissue homogenate and subsequent determination of the free hydroxyproline in hydrolysate. Chloramine-T was used to oxidize the free hydroxyproline for production of a pyrrole. The addition of Ehrlich reagent resulted in the formation of a chromophore that can be measured at 550 nm. Calculations were made to express the result as micrograms of hydroxyproline per gram of dry tissue (μg/g dry tissue).

**Histopathological Analysis**

A strip of the anastomosis was immediately fixed in 10% phosphate-buffered formaldehyde, and then embedded in paraffin. One sample was randomly selected from each of the four groups. Histopathological sections (4 μm) were stained with hematoxylin-eosin (HE) and Masson trichrome (MT). The degree of collagen deposition was descriptively assessed in a blinded manner by two pathologists.

**Statistics**

All data are presented as the mean value ± standard deviation (SD). Comparisons among the four groups were tested by non-repeated measures ANOVA. Multiple comparisons
were examined by Bonferroni test, and differences were considered statistically significant at p < 0.05.

RESULTS

5 Number of platelets and quantification of growth factor levels for three different concentrations of platelets

Blood samples obtained from SD male rats exhibited an original platelet count of 0.91 ± 0.12 × 10⁶/mm³ and an original white blood cell count of 6.45 ± 1.53 × 10³/mm³. PPP (n=5), L-PRP (n=5), and H-PRP (n=5) exhibited average platelet concentrations of 0.03 ± 0.01, 2.11 ± 0.18, and 5.07 ± 0.19 × 10⁶/mm³, respectively. These platelet concentrations represented 0.03-, 2.33-, and 5.60-fold increases compared with the original platelet count.

None of PPP, L-PRP, and H-PRP included white blood cells.

For PDGF-BB, 3439 ± 2817, 18462 ± 4671, and 49365 ± 4299 pg/ml were respectively determined in PPP, L-PRP, and H-PRP (Fig. 2). For TGF-β1, 5824 ± 1647, 101008 ± 24584, and 210453 ± 37188 pg/ml were respectively determined in PPP, L-PRP, and H-PRP (Fig. 3). Both PDGF-BB and TGF-β1 values of H-PRP and L-PRP were significantly higher than those of PPP (p<0.01). Both PDGF-BB and TGF-β1 values of
H-PRP were significantly higher than those of L-PRP (p<0.01).

**Numbers of platelets in three different concentrations of platelets for surgical procedure**

Blood samples obtained from SD male rats exhibited an original platelet count of 0.93 ± 0.12 × 10^6/mm³ and an original white blood cell count of 6.69 ± 2.34 × 10^3/mm³. PPP (n=12), L-PRP (n=12), and H-PRP (n=12) exhibited average platelet concentrations of 0.06 ± 0.02, 1.93 ± 0.13, and 5.15 ± 0.30 × 10^6/mm³, respectively. These platelet concentrations were equivalent to 0.06-, 2.08-, and 5.53-fold increases compared with the original platelet count. None of PPP, L-PRP, and H-PRP included white blood cells.

**Nutritional parameters**

The initial body weight and body weight change 5 days after operation were similar in the four groups (Table 1). Total infusion volume, total energy intake, and total urine volume did not differ between the four groups (Table 1). There were no significant differences between the four groups in serum levels of total protein, albumin, glucose, or blood urea nitrogen (Table 2). These results indicated that there was no crucial factor directly impacting on intestinal anastomotic healing other than the effects of PRP.

**Anastomotic bursting pressure (ABP)**
There was no intraperitoneal abscess formation, and no anastomotic leakage was noted in any animal. All tested anastomoses burst exactly at the suture line. The ABP values of the control group, the PPP group, the L-PRP group, and the H-PRP group were 171 ± 20, 174 ± 23, 189 ± 17, and 148 ± 25 mmHg, respectively. In the L-PRP group, the ABP values were significantly higher compared with those in the control group, the PPP group, and the H-PRP group (p<0.05). On the other hand, the ABP values of the H-PRP group were significantly lower than those of the other three groups (p<0.05) (Fig. 4). No significant difference was detected between the PPP group and the control group.

**Collagen concentration in jejunal anastomotic tissue**

As a surrogate marker of collagen, the hydroxyproline concentrations of the control group, the PPP group, the L-PRP group, and the H-PRP group were 515 ± 130, 495 ± 123, 629 ± 120, and 407 ± 143 μg/g dry tissue, respectively. In the L-PRP group, the hydroxyproline concentrations were significantly higher than those in the control group, the PPP group, and the H-PRP group (p<0.05). The hydroxyproline concentrations of the H-PRP group were significantly lower than those of the other three groups (p<0.05) (Fig. 5). There was no significant difference between the PPP group and the control group.

**Histopathological examination in jejunal anastomotic tissue (Fig. 6)**
In the L-PRP group, collagen deposition mainly in the serosal layer was most pronounced, a clear sign of increased collagen production, compared with those of the control group, the PPP group, and the H-PRP group. In contrast, the H-PRP group exhibited the smallest collagen deposition among the four groups. There was little difference between the control group and the PPP group.

DISCUSSION

Gastrointestinal wound healing is characterized by three phases of healing, namely, the inflammation phase, the proliferation phase, and the maturation phase. At the early phase, platelets play a key role in the wound healing process. After the formation of fibrin clots that serve as a scaffolding for platelets, platelets act during the first 72 hours of the inflammation phase. The initial and brief release of growth factors from platelets mediates the entire process by controlling growth, differentiation, and cell metabolism [18]. After that, macrophages take over the work of platelets, and initiate both wound debridement and the production of growth factors that modulate fibroplasia and angiogenesis. Because the anastomotic mechanical strength reaches the lowest value 3 or 4 days after gastrointestinal anastomosis, the anastomotic leakage most commonly occurs
during this period [19]. From the above-mentioned viewpoint, the purpose of PRP application to gastrointestinal anastomosis is to accelerate the activation of fibroblasts and collagen formation by multiple viable growth factors released from platelets, which enables an increase in anastomotic strength at the earliest stage possible during the inflammation phase.

In this study, PRP was prepared after two-step centrifugation of blood obtained from rats without any commercial kits. The advantage of this method is that both PRP and platelet-poor plasma (PPP) are easily available at the time, and the desired PRP concentration is fully adjustable by the addition of PPP. The concentration of each growth factor released from PRP is predictable on the basis of PRP concentrations. There is a possibility that concentrations of growth factors in PRPs differ according to species, such as rat, goat, and human [20]. Therefore, we quantified the numbers of platelets and two representative growth factors (PDGF-BB, TGF-β1) in three different concentrations of platelets (PPP, L-PRP, H-PRP) obtained from rat, and verified the relationship between numbers of platelets and concentrations of growth factors (Fig 2 and Fig 3). In our method, PRP excludes buffy coat completely. The influence of buffy coat including leukocytes is still unclear [21]. Some authors recommend the elimination of leukocytes
[22], while several studies have shown that leukocytes in buffy coat play a part in the effects of PRP [23]. In our experiment, the buffy coat layer was eliminated because the aim of this study is to investigate effects of PRP simply depending on different platelet concentrations. By adding thrombin to PRP as an activator, PRP works via the degranulation of the α-granules, which leads to the release of the following 7 fundamental growth factors: the three isomers of platelet-derived growth factor (PDGF-AA, PDGF-BB, and PDGF-AB), two forms of transforming growth factor-β (TGF-β1 and TGF-β2), vascular endothelial growth factor (VEGF), and epidermal growth factor [24]. After this initial burst of PRP-related growth factors, platelets synthesize and secrete additional growth factors for the remainder of their life-span [10][11]. The addition of calcium chloride promotes the physiological clotting process, which leads to sound attachment of platelets to the anastomotic site and a sustained higher level of delivery of viable growth factors. Besides, clots also act as a suitable scaffold, where fibroblasts and endothelial cells can receive stimuli from several growth factors and proliferate. These growth factors are biologically active polypeptides, and most of them have key roles in every step of wound healing. Some reports demonstrate that the local application of PDGF-BB and/or TGF-β1 as a single agent enhances wound
healing [9][25][26]. Moreover, it was reported that the local injection of VEGF into the muscularis propria accelerates colonic anastomotic healing and strengthens the anastomosis by increased angiogenesis [27]. The specific characteristic of PRP is that it acts as a combination of multiple growth factors, each of which exerts a unique influence on the complex cascade of the wound healing process, and therefore should generate a synergistic effect unlike single recombinant growth factors that focus on a single function.

As just described, the biological rationale for PRP application to various wounds seems to be theoretically sound, but its effects on intestinal anastomotic wound healing remain controversial [10][11]. To address this issue, we focused on the assumption that PRP has a dose-specific effect on intestinal anastomoses, and thus PRP concentration affects the outcome.

In our study, the ABP values of proximal jejunal anastomoses in the L-PRP group, of which the PRP concentration corresponds to about a 2-fold increase over the original platelet concentration, were significantly higher than those of the other three groups (increased by 11% compared with that of the control group). There was no significant difference in the ABP values between the PPP group and the control group. PPP contains serum proteins such as fibrin, fibronectin, and vitronectin, known to act as cell adhesion
molecules, but very few platelets [28]. These findings might provide an explanation for the underlying mechanism of PRP action in intestinal anastomotic healing. The essential PRP action appears to be driven by platelets themselves, but not some serum proteins. On the other hand, the ABP values of the H-PRP group, of which the PRP concentration corresponds to about a 5-fold increase over the original platelet concentration, were significantly lower than those of the other three groups (decreased by 13.5% compared with that of the control group). The ABP is a more reliable measure for evaluating early postoperative anastomotic mechanical strength, especially within a week of the operation [16][29]. Generally, the bursting pressure is considered to reflect the physiological strain in the intestine more accurately than the breaking strength [16]. In addition, the bursting pressure shows indirect collagen formation and reflects the balance between collagen deposition and lysis [30]. Therefore, the ABP might be regarded as not only an integrated measure of anastomotic wound healing but also the absolute outcome of gastrointestinal anastomoses. Considering the physiological aspects of the ABP, we could conclude that L-PRP application significantly promoted jejunal anastomotic healing, but H-PRP application definitely impaired the wound healing process. Needless to say, the data on collagen concentration at the anastomosis coincided with the ABP results and reinforced
them. Furthermore, self-explanatory histopathological findings supported the above-mentioned conclusion. In the L-PRP group, there was clear evidence of increased fibroblast density and collagen formation primarily in the serosal layer. In this experiment, the proximal jejunum was divided, and then single-layer, end-to-end anastomosis was performed in an inverted interrupted fashion. This is also known as serosa-to-serosa anastomosis. The essence of this anastomotic technique is full-thickness intestinal anastomosis with serosal apposition, which places importance on union of the wound, especially in the serosal layer. In fact, the application of L-PRP increased the anastomotic mechanical strength by accelerating collagen formation in the serosal layer, which should be central to wound healing. To the best of our knowledge, this paper is the first report demonstrating that PRP concentration plays a crucial role in the effect of PRP on intestinal anastomotic healing. Although PRP might exert positive effects in a dose-dependent manner up to a certain level, excessive PRP concentration results in serious adverse effects rather than no effect. Our results correlated with the findings of several previous studies [24][31]. By evaluating the effect of different concentrations of PRP on osteoblast and fibroblast functions in vitro, Graziani et al. reported that maximal PRP concentrations used in their study, which varied between 4.2- and 5.5-fold increases
over the original platelet concentration, might not provide the optimal environment for the promotion of wound healing. Optimal results were observed at a platelet concentration of 2.5-fold, which was approximately half of the maximum concentration that could be obtained [24]. It has remained unclear how PRP highly concentrated above a certain level interferes with the normal wound healing process. However, one-dimensional thinking of “if some is good, more is better” is not recommended for PRP application to intestinal anastomoses. To achieve the desired effect of PRP on intestinal wound healing, the use of PRP at the optimal density is thought to be imperative. The present results provide a possible explanation for the existing criticism of the efficacy of PRP. Future studies should be designed with the understanding that the PRP concentration plays a crucial role in the efficacy of PRP.

Some limitations of our study need to be addressed. First, a potential weakness of our experiment is the use of homologous PRP, but not autologous PRP. Originally, the term PRP refers to an autologous concentration of platelets in a small volume of plasma. In small animal models such as rats, the blood volume is too small to produce autologous PRP. Inevitably, donor blood is commonly used. It has been pointed out that the use of donor animal blood platelets conveys a risk of imparting an overt immune reaction,
which could lead to false-negative results [31]. However, our study could remove such doubts because of the demonstration of positive effects of PRP in the L-PRP group. Second, we just investigated the overall biological properties of PRP but have yet to evaluate all sorts of growth factors included in PRP individually in this study. In the next stage, we need to identify the factor or factors that actually improve the anastomotic strength and healing.

Finally, we propose the following vision for clinical practice. PRP can be easily prepared from only 20-40 cc of autologous blood of the patient. After obtaining autologous blood of the patient, the whole process takes no more than one hour. When centrifugal machines are present in the operating room, we can proceed with the preparation of PRP depending on the progress of the operation and accomplish the whole process inside the operating room. Needless to say, quality control mechanisms are always essential. First, PRP applied to the wound should be prepared at the optimal concentration that allows for maximum enhancement of wound healing. Second, “fresh PRP” needs to be used. “Fresh PRP” implies that each platelet is alive and active in PRP. Otherwise, PRP cannot fully exert its positive effect on wound healing because platelets are expected to synthesize and secrete additional growth factors for the remainder of their
lifespan after the initial burst of PRP-related growth factors. In concrete terms, we think that the clinical application of PRP can be targeted at patients who undergo gastrointestinal anastomoses under unfavorable conditions for wound healing. In such cases, leakage of intestinal anastomosis can occasionally occur, even though gastrointestinal anastomosis is performed correctly and complies with the tension-free construction of the suture, with adequate tissue perfusion. There should be a factor that delays or impairs wound healing, but it is difficult to determine the specific cause of anastomotic leak because the process of wound healing is very complex and depends on many factors. Not surprisingly, specific individual countermeasures cannot be taken because the specific cause cannot be identified. Therefore, we focus on PRP working as a storage vehicle of growth factors. PRP is expected to coordinate the process of wound healing and handle various dangerous situations prone to the occurrence of anastomotic leak. PRP might achieve the desired effect to accelerate wound healing and ensure adequate anastomotic mechanical strength earlier than normal, which would lead to the prevention of anastomotic leak under adverse conditions. In the case of patients with cancer, the risks of using PRP should be taken into full account. PRP application to an anastomotic site is essentially a local therapy, not a systemic therapy (e.g., blood
transfusion). In addition, the actual number of platelets administered as the modality of PRP is extremely low compared with the total number of platelets given as the modality of platelet transfusion. Therefore, it is unlikely that PRP application to an anastomotic site stimulates remnant cancer cells in other remote regions. Generally, there seems to be little likelihood that PRP application worsens clinical outcome after cancer operation.

However, there are some alarming pathological conditions. In particular, in the case of subclinical peritoneal metastasis, PRP application might pose a risk of local recurrence in the area surrounding the anastomotic site by stimulating cancer cells with a variety of growth factors derived from PRP. In such cases, PRP application might hasten cancer recurrence as well.

**CONCLUSION**

PRP might exert positive effects on intestinal anastomotic healing in a dose-dependent manner up to a certain level, but adverse effects occur at a high concentration. The essential PRP action might be driven by the platelets themselves, but not by some serum proteins. Further investigations are needed to clarify the optimal PRP concentration that allows for maximum enhancement of anastomotic wound healing.
REFERENCES


Figure legends

Fig 1. Actual preparation of PRP and application of PRP to the jejunal anastomotic site.

A: The arrow indicates PRP and the arrowhead indicates platelet-poor plasma. B: The arrow indicates activated PRP covering the anastomotic site.

Fig 2. The determination of PDGF-BB concentrations in three different concentrations of platelets.

PPP: platelet-poor plasma, L-PRP: low-concentrated platelet-rich plasma, H-PRP: high-concentrated platelet-rich plasma. Each column represents the mean value ± standard deviation of PDGF-BB in each concentration of platelets; *: $p<0.01$ versus the PPP, †: $p<0.01$ versus the L-PRP (non-repeated measures ANOVA and Bonferroni’s multiple t-test).
**Fig 3.** The determination of TGF-β1 concentrations in three different concentrations of platelets.

PPP: platelet-poor plasma, L-PRP: low-concentrated platelet-rich plasma, H-PRP: high-concentrated platelet-rich plasma. Each column represents the mean value ± standard deviation in each concentration of platelets; *: $p<0.01$ versus the PPP, †: $p<0.01$ versus the L-PRP (non-repeated measures ANOVA and Bonferroni’s multiple t-test).

**Fig 4.** Anastomotic bursting pressure 5 days postoperatively in the four groups.

Control: without application of PPP or PRP to the anastomosis line; PPP: with application of PPP to the anastomosis line; L-PRP: with application of low-concentrated PRP to the anastomosis line; H-PRP: with application of high-concentrated PRP to the anastomosis line. Each column represents the mean value ± standard deviation of rats in
the four groups; *: \( p < 0.05 \) versus the control, PPP, and H-PRP groups, †: \( p < 0.05 \) versus the control, PPP, and L-PRP groups (non-repeated measures ANOVA and Bonferroni’s multiple t-test).

**Fig 5.** The concentration of hydroxyproline in jejunal anastomotic tissue 5 days postoperatively in the four groups.

Control: without application of PPP or PRP to the anastomosis line; PPP: with application of PPP to the anastomosis line; L-PRP: with application of low-concentrated PRP to the anastomosis line; H-PRP: with application of high-concentrated PRP to the anastomosis line. Each column represents the mean value ± standard deviation of rats in the four groups; *: \( p < 0.05 \) versus the control, PPP, and H-PRP groups, †: \( p < 0.05 \) versus the control, PPP, and L-PRP groups (non-repeated measures ANOVA and Bonferroni’s
multiple t-test).

**Fig 6.** Typical histopathological appearance of the jejunal anastomosis 5 days postoperatively in the four groups.

5  **A1-A3**: Control without application of PPP or PRP to the anastomosis line;  **B1-B3**: PPP with application of PPP to the anastomosis line;  **C1-C3**: L-PRP with application of low-concentrated PRP to the anastomosis line;  **D1-D3**: H-PRP with application of high-concentrated PRP to the anastomosis line.  **A1-D1**: 40-fold magnified hematoxylin and eosin stain (HE 40×);  **A2-D2**: 40-fold magnified Masson trichrome stain (MT 40×);  **A3-D3** 100-fold magnified Masson trichrome stain (MT 100×).
Table 1. Nutritional parameters in the four groups.

<table>
<thead>
<tr>
<th></th>
<th>Control</th>
<th>PPP</th>
<th>L-PRP</th>
<th>H-PRP</th>
</tr>
</thead>
<tbody>
<tr>
<td>Initial body weight (g)</td>
<td>195 ± 21</td>
<td>209 ± 19</td>
<td>199 ± 16</td>
<td>207 ± 15</td>
</tr>
<tr>
<td>Body weight change (%)</td>
<td>1.7 ± 2.3</td>
<td>3.0 ± 1.3</td>
<td>2.0 ± 2.1</td>
<td>2.8 ± 3.4</td>
</tr>
<tr>
<td>Infusion volume (ml/5 days)</td>
<td>190 ± 23</td>
<td>196 ± 21</td>
<td>191 ± 14</td>
<td>197 ± 14</td>
</tr>
<tr>
<td>Energy intake (kcal/5 days)</td>
<td>155 ± 19</td>
<td>161 ± 17</td>
<td>157 ± 11</td>
<td>162 ± 12</td>
</tr>
<tr>
<td>Urine volume (ml/g/5 days)</td>
<td>0.53 ± 0.06</td>
<td>0.54 ± 0.05</td>
<td>0.59 ± 0.05</td>
<td>0.50 ± 0.07</td>
</tr>
</tbody>
</table>

Control: without application of PPP or PRP to the anastomosis line; PPP: with application of PPP to the anastomosis line; L-PRP: with application of low-concentrated PRP to the anastomosis line; H-PRP: with application of high-concentrated PRP to the anastomosis line. All values represent the mean value ± standard deviation of rats in each of the four groups. Total infusion volume, total energy intake, and total urine volume did not differ between the four groups (non-repeated measures ANOVA). N.S.: no significance.
**Table 2.** Biochemical nutritional parameters in the four groups.

<table>
<thead>
<tr>
<th></th>
<th>Control</th>
<th>PPP</th>
<th>L-PRP</th>
<th>H-PRP</th>
<th>N.S</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total protein (g/dl)</td>
<td>4.6 ± 0.5</td>
<td>4.5 ± 0.3</td>
<td>4.7 ± 0.3</td>
<td>4.8 ± 0.3</td>
<td>N.S</td>
</tr>
<tr>
<td>Serum albumin (g/dl)</td>
<td>2.5 ± 0.5</td>
<td>2.6 ± 0.3</td>
<td>2.5 ± 0.3</td>
<td>2.6 ± 0.3</td>
<td>N.S</td>
</tr>
<tr>
<td>Glucose (mg/dl)</td>
<td>183 ± 102</td>
<td>229 ± 221</td>
<td>175 ± 68</td>
<td>153 ± 41</td>
<td>N.S</td>
</tr>
<tr>
<td>Blood urea nitrogen (mg/dl)</td>
<td>12.0 ± 2.6</td>
<td>10.5 ± 5.3</td>
<td>13.3 ± 4.8</td>
<td>12.4 ± 2.7</td>
<td>N.S</td>
</tr>
</tbody>
</table>

Control: without application of PPP or PRP to the anastomosis line; PPP: with application of PPP to the anastomosis line; L-PRP: with application of low-concentrated PRP to the anastomosis line; H-PRP: with application of high-concentrated PRP to the anastomosis line. All values represent the mean value ± standard deviation of rats in each of the four groups. There were no significant differences in serum levels of total protein, albumin, glucose, and blood urea nitrogen between the four groups (non-repeated measures ANOVA). N.S.: no significance.
Fig. 2 The determination of PDGF-BB.

- PPP
- L-PRP
- H-PRP

** p<0.01
†† p<0.01

PDGF-BB [pg/ml]
Fig. 3 The determination of TGF-β1.
Fig. 4 Anastomotic bursting pressure in the four groups

Anastomotic Bursting Pressure [mmHg]

- Control
- PPP
- L-PRP
- H-PRP

* p<0.05
† p<0.05
Fig. 5 Concentration of hydroxyproline in jejunal anastomotic tissue in the four groups

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Hydroxyproline concentration [μg / g dry tissue]</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>300</td>
</tr>
<tr>
<td>PPP</td>
<td>300</td>
</tr>
<tr>
<td>L-PRP</td>
<td>900</td>
</tr>
<tr>
<td>H-PRP</td>
<td>600</td>
</tr>
</tbody>
</table>

* * p<0.05
† † p<0.05
Fig. 6 Histological finding of anastomosis